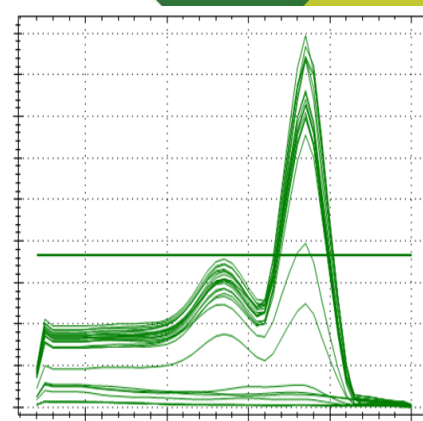


# Experiences and challenges of *Fusarium oxysporum* f. sp. *cubense* Tropical Race 4 diagnostic in banana crops in Colombia



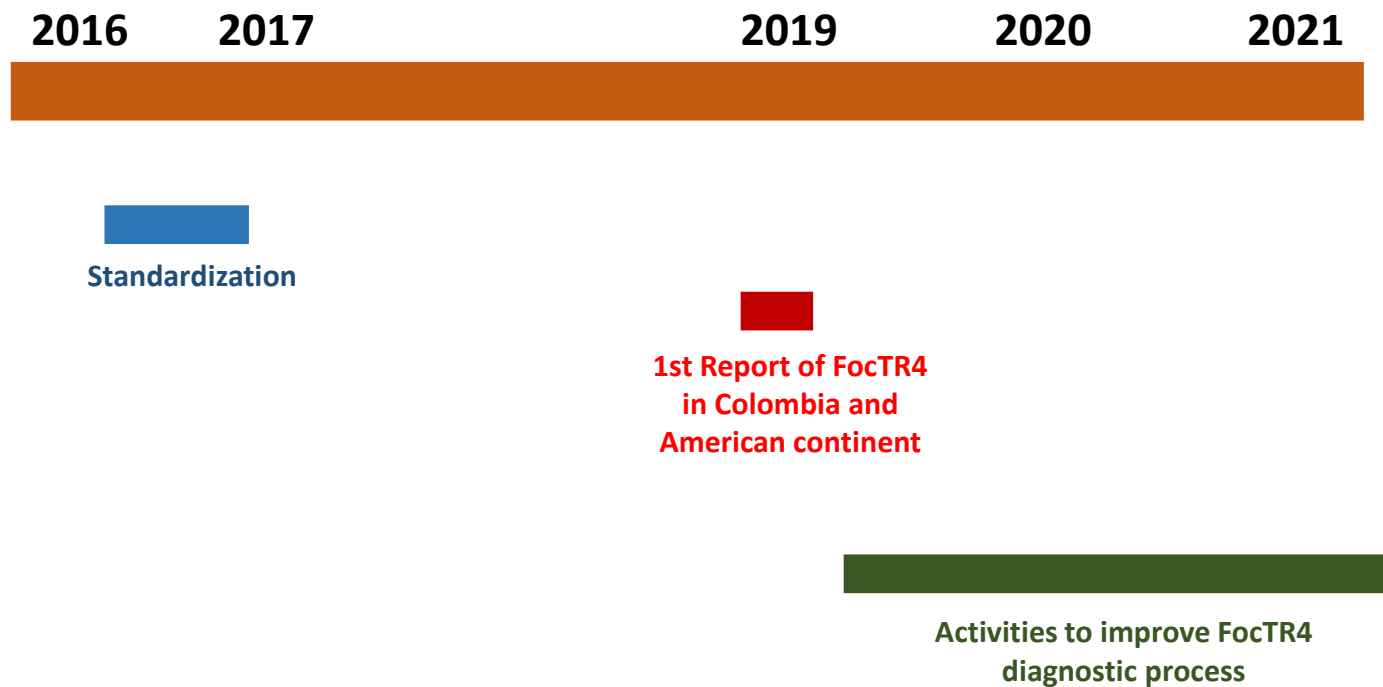
Dirección Técnica de Análisis y Diagnóstico Agrícola

Subgerencia de Análisis y Diagnóstico

**Instituto Colombiano Agropecuario**

By Mariluz Ayala Vásquez

# Background and preliminary process of *FocTR4* detection



# 2016-2017: Memories

Early activities  
for *FocTR4*  
detection in  
Colombia

**2016: Training (Banana Research Group-  
Wageningen University)**

**2016-2017: Development and standardization of  
*FocTR4* detection method in ICA's laboratories**



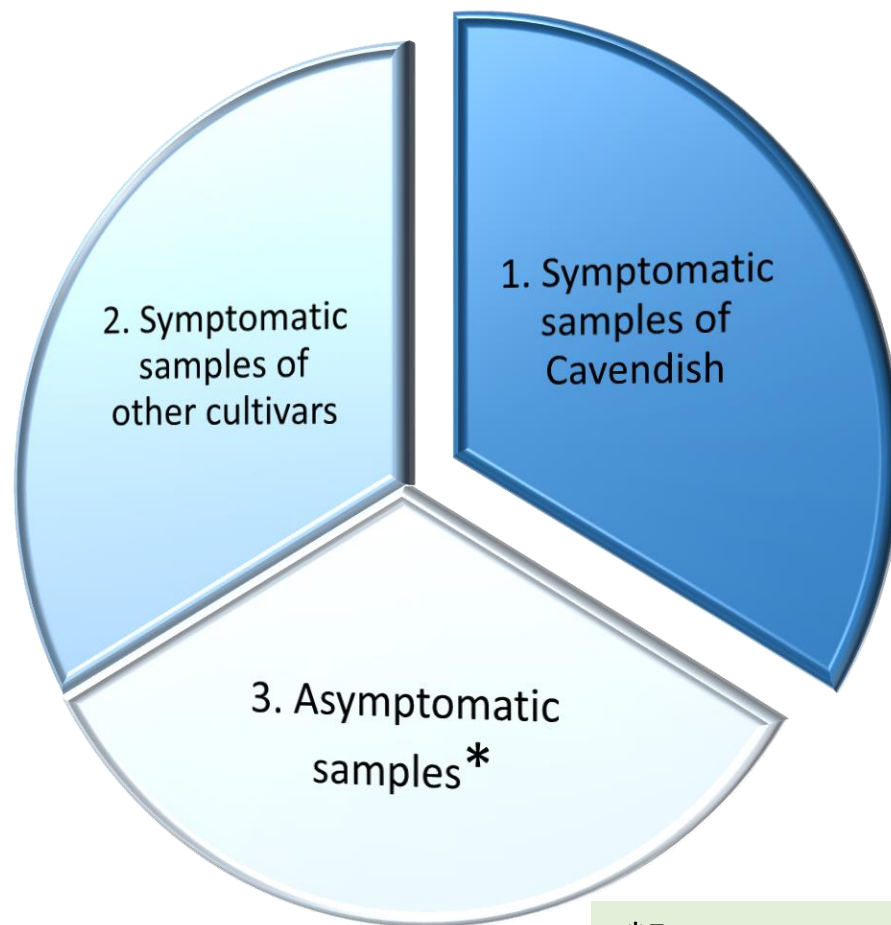
**WAGENINGEN**  
UNIVERSITY & RESEARCH



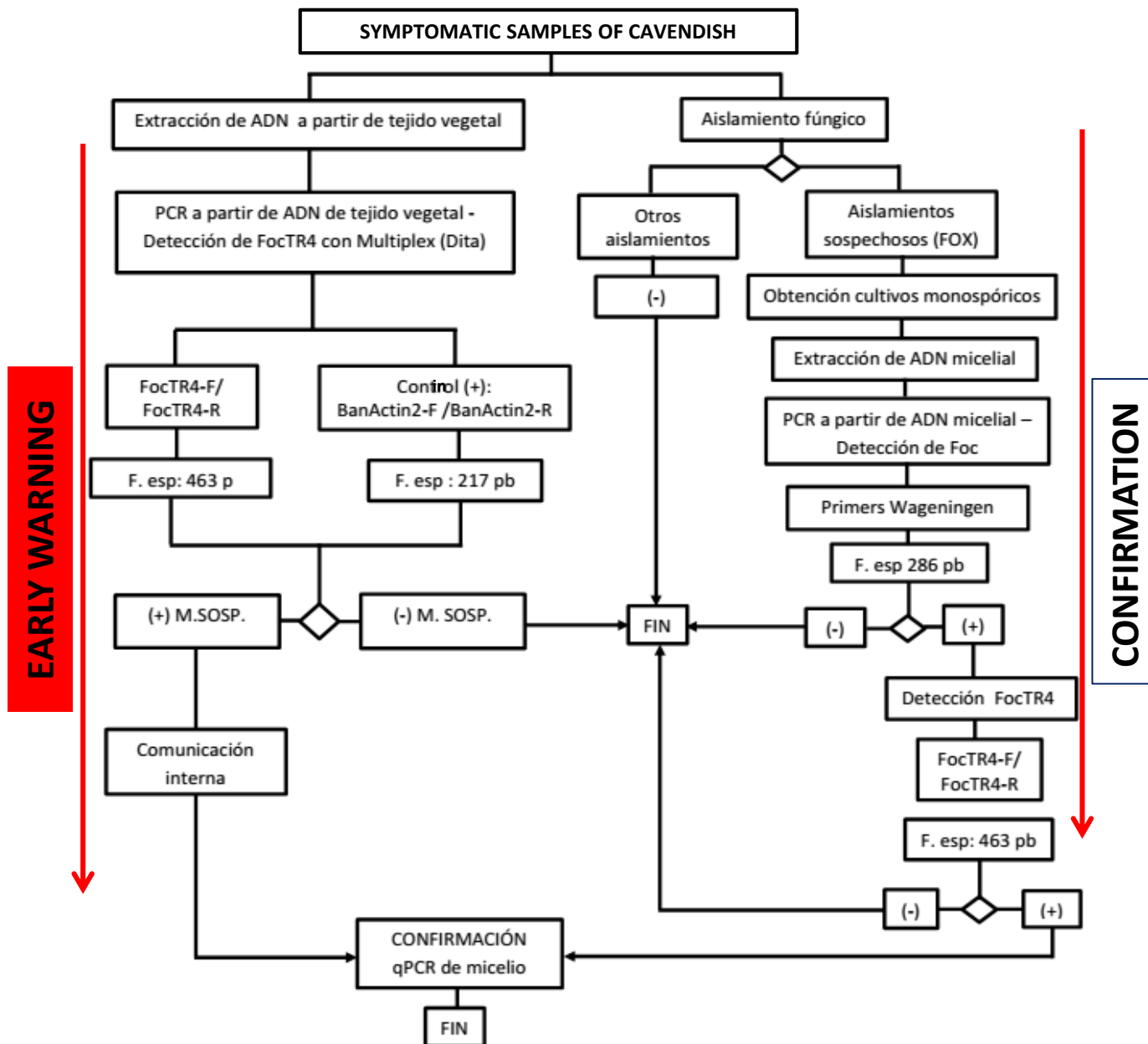
# Preliminary aspects for the FocTR4 detection method

The procedure was initially determined according to the type of sample to be processed!

Generation of different alarm levels



\*Frequent samples



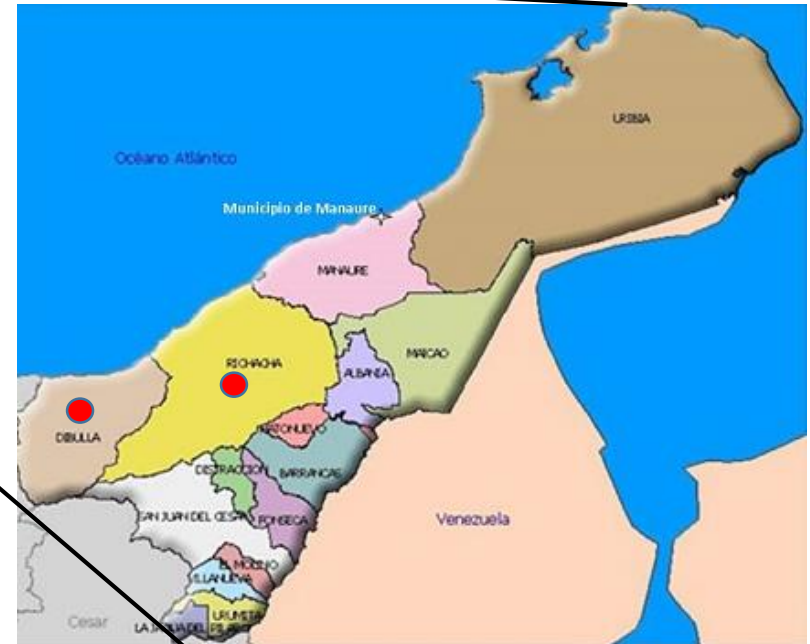
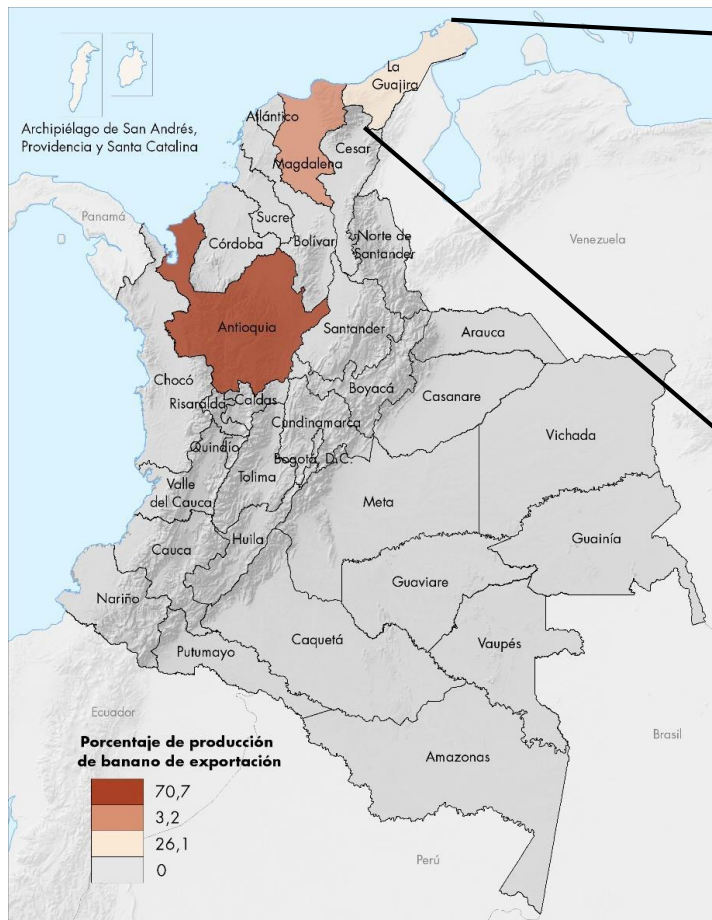


# 2019: First report of FocTR4 in Colombia



**Cavendish plants with suspicious symptoms are identified in the field**

# 2019: First report of FocTR4 in Colombia



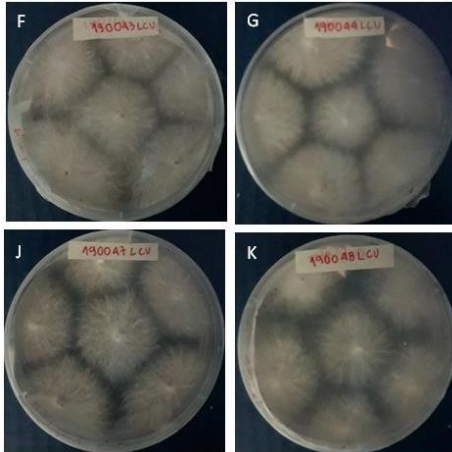
La Guajira County – Riohacha and Dibulla municipalities affected areas



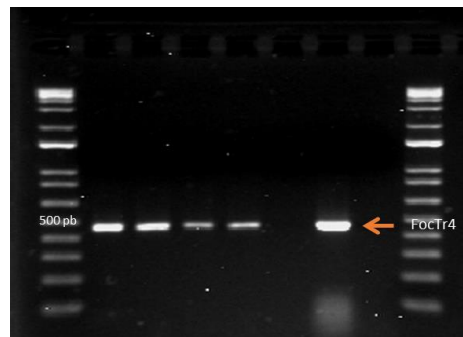
# 2019: First report of FocTR4 in Colombia

## Official Samples

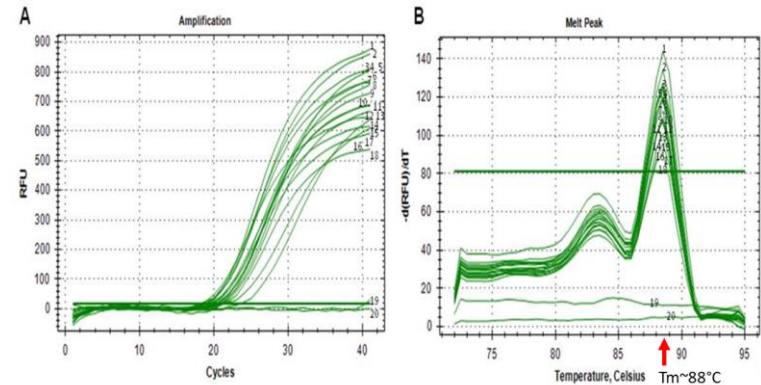
Fungus Isolation



IGS PCR end point



IGS qPCR Detection in tissue



Confirmation process of preliminary results  
obtained in ICA's laboratories



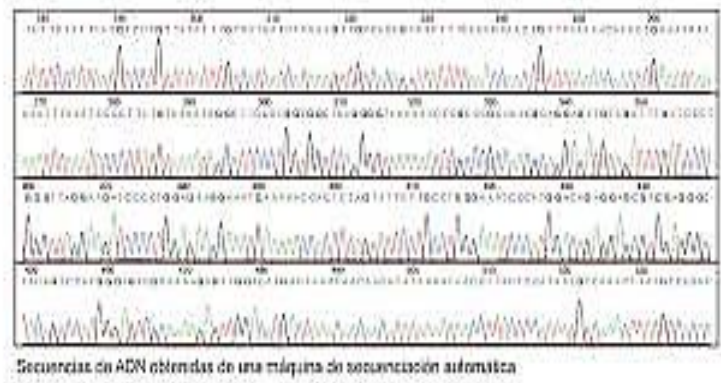
International Reference Laboratory- Wageningen University  
Dr. Fernando García



# 2019: First report of FocTR4 in Colombia

Implementation of complementary methodologies for confirmation process

## Next Generation Sequencing - NGS



VCGs were not performed  
Spend a lot of time!!

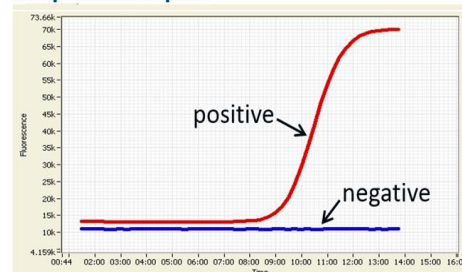
## Pathogenicity tests



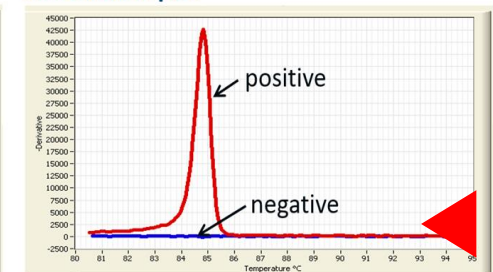
## LAMP: Loop Mediated Isothermal Amplification



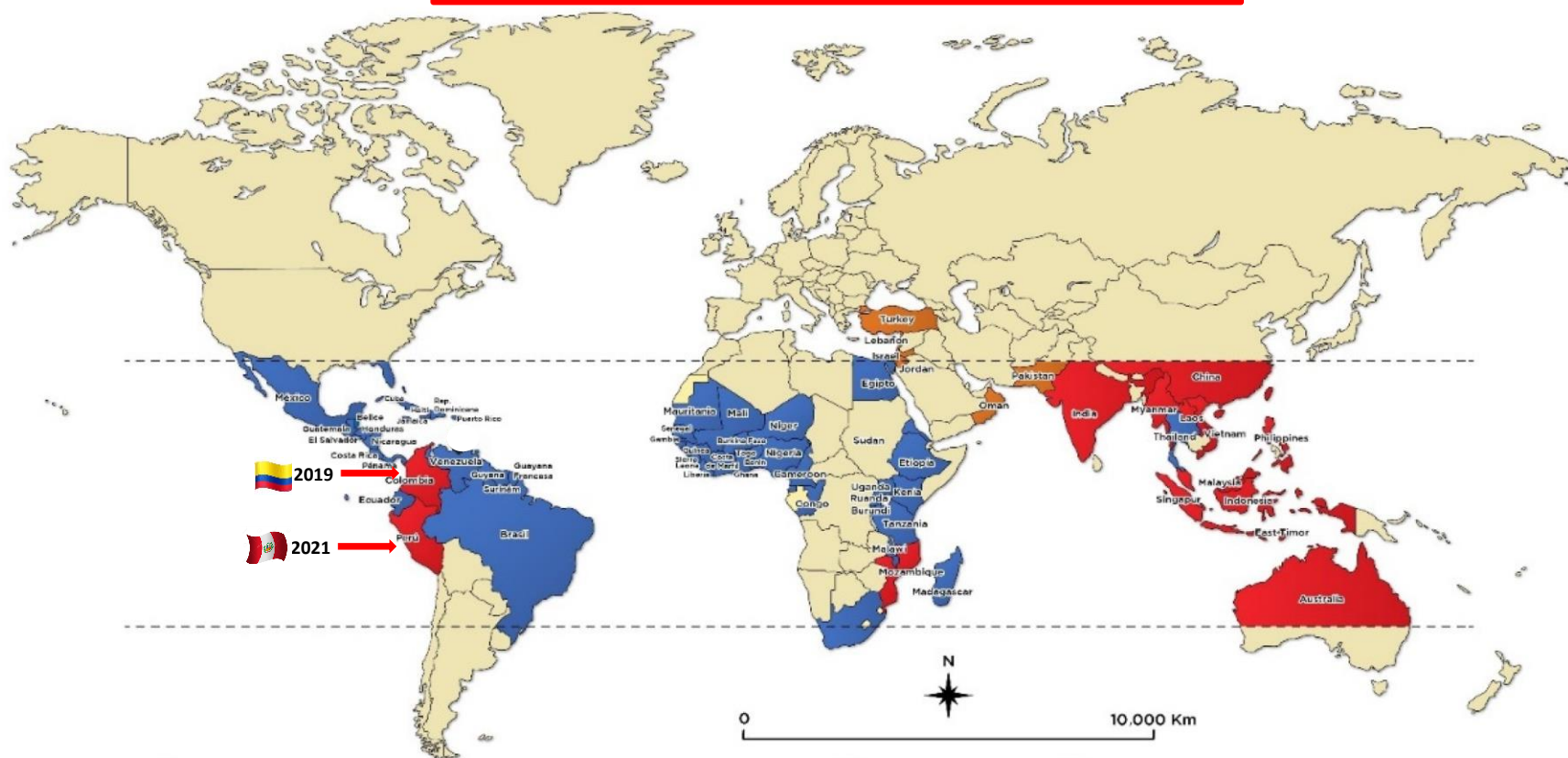
### Amplification plot



### Dissociation plot



# FocTR4 Current status



**R1  
R2**

Belice	Cuba	Guayana	Mexico	Sierra leona
Benin	Ecuador	Guayana F.	Nicaragua	South Africa
Brasil	Egipto	Haiti	Niger	Surinam
Burkina F.	Honduras	Kenia	Nigeria	Tanzania
Burundi	Etiopia	Madagascar	Panamá	Thailand
Cameron	Liberia	Puerto Rico	Togo	Uganda
Congo	Gambia	Rep. Dom.	Uganda	Venezuela
Costa de M.	Guatemala	Ruanda		
Costa Rica	Guinea	Senegal		

**TR4**

Israel  
Jordan  
Lebanon  
Mayotte  
Oman  
Pakistan  
Turkey

**TR4,  
R1, R2**

Australia  
China  
Colombia  
India  
Indonesia  
Laos  
Malaysia  
Mozambique  
Myanmar  
Peru  
Philippines  
Taiwan  
Vietnam

Modified from Olivares *et al.*, 2021

# Activities for improving *FocTR4* diagnostic process

With the new *FocTR4* phytosanitary status for Colombia.....

## • Diagnostic Process

- To strengthen the *FocTR4* detection methodologies: different molecular markers
- To ensure reliability and improve the results report timeframe delivery.
- To implement the guidelines describe in the ISO/IEC-17025 standard

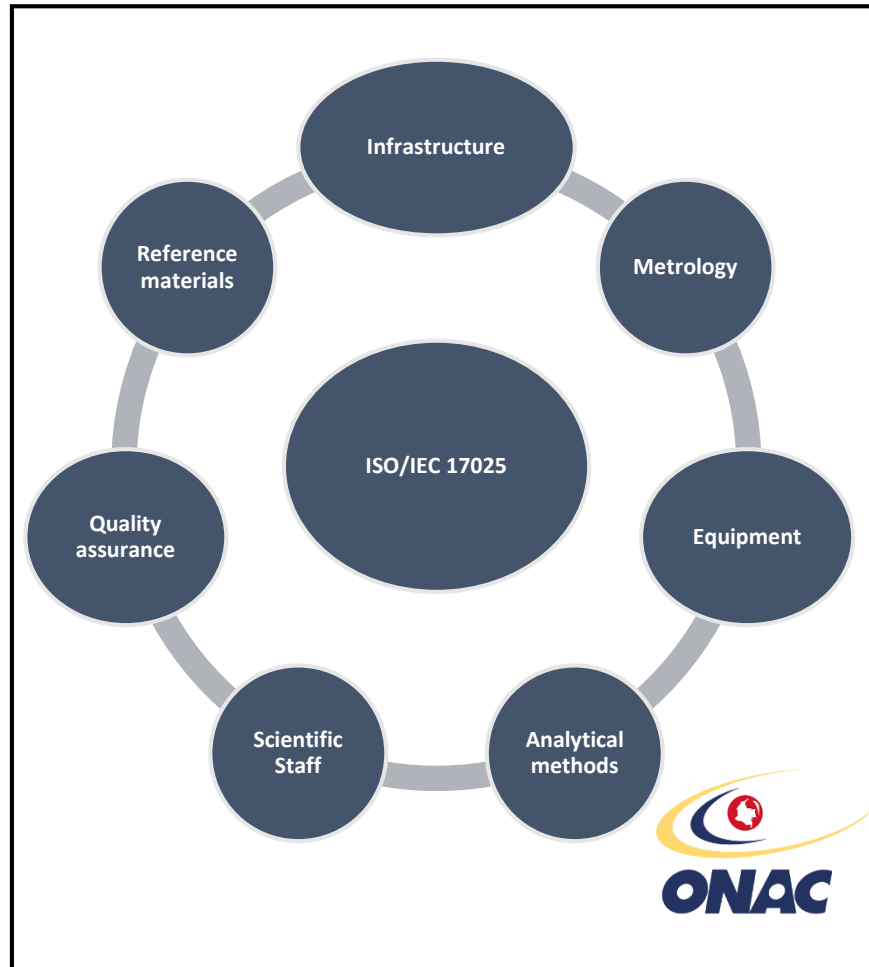
## • Analytical capacity

- To enable others ICA's laboratories to face the increased demand generated by official surveillance plans.

## • Biosecurity

- To minimize the risks of dispersion related to the mobilization of samples.
- To ensure the biological contention in the laboratories using a specific area with Good Laboratory Practice and biosecurity infrastructure.

# Activities for improving *FocTR4* diagnostic process



**Official control beyond of the detection protocols!!.**

The analytical results

For taking good decisions in the diseases management, legal actions, sanctioning processes and phytosanitary protection for the country is necessary adopt the guidelines of a quality management system.



# Activities for improving *FocTR4* diagnostic process

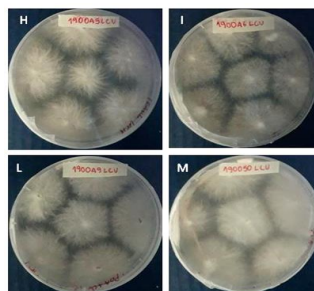
## Samples



### Recommendations

- Refrigeration, packaging reconditioning.
- Avoid excess moisture that accelerates tissue degradation processes.
- Unacceptable sample: high state of oxidation or advanced decomposition.

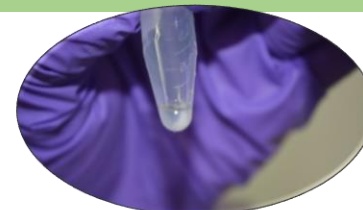
## *FocTR4* isolation conditions



**Tissue disinfection:** NaClO 3%- 1 min, Sterile H<sub>2</sub>O 1 min, Drying with sterile paper

**Tissue planting:** 8 tissue fragments in each Petri plate, 2 repetitions, Incubation 25-28°C, Monitoring 2-3 days

## DNA Extraction



### Tissue/Mycelium Homogenization:

by Liquid Nitrogen-zircon beads

**Cell Lysis:** with 320 µl Extraction Buffer (sorbitol 350 mM, Tris 100 mM pH 8.0, EDTA 5 mM, 0.2% de beta-mercaptoetanol), 320 µl Lysis Buffer (CTAB 55 mM, Tris 200 mM pH 8.0, EDTA 50 mM y NaCl 2M), 100 µl Lauroyl sarcosin 5%

### Proteins and lipids separations:

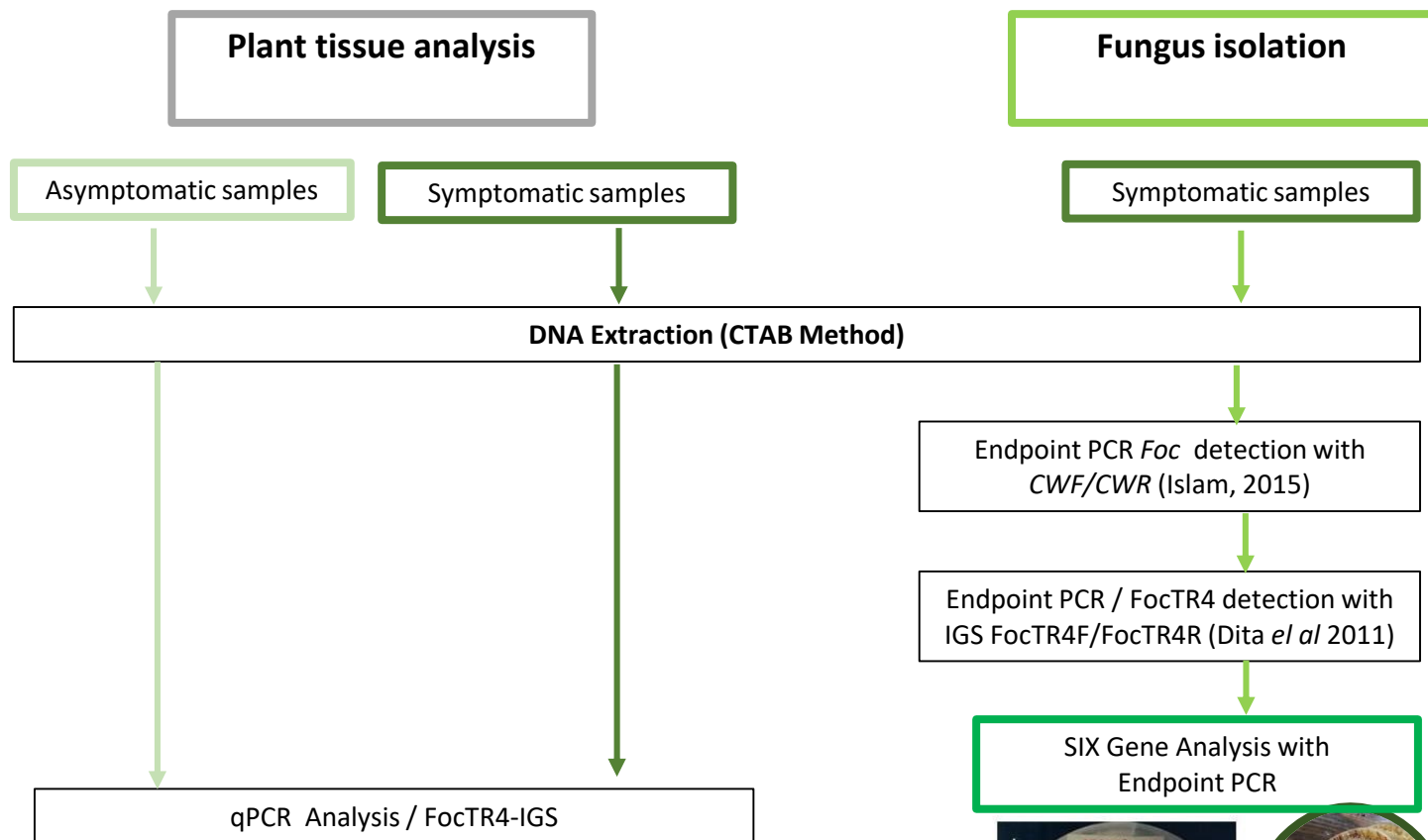
Washing with 1 vol chloroform:  
Isoamilic Alcohol

**Precipitation:** 1 vol Isoamilic Alcohol, 50 µl potassium acetate 5M

**DNA Cleaning:** Washing Ethanol (2), resuspension 100-200 µl H<sub>2</sub>O and quantification

# Activities for improving *Foc*TR4 diagnostic process

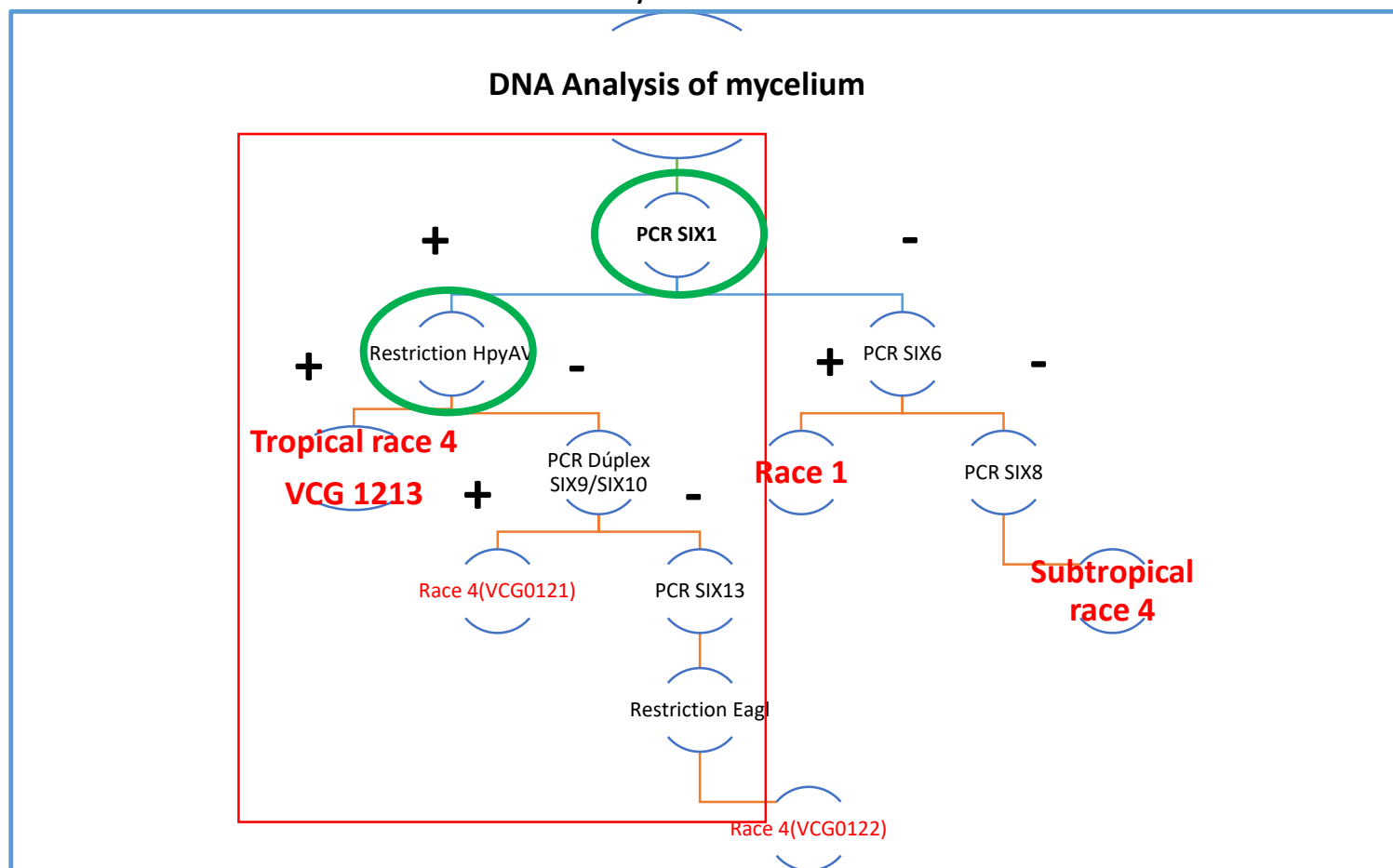
Current scheme for the detection of *Fusarium oxysporum* f. sp. *cubense* TR4



# Activities for improving *FocTR4* diagnostic process

FocTR4-PCR Detection (Carvalho *et al.*, 2019)

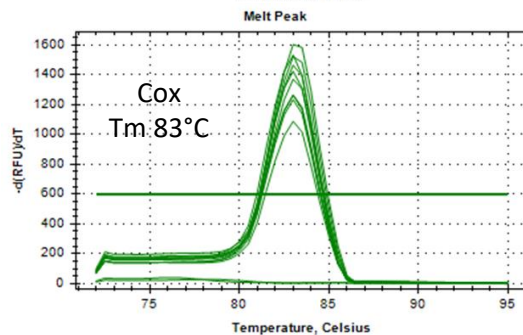
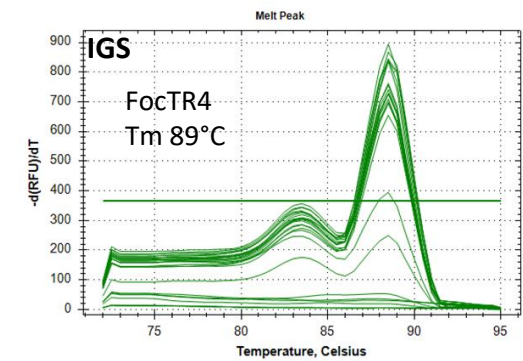
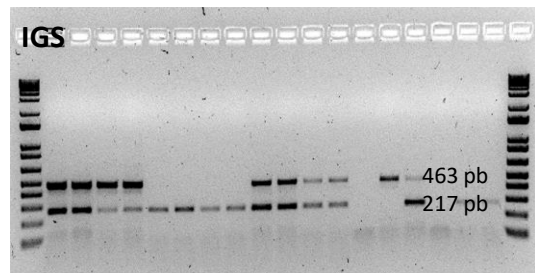
\* Flowchart SIX gene analysis as a complementary protocol for *FocTR4* detection from mycelium



## Activities for improving *FocTR4* diagnostic process

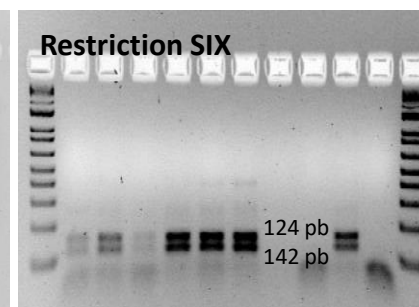
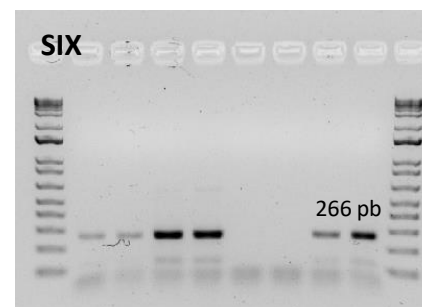
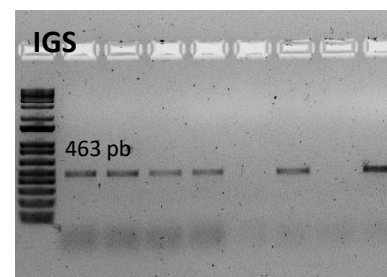
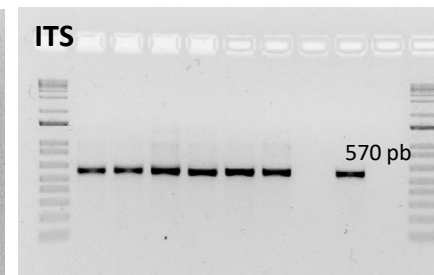
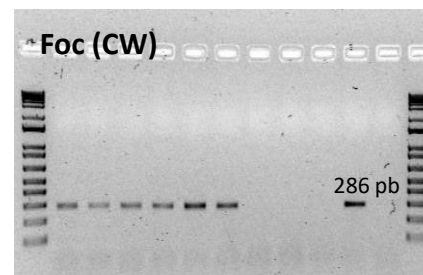
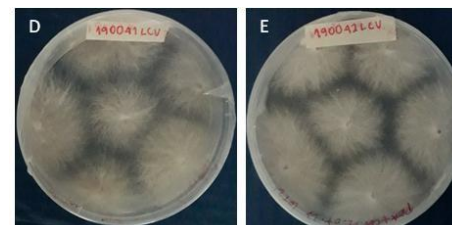
Primers	Sequence 5' - 3'	Fragment size (pb)	Gen Target	References
FocTR4-F	CACGTTTAAGGTGCCATGAGAG	463	IGS	Dita <i>et al.</i> (2010); Dita <i>et al.</i> (2011)
FocTR4-R	GCCAGGACTGCCTCGTGA			
BanActin2-F	ACAGTGCTCTGGATTGGAGGC	217	Banana's Actin Gen (Internal control)	Dita <i>et al.</i> (2010)
BanActin2-R	GCACTTCATGTGGACAATGG			
CWF1	CCTGATACCCAGACGGCTAA	286	Putative protein	Islam (2015)
CWR1	CTGTCGGCTTCACCGTTATT			
SIX1_266_F	GTGACCAGAACTTGCCCACA	266	SIX Gen	Carvalhais <i>et al.</i> (2019)
SIX1_266_R	CTTTGATAAGCACCATCAA			
ITS-4	TCCTCCGCTTATTGATATGC	575	ITS	White <i>et al.</i> , 1990
ITS-5	GGAAGTAAAAGTCGTAACAAGG			





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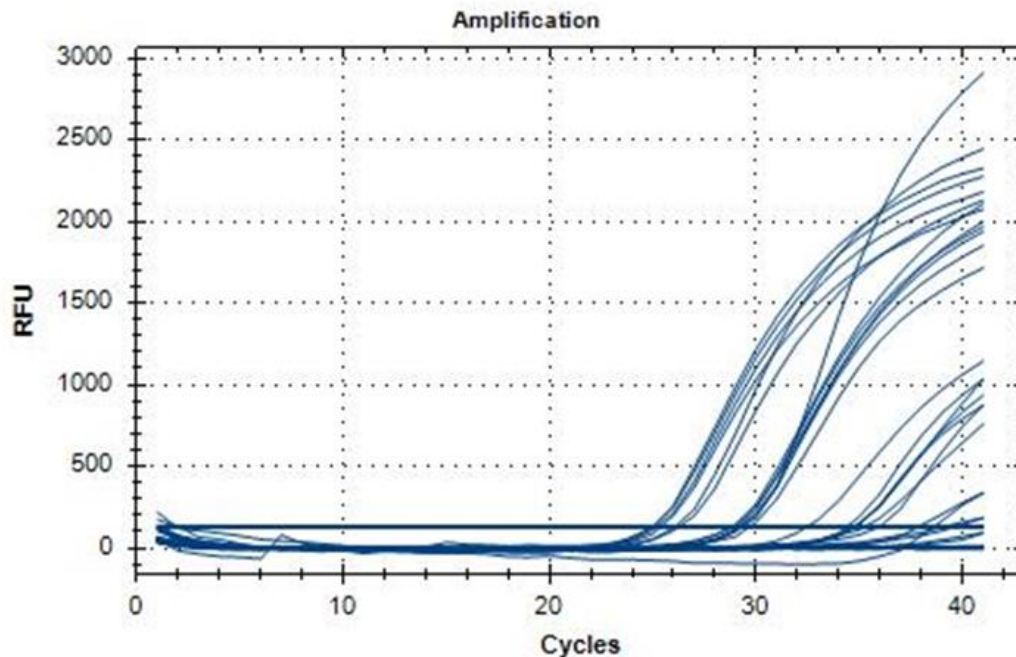
M  
Y  
C  
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U  
M



# Activities for improving *FocTR4* diagnostic process

## Additional *FocTR4* detection Methodology

Primers / Probe	Sequence 5' - 3'	Gen Target	References
FWB-TR4 F	CGGTCTCGGCCAAATCTGATT	Hypothetical protein	Aguayo <i>et al.</i> (2017)
FWB-TR4 R	ACGACTTATCTAGCGGTTGATGTG		
FWB-TR4 P	ACCCTTCAACTCCACTCGATCGCA		



# Activities for improving *FocTR4* diagnostic process

## ... Additional activities carried out by ICA

Andean Community- CAN project: Regional standardization of the diagnosis of *Fusarium oxysporum* f. sp. *cubense* Tropical Race 4 (FocTR4)

- Workshop capacities in FocTR4 diagnostic- for CAN countries
- Standardization of diagnostic protocols
- Proficiency test: quality assurance to verify technical competence of laboratories within the framework of ISO IEC17043 standard.

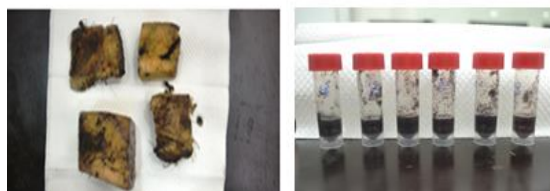
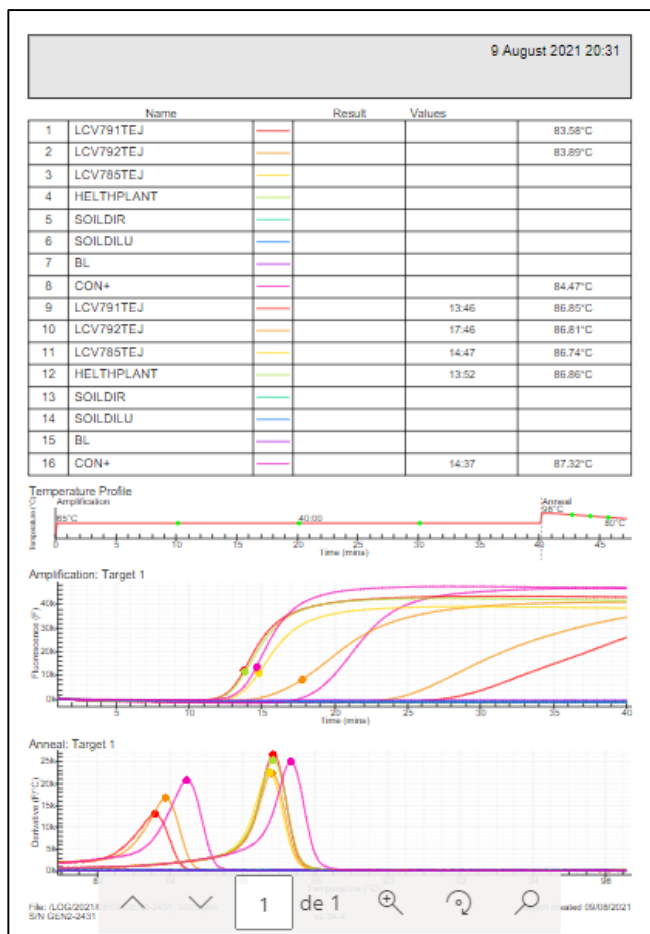




# New Methodology evaluation

## LAMP: Loop-mediated isothermal amplification

**Great POTENTIAL for FocTR4 Diagnosis/ Specificity in-situ**





## ...Working

- Performance evaluation of molecular markers designed by AGROSAVIA for FocTR4 detection .
- ICA: Selection of molecular markers by comparative evaluation of attributes for further validation
- Development and implementation new methods for FocTR4 in soil.

**AGROSAVIA**  
Corporación colombiana de investigación agropecuaria

**ICA**  
Instituto Colombiano Agropecuario

**AGUGRA**  
Asociación de Agricultores y Ganaderos de Colombia

**ASBAMA**  
Asociación de Bancos de Semillas de Colombia

**ICA** 55 años  
Instituto Colombiano Agropecuario

 **MINAGRICULTURA**

 **GOBIERNO DE COLOMBIA**



# Analytical capacity

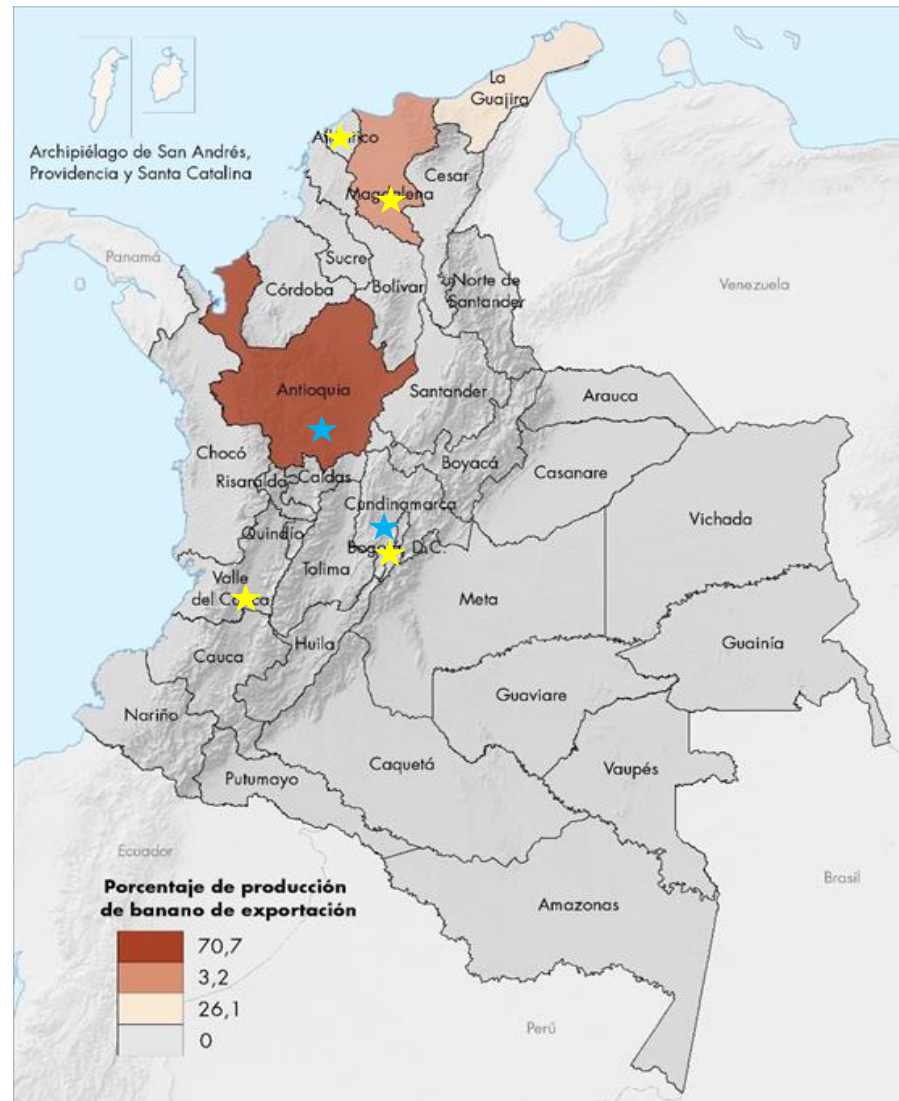
2017

**2** Authorized Laboratories

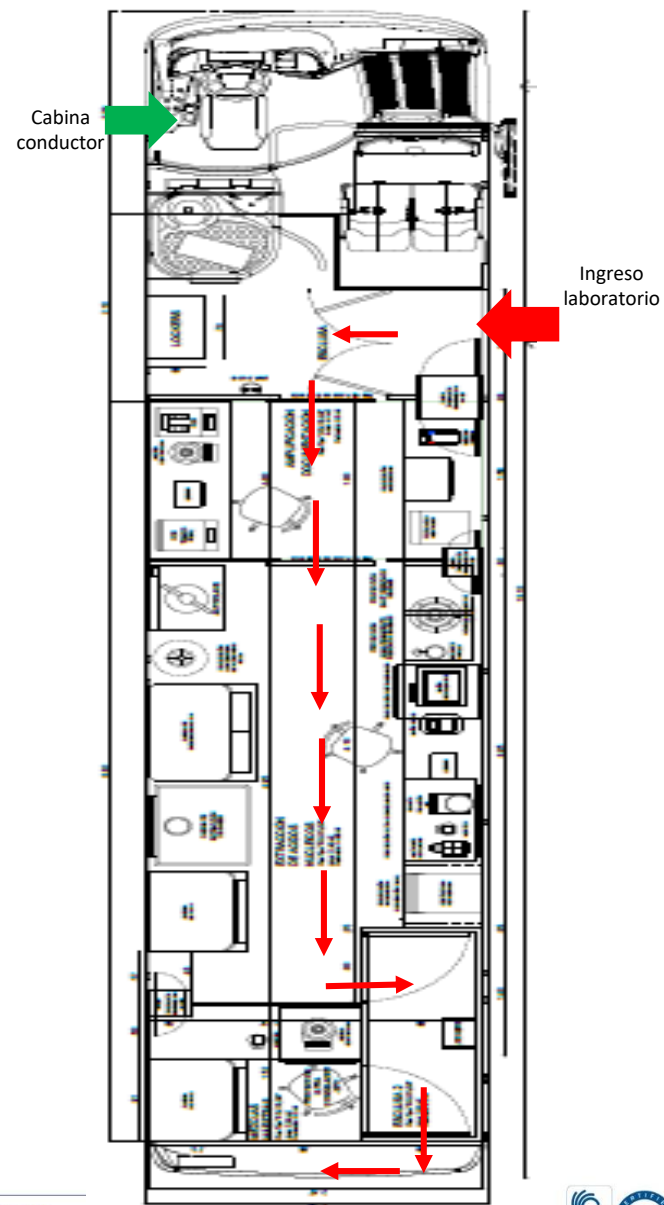


2021

**6** Authorized Laboratories



# Mobile Laboratory

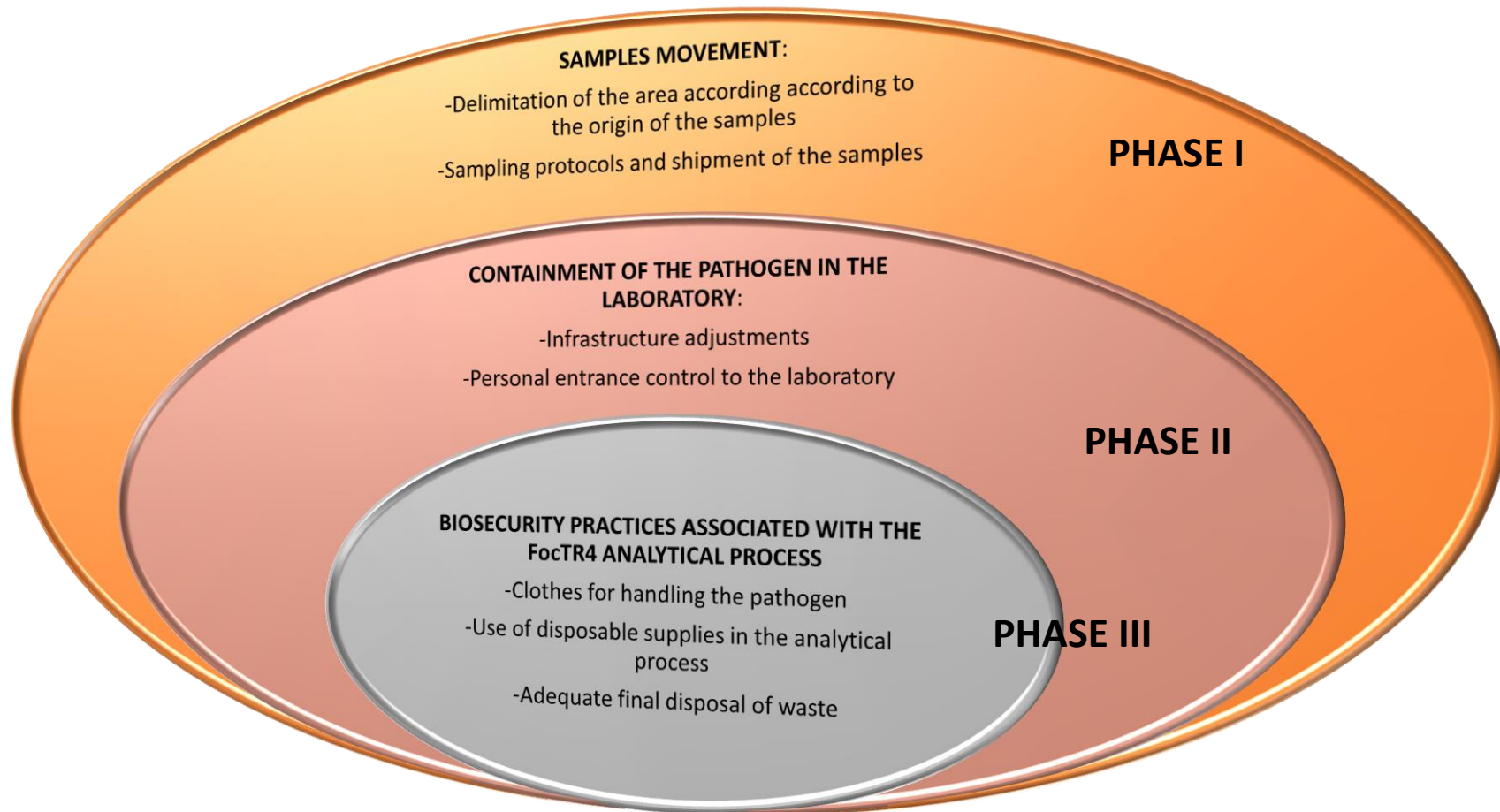


## BIOSECURITY PRACTICES IMPLEMENTED

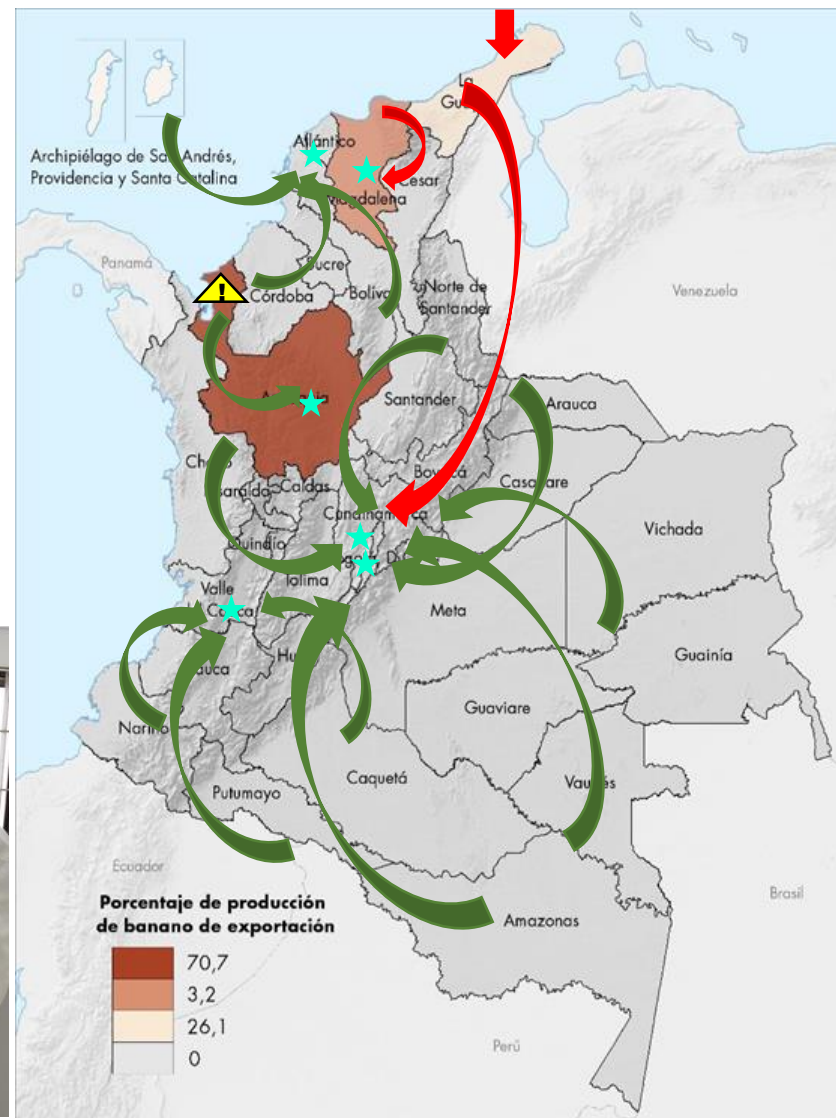
1. To establish protocols for taking, packaging and sending samples to the laboratories.
1. To manage the samples shipment according to the origin and risk of the area sampling.
2. To define and build contention areas inside of the laboratories.
3. To design of biosecurity protocols for sample handling, and waste final disposition.



## BIOSECURITY PRACTICES IMPLEMENTED



## BIOSECURITY PRACTICES IMPLEMENTED





# CONTENTION AREA

## BIOSECURITY PRACTICES IMPLEMENTED



# Phytosanitary requirements verification


**LABORATORIO DE CUARENTENA VEGETAL-LCV**  
**2516 masl**



**Diagnostic Platform:**  
**8 Viruses**  
**1 Fungus**  
**1 Bacteria**





VIRUS	MÉTODOS	CONTROLES	REGIÓN GENÓMICA	REFERENCIA
<i>Banana bunchy top virus</i> (BBTV) Babuvirus – Nanoviridae	PCR	Fragmento Clonado	Proteína de la Master Replicasa 479 pb	Selvajaran et al (2011) Current Science. 100(1):10
<i>Abaca bunchy top virus</i> (ABTV) Babuvirus – Nanoviridae	PCR	Sintético	DNA S	Sharma et al (2007) Arch Virol (2008) 153: 135–147.
<i>Cucumber mosaic virus</i> (CMV) Cucumovirus – Bromoviridae	RT – PCR	Material Vegetal comercial para diagnóstico	Proteína de la cápside	Choi et al (1999) Journal of Virological Methods 83:67–73
<i>Banana bract mosaic virus</i> (BBrMV) Potyviridae - Potyvirus	RT – PCR	Sintético	Proteína de la cápside	Iskra-Caruana et al. (2008) J. Virol. Methods 153:223, 2008. (ANSES)
<i>Banana mild mosaic virus</i> (BanMMV) - Betaflexiviridae	RT – PCR	Sintético	Proteína de cápside	Teycheney (2005) Journal of General Virology, 86, 3179–3187.
<i>Banana virus X</i> (BVX) Betaflexiviridae	RT – PCR	Sintético	Proteína de la replicación	Teycheney (2005) Arch. Virol. 150: 1715-1727.
<i>Abaca mosaic virus</i> – (SCMV, Ab) Potyviridae – Potyvirus	RT – PCR	Sintético	Extremo 3' y Proteína de cápside	Gambley et al (2004) Australasian Plant Pathology. 33, 475–484
VIRUS	MÉTODOS	CONTROLES	REGIÓN GENÓMICA	REFERENCIA
<i>Banana streak badnavirus</i> (BSVs) Badnavirus – Caulimoviridae	Inmuncaptura PCR • Imové BSIMV • Mysore BSMYV • Gold Finger BSGFV • Obino l'ewai BSOLV			Geering et al., unpublished; Le Provost et al., 2006, Journal of Virological Methods
HONGOS	MÉTODOS	CONTROLES	REGIÓN GENÓMICA	REFERENCIA
<i>Fusarium oxysporum</i> f. sp. <i>cubense</i> Raza 4 Tropical <i>FocR4T</i> (Asintomáticas)	Aislamiento del hongo, PCR y qPCR	Hongo purificado		
BACTERIA	MÉTODOS	CONTROLES	REGIÓN GENÓMICA	REFERENCIA
<i>Ralstonia solanacearum</i> Raza 2	PCR duplex	Bacteria purificada	Filotipo II patogénicas del Moko (secuevar IIB-4) y su variante fitopatológica no patogénica a musáceas (IIB-4NPB)	

# THANK YOU!!

# Receipt of samples

## Necrosis in vascular bundles



- Refrigeration, packaging reconditioning.
- Avoid excess moisture that accelerates tissue degradation processes.



- Unsuitable sample: in a state of oxidation or advanced decomposition.

Fotos: LDFAN, ICA 2016

# Isolation



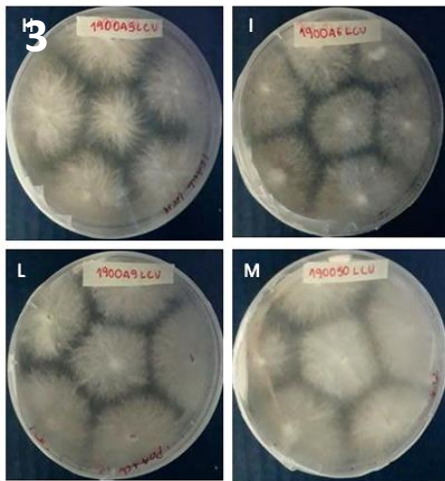
## Disinfection of tissue :

- NaClO 3%- 1 min
- Sterile H2O 1 min
- Drying with sterile paper



## Tissue planting :

- 8 explantes
- 2 repetitions
- Incubation 25-28°C
- Monitoring 2-3 days



## Obtaining isolates :

- DNA Extraction





# DNA Extraction

## 1. Tissue maceration/mycelium(100-200 mg)

Liquid Nitrogen  
TissueHomogenizer- zircon beads

## 2. Cell Lysis: Incubation 1h - 65°C

320 µl Extraction Buffer (sorbitol 350 mM, Tris 100 mM pH 8.0, EDTA 5 mM, 0.2% de beta-mercaptoetanol)

320 µl Lysis Buffer (CTAB 55 mM, Tris 200 mM pH 8.0, EDTA 50 mM y NaCl 2M)

100 µl Lauroyl sarcosin 5%

## 7. Quantification



## 6. Resuspension

100-200 µl H<sub>2</sub>O

## 5. DNA Cleaning

Washing Ethanol (2)

## 3. Separation of proteins and lipids

Washing with 1 vol  
chloroform :Isoamlic Alcohol

## 4. Precipitation

1 vol Isoamlic Alcohol  
50 µl potassium acetate 5M