Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
[1]	G	Editorial	Use latin name only for the pest throughout the text, Add authority (Everts) after the name in the title Ensure that authority names are used for species names at their first mention in the text and not thereafter (e.g. paragraphs 88, 112,113, 118) Consider putting “Khapra beetle” in brackets in the title	1. Consistency. So-called common names vary between countries, and in a global standard it is inappropriate to use one country's preference. 2 & 3. Consistency with Thrips palmi protocol 4. As the name Khapra beetle is globally known, it might be helpful for NPPOs to have a reference in the title.	EPPO,Russian Federation ,European Union ,Ukraine ,Morocco ,Uzbekistan
[2]	G	Editorial	Suggest that a ‘most similar to’ or ‘easily confused with’ paragraph/section be added for each species (adults and larvae)		Australia
[3]	G	Editorial		Rewording is needed to add clarity to the text because some words are joined together in the sentences. Paragraphs: 158,161,179.	Costa Rica ,Nicaragua ,El Salvador ,OIRSA
[4]	G	Editorial	1) We recommend this Annex, as well as all the other diagnostic protocols, to be made into technical documents. 2) We recommend that the 'k' in Khapra beetle not be capitalized wherever the name appears in the text and figure captions. 3) We recommend that listings of citations be ordered chronologically rather than alphabetically. See paragraphs 12, 55, 81, and 106. If IPPC prefers alphabetically, then all places in document should follow the same rule.	1) Diagnostic documents can become out of date fast as new tools, methodologies, and methods are continually developed. If made into a technical document, it would be easier to make updates versus if it is made into an annex to an ISPM. 2) "Khapra" means destroyer in the Hindi language. It is not a proper name or place. The Entomological Society of America List of Common Names officially cites 'khapra beetle' as the spelling, without capitalization. Webster's New World Dictionary and many other sources on the Internet do not capitalize the word 'khapra'. 3) The ordering of multiple citations should be done in a consistent manner throughout the document.	United States of America
[5]	G	Substantive	1. Reorder section 3 to make it clearer e.g. start with general information on life stages, followed by detection in commodities including methods used, detection in storage places and methods and finally how to handle specimens (paras 27, 28, 29, 31, 32, 30, 34, 33); 2. In section 4, keep all the information on identification of larvae together (sections 4.1, 4.3.1, 4.4, 4.5) followed by information on adults (sections 4.2, 4.3.2, 4.6, 4.7). Numbering and some titles would have to be adjusted as a result; 3. Tabulate certain sections e.g. keys (4.3.1, 4.3.2, 4.5.1, 4.7.10) or the distinguishing characteristics as in T. palmi (sections 4.4.2, 4.5.2, 4.6.2). Tables could be formatted to ensure that names and numbers are aligned to the right.	Parts of the protocol are confusing and proposals have been made to improve the draft and bring it into line with the Thrips palmi protocol. Some proposed micrographs are provided at para 159. EPPO Secretariat to obtain a document from the Russian expert with the photos at paragraph 159	EPPO,Russian Federation ,Ukraine ,Morocco ,Uzbekistan

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			4. Consider simplification by deletion of sections 4.3.1 and 4.6. The information appears to be repeated in other parts of the protocol. If the characters listed in these sections are essential for diagnosis this should be made clear. 5. Include micrographs where possible.		
[6]	G	Substantive		It is recommended to add photographic images of all life stages of the beetle and images of damages. This should	Yemen ,Oman
[7]	G	Substantive		A general review of photographs and pictures would be necessary. The images that appear in the protocol are not of good quality, making it difficult the observation of the details cited in the text.	COSAVE,Paraguay ,Chile,Brazil
[8]	G	Substantive		Will be necessary to specify the numbers of days or hours in such parts of the text that we found for exmple periods of time, as paragraph 52: "Slides and pinned insects should be labelled immediately after mounting the specimens. The slides should be placed in an oven for a few days at 40ºC. After drying all slides should be ringed (see 4.1). " In this case, How many days will be suficient to obtain the better results?	Mexico
[9]	G	Substantive		A general review of photographs and pictures would be necessary. The images that appear in the protocol are not of good quality, making it difficult the observation of the details cited in the text	Argentina
[10]	G	Substantive	1. Reorder section 3 to make it clearer e.g. start with general information on life stages, followed by detection in commodities including methods used, detection in storage places and methods and finally how to handle specimens (paras 27, 28, 29, 31, 32, 30, 34, 33); 2. In section 4, keep all the information on identification of larvae together (sections 4.1, 4.3.1, 4.4, 4.5) followed by information on adults (sections 4.2, 4.3.2, 4.6, 4.7). Numbering and some titles would have to be adjusted as a result; 3. Tabulate certain sections e.g. keys (4.3.1, 4.3.2, 4.5.1, 4.7.10) or the distinguishing characteristics as in T. palmi (sections 4.4.2, 4.5.2, 4.6.2). Tables could be formatted to ensure that names and numbers are aligned to the right.	Parts of the protocol are confusing and proposals have been made to improve the draft and bring it into line with the Thrips palmi protocol. Some proposed micrographs are provided at para [159].	European Union

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			4. Consider simplification by deletion of sections 4.3.1 and 4.6. The information appears to be repeated in other parts of the protocol. If the characters listed in these sections are essential for diagnosis this should be made clear. 5. Include micrographs where possible.		
[11]	G	Technical	Figures are very helpful in correct and rapid identification in both larval and adult stages. As there are many characters difficult to use in the identification without the ir figures, the following with morphological terms should be added; -habitus of adults:see Peacock, E.R., 1993. P83 -habitus of larvae: see Peacock, E.R. 1993. P121 -patterns of elytra: see Beal, R.S. 1954. P39 -position of hastisetae: see Peacock, E.R. 1993. P122 -antennae of larvae: see Peacock, E.R. 1993. P138		Japan
[12]	G	Technical		A general review of photographs and pictures will be necessary. The images that appear in the protocol are not of good quality, making it difficult to observe the details cited in the text. The specific time frame should be mentioned in those paragraphs that states "a few days" to provide better guidance. Example: paragraph 45 and 52. There are more suitable mounting media available other than Hoyer´s medium, therefore throughout the text we propose to add "or similar media" after mentioning Hoyer´s.	Costa Rica ,Nicaragua ,El Salvador ,OIRSA
[13]	G	Technical		A general review of photographs and pictures will be necessary. The images that appear in the protocol are not of good quality, making it difficult to observe the details cited in the text. The specific time frame should be mentioned in those paragraphs that states “few days” to provide better guidance (example paragraph 45 and 52). Rewording is needed to add clarity to the text because some words are joined together in the sentences in paragraphs 158, 161 and 179	Uruguay
[14]	G	Technical		A general review of photographies and pictures will be neccesary. The images that appear in the protocol does not have good resolution and quality, making it difficult to observe some	Mexico

<table>
<tr><th>Comment no.</th><th>Paragraph no.</th><th>Comment type</th><th>Comment</th><th>Explanation</th><th>Country</th></tr>
<tr><td></td><td></td><td></td><td></td><td>relevant taxonomical detail cited in the text.</td><td></td></tr>
<tr><td>[15]</td><td>G</td><td>Technical</td><td>The quality of this diagnostic protocol is very good. However, the diagnostic protocol does not mention the use of Berlese funnnels to verify for the presence of T. granarium.
Canada and other countries use Berlese funnels regularly and this type of equipment has proven to be successful in collecting adults and larvae of Trogoderma.
The protocol does not make reference to Gorham's 1991 Insect and Mite Pests in Food (Vol 1 & II) or Y. Bousquet's (1990), Beetles associated with stored products in Canada: An identification guide in the literature. These are important publications that are commonly used and should be added to the list of references.
Suggestions for rapid clearing and mounting would be benefical for the end users of this diagnostic protocol. Results typically need to be supplied within 24 hours of the initial inspection, so there is no time for heat treatment of Hoyer's.</td><td></td><td>Canada</td></tr>
<tr><td>[16]</td><td>3</td><td>Editorial</td><td><table>
<tr><td>Date of this document</td><td>2011-06-20</td></tr>
<tr><td>Document category</td><td>Draft new annex to ISPM 27:2006 (Diagnostic protocols for regulated pests)</td></tr>
<tr><td>Current document stage</td><td>Draft for member consultation 2011.</td></tr>
<tr><td>Origin</td><td>SC added subject:2004-006 under topic:2006-007, Insects and mites</td></tr>
<tr><td>Major stages</td><td>Approved for member consultation by the SC</td></tr>
<tr><td>Consultation on technical level</td><td>The first draft of this diagnostic protocol was written by:
The first draft of this protocol was written by Andras Szito (Department of Agriculture and Food Western Australia, Plant Biosecurity Branch, South Perth, Australia); Witold Karnkowski (Main Inspectorate of Plant Health and Seed Service, Central Laboratory, Toruń, Poland) and Alba Enrique de Briano</td></tr>
</table></td><td>Correction to name and institution title.</td><td>EPPO,Russian Federation ,Ukraine ,Morocco ,Uzbekistan</td></tr>
</table>

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			(Laboratorio de Plagas y Enfermedades de las Plantas, SENASA, Buenos Aires, Argentina). This proposal has been reviewed by Dr. R. S. Beal Jr (Prescott, USA), Dr Marcin Kadej (Instytut Zoologiczny, Uniwersytet Wrocławski, Wrocław, Poland), Dr Alan V. Barak (USDA, APHIS, PPQ, CPHST- Otis Laboratory, Buzzards Bay MA, USA), Prof. Chris Haines (Natural Resources Institute, University of Greenwich at Medway, Kent, UK) and MDr. J.C. Ostoja-Starzewski (The Food and Environmental Research Agency, York, United Kingdom)		
			Main discussion points during development of the diagnostic protocol: N/A		
			Notes: Formatted in template of February 2010. Auto numbered ¶.		
[17]	3	Editorial	**Date of this document**: 2011-06-20	The sentence is repeated.	Costa Rica ,Nicaragua ,OIRSA
			Document category: Draft new annex to ISPM 27:2006 (*Diagnostic protocols for regulated pests)*		
			Current document stage: Draft for member consultation 2011.		
			Origin: SC added subject:2004-006 under topic:2006-007, Insects and mites		
			Major stages: Approved for member consultation by the SC		
			Consultation on technical level: ~~The first draft of this diagnostic protocol was written by:~~ The first draft of this protocol was		

Comment no.	Paragraph no.	Comment type	Comment		Explanation	Country
				written by Andras Szito (Department of Agriculture and Food Western Australia, Plant Biosecurity Branch, South Perth, Australia); Witold Karnkowski (Main Inspectorate of Plant Health and Seed Service, Central Laboratory, Toruń, Poland) and Alba Enrique de Briano (Laboratorio de Plagas y Enfermedades de las Plantas, SENASA, Buenos Aires, Argentina). This proposal has been reviewed by Dr. R. S. Beal Jr (Prescott, USA), Dr Marcin Kadej (Instytut Zoologiczny, Uniwersytet Wrocławski, Wrocław, Poland), Dr Alan V. Barak (USDA, APHIS, PPQ, CPHST- Otis Laboratory, Buzzards Bay MA, USA), Prof. Chris Haines (Natural Resources Institute, University of Greenwich at Medway, Kent, UK) and Dr. C. Ostoja-Starzewski (The Food and Environmental Research Agency, York, United Kingdom)		
			Main discussion points during development of the diagnostic protocol	N/A		
			Notes	Formatted in template of February 2010. Auto numbered ¶.		
[18]	3	Editorial	**Date of this document**	2011-06-20	The sentence is repeated	COSAVE,Paraguay ,Chile,Brazil
			Document category	Draft new annex to ISPM 27:2006 (*Diagnostic protocols for regulated pests)*		
			Current document stage	Draft for member consultation 2011.		
			Origin	SC added subject:2004-006 under topic:2006-007, Insects and mites		

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			Major stages: Approved for member consultation by the SC **Consultation on technical level**: The first draft of this diagnostic protocol was written by~~:The first draft of this protocol was written by~~ Andras Szito (Department of Agriculture and Food Western Australia, Plant Biosecurity Branch, South Perth, Australia); Witold Karnkowski (Main Inspectorate of Plant Health and Seed Service, Central Laboratory, Toruń, Poland) and Alba Enrique de Briano (Laboratorio de Plagas y Enfermedades de las Plantas, SENASA, Buenos Aires, Argentina). This proposal has been reviewed by Dr. R. S. Beal Jr (Prescott, USA), Dr Marcin Kadej (Instytut Zoologiczny, Uniwersytet Wrocławski, Wrocław, Poland), Dr Alan V. Barak (USDA, APHIS, PPQ, CPHST- Otis Laboratory, Buzzards Bay MA, USA), Prof. Chris Haines (Natural Resources Institute, University of Greenwich at Medway, Kent, UK) and Dr. C. Ostoja-Starzewski (The Food and Environmental Research Agency, York, United Kingdom) **Main discussion points during development of the diagnostic protocol**: N/A **Notes**: Formatted in template of February 2010. Auto numbered ¶.		
[19]	3	Editorial	**Date of this document**: 2011-06-20 **Document category**: Draft new annex to ISPM 27:2006 (*Diagnostic protocols for regulated pests)*	Add a state abbreviation for the city of Prescott.	United States of America

Comment no.	Paragraph no.	Comment type	Comment		Explanation	Country
			Current document stage	Draft for member consultation 2011.		
			Origin	SC added subject:2004-006 under topic:2006-007, Insects and mites		
			Major stages	Approved for member consultation by the SC		
			Consultation on technical level	The first draft of this diagnostic protocol was written by: The first draft of this protocol was written by Andras Szito (Department of Agriculture and Food Western Australia, Plant Biosecurity Branch, South Perth, Australia); Witold Karnkowski (Main Inspectorate of Plant Health and Seed Service, Central Laboratory, Toruń, Poland) and Alba Enrique de Briano (Laboratorio de Plagas y Enfermedades de las Plantas, SENASA, Buenos Aires, Argentina). This proposal has been reviewed by Dr. R. S. Beal Jr (Prescott AZ, USA), Dr Marcin Kadej (Instytut Zoologiczny, Uniwersytet Wrocławski, Wrocław, Poland), Dr Alan V. Barak (USDA, APHIS, PPQ, CPHST- Otis Laboratory, Buzzards Bay MA, USA), Prof. Chris Haines (Natural Resources Institute, University of Greenwich at Medway, Kent, UK) and Dr. C. Ostoja-Starzewski (The Food and Environmental Research Agency, York, United Kingdom)		
			Main discussion points during development of the diagnostic protocol	N/A		
			Notes	Formatted in template of February 2010. Auto numbered ¶.		

Comment no.	Paragraph no.	Comment type	Comment		Explanation	Country
[20]	3	Editorial	**Date of this document**	2011-06-20	Delete repeted sentence	Mexico
			Document category	Draft new annex to ISPM 27:2006 (*Diagnostic protocols for regulated pests)*		
			Current document stage	Draft for member consultation 2011.		
			Origin	SC added subject:2004-006 under topic:2006-007, Insects and mites		
			Major stages	Approved for member consultation by the SC		
			Consultation on technical level	~~The first draft of this diagnostic protocol was written by:~~ The first draft of this protocol was written by Andras Szito (Department of Agriculture and Food Western Australia, Plant Biosecurity Branch, South Perth, Australia); Witold Karnkowski (Main Inspectorate of Plant Health and Seed Service, Central Laboratory, Toruń, Poland) and Alba Enrique de Briano (Laboratorio de Plagas y Enfermedades de las Plantas, SENASA, Buenos Aires, Argentina). This proposal has been reviewed by Dr. R. S. Beal Jr (Prescott, USA), Dr Marcin Kadej (Instytut Zoologiczny, Uniwersytet Wrocławski, Wrocław, Poland), Dr Alan V. Barak (USDA, APHIS, PPQ, CPHST- Otis Laboratory, Buzzards Bay MA, USA), Prof. Chris Haines (Natural Resources Institute, University of Greenwich at Medway, Kent, UK) and Dr. C. Ostoja-Starzewski (The Food and Environmental Research Agency, York, United Kingdom)		

Comment no.	Paragraph no.	Comment type	Comment		Explanation	Country
			Main discussion points during development of the diagnostic protocol	N/A		
			Notes	Formatted in template of February 2010. Auto numbered ¶.		
[21]	3	Editorial	**Date of this document**	2011-06-20	The sentence is repeated	Argentina
			Document category	Draft new annex to ISPM 27:2006 (*Diagnostic protocols for regulated pests)*		
			Current document stage	Draft for member consultation 2011.		
			Origin	SC added subject:2004-006 under topic:2006-007, Insects and mites		
			Major stages	Approved for member consultation by the SC		
			Consultation on technical level	The first draft of this diagnostic protocol was written by: ~~The first draft of this protocol was written by~~ Andras Szito (Department of Agriculture and Food Western Australia, Plant Biosecurity Branch, South Perth, Australia); Witold Karnkowski (Main Inspectorate of Plant Health and Seed Service, Central Laboratory, Toruń, Poland) and Alba Enrique de Briano (Laboratorio de Plagas y Enfermedades de las Plantas, SENASA, Buenos Aires, Argentina). This proposal has been reviewed by Dr. R. S. Beal Jr (Prescott, USA), Dr Marcin Kadej (Instytut Zoologiczny, Uniwersytet Wrocławski, Wrocław, Poland), Dr Alan V. Barak (USDA, APHIS, PPQ, CPHST- Otis Laboratory, Buzzards Bay MA, USA),		

Comment no.	Paragraph no.	Comment type	Comment		Explanation	Country
				Prof. Chris Haines (Natural Resources Institute, University of Greenwich at Medway, Kent, UK) and Dr. C. Ostoja-Starzewski (The Food and Environmental Research Agency, York, United Kingdom)		
			Main discussion points during development of the diagnostic protocol	N/A		
			Notes	Formatted in template of February 2010. Auto numbered ¶.		
[22]	3	Editorial	**Date of this document**	2011-06-20	Correction to name and institution title.	European Union
			Document category	Draft new annex to ISPM 27:2006 (*Diagnostic protocols for regulated pests)*		
			Current document stage	Draft for member consultation 2011.		
			Origin	SC added subject:2004-006 under topic:2006-007, Insects and mites		
			Major stages	Approved for member consultation by the SC		
			Consultation on technical level	The first draft of this diagnostic protocol was written by: The first draft of this protocol was written by Andras Szito (Department of Agriculture and Food Western Australia, Plant Biosecurity Branch, South Perth, Australia); Witold Karnkowski (Main Inspectorate of Plant Health and Seed Service, Central Laboratory, Toruń, Poland) and Alba Enrique de Briano (Laboratorio de Plagas y Enfermedades de las Plantas, SENASA, Buenos Aires, Argentina). This proposal has been reviewed by		

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			Dr. R. S. Beal Jr (Prescott, USA), Dr Marcin Kadej (Instytut Zoologiczny, Uniwersytet Wrocławski, Wrocław, Poland), Dr Alan V. Barak (USDA, APHIS, PPQ, CPHST- Otis Laboratory, Buzzards Bay MA, USA), Prof. Chris Haines (Natural Resources Institute, University of Greenwich at Medway, Kent, UK) and Dr. J.C. Ostoja-Starzewski (The Food and Environmental Research Agency, York, United Kingdom) **Main discussion points during development of the diagnostic protocol**: N/A **Notes**: Formatted in template of February 2010. Auto numbered ¶.		
[23]	6	Editorial	The Khapra beetle, *Trogoderma granarium* Everts (Coleoptera: Dermestidae), is a stored product pest of great importance. Its importance lies not only in its ~~capabilities of causing~~capability to cause serious damage to stored dry commodities but ~~also in that~~ countries also face export restrictions for their produce when having established populations of this pest ~~face export restrictions for their produce~~.	To clarify the sentence.	Yemen ,Oman
[24]	6	Editorial	The ~~K~~ khapra beetle, *Trogoderma granarium* Everts (Coleoptera: Dermestidae), is a stored product pest of great importance. Its importance lies not only in its capabilities of causing serious damage to stored dry commodities but also in that countries having established populations of this pest face export restrictions for their produce.	See general comment.	United States of America
[25]	6	Substantive	The Khapra beetle, *Trogoderma granarium* Everts (Coleoptera: Dermestidae), is a stored product pest of great importance. Its importance lies not only in its capabilities of causing serious damage to stored dry commodities but also in that countries having established populations of this pest face export restrictions for their produce. It also found in thye hold of vessel from infested country.		Indonesia
[26]	6	Substantive	~~The Khapra beetle, *Trogoderma granarium* Everts (Coleoptera: Dermestidae), is a stored product pest of great importance. Its importance lies not only in its capabilities of causing serious damage to stored dry commodities but also in that countries having established populations of~~	Add more background information.	China

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			~~this pest face export restrictions for their produce.~~ The Khapra beetle, Trogoderma granarium Everts (Coleoptera: Dermestidae), is a stored product pest of great importance. Its importance lies not only in its capabilities of causing serious damage to stored dry commodities but also in that countries having established populations of this pest face export restrictions for their produce. It also found in the hold of vessel from infested country.		
[27]	6	Substantive	The Khapra beetle, *Trogoderma granarium* Everts (Coleoptera: Dermestidae), is a stored product pest of great importance. Its economic importance lies not only in its capabilities of causing serious damage to stored dry commodities but also in that countries having established populations of this pest face export restrictions for their produce.	To emphasise that the impact is economic as export restrictions may be imposed.	South Africa
[28]	6	Technical	The Khapra beetle, *Trogoderma granarium* Everts (Coleoptera: Dermestidae), is a stored product pest of great importance. Its importance lies not only in its capabilities of causing serious damage to stored dry commodities but also in that countries having established populations of this pest face export restrictions for their produce. It is one of the insects responsible for the spread of *Aspergillus flavus* in stored dried commodities (Simha, 1990).	Another relevant information about the pest was added to emphazise its importance. Aflatoxins are a group of toxic compounds produced by certain strains of the fungus Aspergillus flavus.	Costa Rica ,Nicaragua ,El Salvador
[29]	6	Technical	The Khapra beetle, *Trogoderma granarium* Everts (Coleoptera: Dermestidae), is a stored product pest of great importance. Its importance lies not only in its capabilities of causing serious damage to stored dry commodities but also in that countries having established populations of this pest face export restrictions for their produce. It is one of the insects responsibles for the spread of *Aspergillus flavus* in stored dried commodities (Simha, 1990).	Another relevant information about the pest was added to emphasize its importance. Aflatoxins are a group of toxic compounds produced by certain strains of the fungus Aspergillus flavus	Uruguay
[30]	6	Technical	The Khapra beetle, *Trogoderma granarium* Everts (Coleoptera: Dermestidae), is a stored product pest of great importance. Its importance lies not only in its capabilities of causing serious damage to stored dry commodities but also in that countries having established populations of this pest face export restrictions for their produce. It is one of the responsibles of the *Aspergillus flavus* spread in stored dry commodities (Simha, 1990).	Aflatoxinas are a group of toxic compounds produced by certain strains of the fungus Aspergillus flavus.	COSAVE,Paraguay ,Chile,Brazil
[31]	6	Technical	The Khapra beetle, *Trogoderma granarium* Everts (Coleoptera: Dermestidae), is a stored product pest of great importance. Its importance lies not only in its capabilities of causing serious damage to stored dry commodities but also in that countries having established populations of this pest face export restrictions for their produce.It is one of the responsibles of the Aspergillus flavus spread in stored dry commodities	Aflatoxinas are a group of toxic compounds produced by certain strains of the fungus Aspergillus flavus	Argentina

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			(Simha, 1990).		
[32]	6	Technical	The Khapra beetle, *Trogoderma granarium* Everts (Coleoptera: Dermestidae), is a stored product pest of great importance. Its importance lies not only in its capabilities of causing serious damage to stored dry commodities but also in that countries having established populations of this pest face export restrictions for their produce. It is one of the insects responsible for the spread of Aspergillus flavus in stored dried commodities (Simha, 1990).	Another relevant information about the pest was added to emphasize its importance. Aflatoxins are a group of toxic compounds produced by certain strains of the fungus Aspergillus flavus.	OIRSA
[33]	7	Editorial	*Trogoderma granarium* is thought to have originated from the Indian subcontinent but it is present in some areas of Asia, the Middle East, Africa and Europe. ~~For more detailed information about *T. granarium*, see the EPPO PQR database (EPPO, 2007).~~ There have been multiple introductions to the United States and Mexico but these were successfully eradicated. It is one of the very few stored products pests that has limited worldwide distribution. *T. granarium* has very limited ability to spread without human aid because it is unable to fly. It is very important to distinguish between records that relate to introductions and those of established infestations.	Move sentence to para 12, containing similar guidance.	EPPO,Russian Federation ,Ukraine ,Morocco ,Uzbekistan
[34]	7	Editorial	*Trogoderma granarium* is thought to have originated from the Indian subcontinent but it is present in some areas of Asia, the Middle East, Africa and Europe. For more detailed information about *T. granarium*, see the EPPO PQR database (EPPO, 2007). ~~There have been multiple introductions to the United States and Mexico but these were successfully eradicated.~~ It is one of the very few stored products pests that has limited worldwide distribution. *T. granarium* has very limited ability to spread without human aid because it is unable to fly. It is very important to distinguish between records that relate to introductions and those of established infestations.	Introduction and eradication happens in other countries so no need to specify USA and Mexico.	Korea, Republic of
[35]	7	Editorial	*Trogoderma granarium* ~~is thought to~~ may have originated from the Indian subcontinent and ~~but~~ it is now present in some areas of Asia, the Middle East, Africa and Europe. For more detailed information about *T. granarium*, see the EPPO PQR database (EPPO, 2007). There have been multiple introductions to the United States and Mexico but these were successfully eradicated. It is one of the very few stored products pests that has limited worldwide distribution. *T. granarium* has very limited ability to spread without human aid because it is unable to fly. It is very important to distinguish between records that relate to introductions and those of established infestations.	For clarification and simplification.	Yemen ,Oman
[36]	7	Editorial	*Trogoderma granarium* is thought to have originated from the Indian subcontinent but it is present in some areas of Asia, the Middle East, Africa	Text moved to paragraph 12, The fourth sentence was relocated for better reading.	Costa Rica ,Nicaragua ,El

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			and Europe. ~~For more detailed information about *T. granarium*, see the EPPO PQR database (EPPO, 2007).~~ It is one of the very few stored product pests that has limited distribution in the world. There have been multiple introductions to the United States and Mexico but these were successfully eradicated~~. It is one of the very few stored products pests that has limited worldwide distribution~~. *T. granarium* has very limited ability to spread without human aid because it is unable to fly. It is very important to distinguish between records that relate to introductions and those of established infestations.		Salvador ,Brazil
[37]	7	Editorial	*Trogoderma granarium* is thought to have originated from the Indian subcontinent but it is present in some areas of Asia, the Middle East, Africa and Europe. ~~For more detailed information about *T. granarium*, see the EPPO PQR database (EPPO, 2007).~~ It is one of the very few stored products pests that has limited worldwide distribution. There have been multiple introductions to the United States and Mexico but these were successfully eradicated. ~~It is one of the very few stored products pests that has limited worldwide distribution~~. *T. granarium* has very limited ability to spread without human aid because it is unable to fly. It is very important to distinguish between records that relate to introductions and those of established infestations.	1) Second sentence moved to paragraph 12. 2) The 4th sentence was relocated for better reading	Uruguay
[38]	7	Editorial	*Trogoderma granarium* is thought to have originated from the Indian subcontinent but it is present in some areas of Asia, the Middle East, Africa and Europe. For more detailed information about *T. granarium*, see the EPPO PQR database (EPPO, 2007). There have been multiple introductions to the United States and Mexico but these were successfully eradicated. It is one of the very few stored products pests that has limited worldwide distribution. *Trogoderma*~~.~~ *granarium* has very limited ability to spread without human aid because it is unable to fly. It is very important to distinguish between records that relate to introductions and those of established infestations.	Genus names are not abbreviated at the beginning of a sentence.	United States of America
[39]	7	Editorial	*Trogoderma granarium* is thought to have originated from the Indian subcontinent but it is present in some areas of Asia, the Middle East, Africa and Europe. ~~For more detailed information about *T. granarium*, see the EPPO PQR database (EPPO, 2007).~~ There have been multiple introductions to the United States and Mexico but these were successfully eradicated. It is one of the very few stored products pests that has limited worldwide distribution. *T. granarium* has very limited ability to spread without human aid because it is unable to fly. It is very important to distinguish between records that relate to introductions and those of established infestations.	Move sentence to para 12, containing similar guidance.	European Union
[40]	7	Editorial	*Trogoderma granarium* is thought to have originated from the Indian	1) Text moved to paragraph 12 2) The 4th	OIRSA

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			subcontinent but it is present in some areas of Asia, the Middle East, Africa and Europe. ~~For more detailed information about *T. granarium*, see the EPPO PQR database (EPPO, 2007).~~ It is one of the very few stored products pests that has limited distribution in the world. There have been multiple introductions to the United States and Mexico but these were successfully eradicated. ~~It is one of the very few stored products pests that has limited worldwide distribution.~~ *T. granarium* has very limited ability to spread without human aid because it is unable to fly. It is very important to distinguish between records that relate to introductions and those of established infestations.	sentence relocated for better reading	
[41]	7	Substantive	*Trogoderma granarium* is thought to have originated from the Indian subcontinent but it is present in some areas of Asia, the Middle East, Africa and Europe. For more detailed information about *T. granarium*, see the EPPO PQR database (EPPO, 2007). ~~There have been multiple introductions to the United States and Mexico but these were successfully eradicated.~~ It is one of the very few stored products pests that has limited worldwide distribution. *T. granarium* has very limited ability to spread without human aid because it is unable to fly. It is very important to distinguish between records that relate to introductions and those of established infestations.	Successful eradications were made in somewhere else too.	Korea, Republic of
[42]	7	Substantive	*Trogoderma granarium* is thought to have originated from the Indian subcontinent but it is present in some areas of Asia, the Middle East, Africa and Europe. For more detailed information about *T. granarium*, see the EPPO PQR database (EPPO, 2007). There have been multiple introductions to the United States ~~and Mexico~~ but these were successfully eradicated. It is one of the very few stored products pests that has limited worldwide distribution. *T. granarium* has very limited ability to spread without human aid because it is unable to fly~~. It is very important to distinguish between records that relate to introductions and those of established infestations.~~	There has been some interceptions in Mexico but the pest was not introduced. Therefore, we suggest a revision of the data base of EPPO. Last sentence deleted because introduction implies establishment so, it is not clear what is intended to distinguish.	Costa Rica ,Nicaragua
[43]	7	Substantive	*Trogoderma granarium* is thought to have originated from the Indian subcontinent but it is present in some areas of Asia, the Middle East, Africa and Europe. For more detailed information about *T. granarium*, see the EPPO PQR database (EPPO, 2007). There have been multiple introductions to the United States ~~and Mexico~~ but these were successfully eradicated. It is one of the very few stored products pests that has limited worldwide distribution. *T. granarium* has very limited ability to spread without human aid because it is unable to fly. ~~It is very important to distinguish between records that relate to introductions and those of established infestations.~~	1) Mexico was deleted because there has been some interceptions in Mexico but the pest was not introduced. Therefore, we suggest a revision of the database of EPPO. 2) Last sentence deleted because introduction implies establishment (ISPM 5) so it is not clear what is intended to distinguish.	Uruguay
[44]	7	Substantive	*Trogoderma granarium* is thought to have originated from the Indian	Information about the regions with the greatest	COSAVE,Parag

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			subcontinent but it is present in some areas of Asia, the Middle East, Africa and Europe. It is one of the very few stored products pests that has limited worldwide distribution.It is found from parallel 35 north to parallel 35 south but occurs mainly in regions near the equator in dry and hot environments (EPPO 1999). ~~For more detailed information about *T. granarium*, see the EPPO PQR database (EPPO, 2007).~~ There have been multiple introductions to the United States and Mexico but these were successfully eradicated. ~~It is one of the very few stored products pests that has limited worldwide distribution.~~ *T. granarium* has very limited ability to spread ~~without human aid because it is unable to fly.~~ so international trade of host commodities is the most important way of spread of this pest. ~~It is very important to distinguish between records that relate to introductions and those of established infestations.~~	risk was added to complete the data. The text "For more detailed information about T. granarium, see the EPPO PQR database (EPPO, 2007)" was moved to para 12. The text "without human aid because it is unable to fly" was changed to highlight that the most important way of pest spread is the international trade of those commodities The text "It is very important to distinguish between records that relate to introductions and those of established infestations" was deleted because introduction implies establishment so it is not clear what is intended to distinguish.	uay ,Argentina ,Chile,Brazil
[45]	7	Substantive	*Trogoderma granarium* is thought to have originated from the Indian subcontinent but it is present in some areas of Asia, the Middle East, Africa and Europe. For more detailed information about *T. granarium*, see the EPPO PQR database (EPPO, 2007). ~~There have been multiple introductions to the United States and Mexico but these were successfully eradicated~~. It is one of the very few stored products pests that has limited worldwide distribution. *T. granarium* has very limited ability to spread without human aid because it is unable to fly. It is very important to distinguish between records that relate to introductions and those of established infestations.	Successful eradications were made in somewhere else too.	Lao People's Democratic Republic,Japan ,India
[46]	7	Substantive	*Trogoderma granarium* is thought to have originated from the Indian subcontinent but it is present in some areas of Asia, the Middle East, Africa and Europe. For more detailed information about *T. granarium*, see the EPPO PQR database (EPPO, 2007). There have been multiple ~~introductions~~interceptions and entries to the United States and Mexico but these were successfully eradicated. It is one of the very few stored products pests that has limited worldwide distribution. *T. granarium* has very limited ability to spread without human aid because it is unable to fly. It is very important to distinguish between records that relate to ~~introductions~~interceptions and entries and those of established infestations.	Introductions do not include the interception records at ports of entry.	United States of America
[47]	7	Substantive	*Trogoderma granarium* is thought to have originated from the Indian subcontinent but it is present in some areas of Asia, the Middle East, Africa and Europe. For more detailed information about *T. granarium*, see the EPPO PQR database (EPPO, 2007). There have been multiple introductions to the United States ~~and Mexico~~ but these were successfully eradicated. It is one of the very few stored products pests that has limited worldwide distribution. *T. granarium* has very limited ability to spread without human aid because it is unable to fly. ~~It is very important to~~	1) Mexico was deleted, there has been some interceptions in Mexico but the pest was not introduced. Therefore, we suggest a revision of the database of EPPO. 2) Last sentence deleted because introduction implies establishment so it is not clear what is intended to distinguish	OIRSA

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			~~distinguish between records that relate to introductions and those of established infestations.~~		
[48]	7	Technical	*Trogoderma granarium* is thought to have originated from the Indian subcontinent but it is present in some areas of Asia, the Middle East, Africa and in a few countries of Europe. For more detailed information about *T. granarium*, see the EPPO PQR database (EPPO, 2007). There have been multiple introductions to the United States and Mexico but these were successfully eradicated. It is one of the very few stored products pests that has limited worldwide distribution. *T. granarium* has very limited ability to spread without human aid because it is unable to fly. It is very important to distinguish between records that relate to introductions interceptions of the pest in important commodities and those of established infestations.	Sentence 1: In Europe its distribution is limited (4 countries, including 3 countries in which its distribution is limited). Last sentence: Proposal to aid clarity. According to ISPM 5 "introduction" means "The entry of a pest resulting in its establishment".	EPPO,Russian Federation ,Ukraine ,Morocco ,Uzbekistan
[49]	7	Technical	*Trogoderma granarium* is thought to have originated from the Indian subcontinent but it is present in some areas of Asia, the Middle East, Africa and Europe. For more detailed information about *T. granarium*, see the EPPO PQR database (EPPO, 2007). There have been multiple introductions to the United States and Mexico but these were successfully eradicated. It is one of the very few stored products pests that has limited worldwide distribution. *T. granarium* has very limited ability to spread without human aid because it is unable to fly, so trade of host commodities is the most important way of spread of this pest. It is very important to distinguish between records that relate to introductions and those of established infestations.	Text was added to highlight that the most important way of pest spread is the trade of those commodities.	Costa Rica ,Nicaragua ,El Salvador ,Brazil
[50]	7	Technical	*Trogoderma granarium* is thought to have originated from the Indian subcontinent but it is present in some areas of Asia, the Middle East, Africa and Europe. For more detailed information about *T. granarium*, see the EPPO PQR database (EPPO, 2007). There have been multiple introductions to the United States and Mexico but these were successfully eradicated. It is one of the very few stored products pests that has limited worldwide distribution. *T. granarium* has very limited ability to spread ~~without human aid because it is unable to fly~~, so trade of host commodities is the most important way of spread of this pest. It is very important to distinguish between records that relate to introductions and those of established infestations.	Text was modified to highlight that the most important way of pest spread is trade of those commodities	Uruguay
[51]	7	Technical	*Trogoderma granarium* is thought to have originated from the Indian subcontinent but it is present in some areas of Asia, the Middle East, Africa and Europe. For more detailed information about *T. granarium*, see the EPPO PQR database (EPPO, 2007). There have been ~~multiple~~ some introductions to the United States ~~and Mexico~~ but these were successfully eradicated. It is one of the very few stored products pests that has limited	Please delete that Mexico has multiple introductions. In the EPPO PQR database (EPPO, 2007) refer that T. granarium was found in Mexico in the past but not established, maybe will be necessary to change introductions by detections because under the definition of ISPM	Mexico

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			worldwide distribution. *T. granarium* has very limited ability to spread without human aid because it is unable to fly. It is very important to distinguish between records that relate to introductions and those of established infestations.	5 in an introduction the entry of a pest result in its establishment and in the case of Mexico T. granarium never be establish. There are some detection but correspond to detections at ports of entry where the consignments never was imported there was under official confinement. To review the EPPO PQR database in order to clarify that in Mexico was only one introduction in the past but not multiple introductions.	
[52]	7	Technical	*Trogoderma granarium* is thought to have originated from the Indian subcontinent but it is present in some areas of Asia, the Middle East, Africa and in a few countries of Europe. For more detailed information about *T. granarium*, see the EPPO PQR database (EPPO, 2007). There have been multiple introductions to the United States and Mexico but these were successfully eradicated. It is one of the very few stored products pests that has limited worldwide distribution. *T. granarium* has very limited ability to spread without human aid because it is unable to fly. It is very important to distinguish between records that relate to ~~introductions~~ interceptions of the pest in important commodities and those of established infestations.	Sentence 1: In Europe its distribution is limited (4 countries, including 3 countries in which its distribution is limited). Last sentence: Proposal to aid clarity. According to ISPM 5 "introduction" means "The entry of a pest resulting in its establishment".	European Union
[53]	7	Technical	*Trogoderma granarium* is thought to have originated from the Indian subcontinent but it is present in some areas of Asia, the Middle East, Africa and Europe. For more detailed information about *T. granarium*, see the EPPO PQR database (EPPO, 2007). There have been multiple introductions to the United States and Mexico but these were successfully eradicated. It is one of the very few stored products pests that has limited worldwide distribution. *T. granarium* has very limited ability to spread without human aid because it is unable to fly, so trade of host commodities is the most important way of spread of this pest. It is very important to distinguish between records that relate to introductions and those of established infestations.	Text was added to highlight that the most important way of pest spread is trade of those commodities	OIRSA
[54]	8	Editorial	The ~~K~~khapra beetle may occur in various dry stored product~~s~~ of primarily vegetable origin. Primary hosts are cereals, buckwheat, cereal products, pulses, alfalfa, various vegetable seeds, herbs, spices and various nuts. It can successfully complete its life cycle in copra, dried fruits, various gums and many different dried products ~~of~~ wholly or partially of animal origin such as milk powder, skins, dried dog food, dried blood, dead insects and dried animal carcasses. As a pest it is most prevalent under hot dry conditions where very heavy infestations can develop. In cooler and also in hot and humid conditions it tends to be out-competed as a pest by other species such as *Sitophilus* spp. and *Rhyzopertha dominica* (Fabricius). Commodities stored in bags in traditional warehouses are more at risk from	See 'general comments' for khapra beetle comment. Grammar for 'stored product' and 'partially of'.	United States of America

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			this pest than bulk-stored commodities.		
[55]	8	Editorial	The Khapra beetle ~~may~~ occurs in various dry stored products of primarily vegetable origin. Primary hosts are cereals, buckwheat, cereal products, pulses, alfalfa, various vegetable seeds, herbs, spices and various nuts. It can also successfully complete its life cycle in copra, dried fruits, various gums and many different dried products of wholly or partial animal origin such as milk powder, skins, dried dog food, dried blood, dead insects and dried animal carcasses. As a pest it is most prevalent under hot dry conditions where very heavy infestations can develop. In cooler and also in hot and humid conditions it tends to be out-competed as a pest by other species such as *Sitophilus* spp. and *Rhyzopertha dominica* (Fabricius). Commodities stored in bags in traditional warehouses are more at risk from this pest than commodities that are stored in bulk ~~stored commodities.~~	Suggested changes make the text more readable and refer to language that is more commonly used and understood.	Canada
[56]	8	Technical	The Khapra beetle may occur in various dry stored products of primarily vegetable origin. Primary hosts are cereals, buckwheat, cereal products, pulses, dehydrated alfalfa, various vegetable seeds, herbs, spices and various nuts. It can successfully complete its life cycle in copra, dried fruits, various gums and many different dried products of wholly or partial animal origin such as milk powder, skins, dried dog food, dried blood, dead insects and dried animal carcasses. As a pest it is most prevalent under hot dry conditions where very heavy infestations can develop. In cooler and also in hot and humid conditions it tends to be out-competed as a pest by other species such as *Sitophilus* spp. and *Rhyzopertha dominica* (Fabricius). Commodities stored in bags in traditional warehouses are more at risk from this pest than bulk-stored commodities.	We recognize that Khapra beetle is associated with acicalada or achicalada alfalfa which is finally dehydrated alfalfa (transported in bales). Also feed on dry herb (herbal mainly). Mexico believe that even though it is repetitive, it must be used the correct terms because it seems like khapra beetle is associated with alfalfa or fresh grass.	Mexico
[57]	8	Technical	The Khapra beetle may occur in various dry stored products of primarily plant ~~vegetable~~ origin. Primary hosts are cereals, buckwheat, cereal products, pulses, alfalfa, various vegetable seeds, herbs, spices and various nuts. It can successfully complete its life cycle in copra, dried fruits, various gums and many different dried products of wholly or partial animal origin such as milk powder, skins, dried dog food, dried blood, dead insects and dried animal carcasses. As a pest it is most prevalent under hot dry conditions where very heavy infestations can develop. In cooler and also in hot and humid conditions it tends to be out-competed as a pest by other species such as *Sitophilus* spp. and *Rhyzopertha dominica* (Fabricius). Commodities stored in bags in traditional warehouses are more at risk from this pest than bulk-stored commodities.	The term "vegetable" has a specific botanical meaning which is not what is intended in this context. The term "plant " is therefore more appropriate within the context of this IPPC diagnostic protocol.	Canada
[58]	10	Editorial	Khapra beetle can have between one and more than ten generations per year depending on food availability and quality, temperature and humidity. A complete life cycle may be as short as 26 days (temperature 32–35 °C) or as long as 220 days or more in a suboptimal environment. In temperate	To highlight because fumigation could be done with different phytosanitary products.	Costa Rica ,Mexico ,Nicaragua ,El Salvador ,Brazil

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			climates larvae become inactive at temperatures below 5 °C, so the pest is able to survive and breed only in protected environments. However, there are two genetic variations of larvae: those that are able to undergo facultative diapause and those that are unable to do so. Larvae of the first type are stimulated into diapause by adverse conditions such as low or high temperatures and/or lack of food. During diapause their respiration drops to an extremely low level leading to tolerance to any type of fumigation. Diapausing larvae are also cold-hardy and may survive temperatures below −10 °C. Should favourable conditions return, the pest is able to multiply rapidly and cause serious damage to the commodity.		
[59]	10	Editorial	Khapra beetle can have between one and more than ten generations per year depending on food availability and quality, temperature and humidity. A complete life cycle may be as short as 26 days (temperature 32–35 °C) or as long as 220 days or more in a suboptimal environment. In temperate climates larvae become inactive at temperatures below 5 °C, so the pest is able to survive and breed only in protected environments. However, there are two genetic variations of larvae: those that are able to undergo facultative diapause and those that are unable to do so. Larvae of the first type are stimulated into diapause by adverse conditions such as low or high temperatures and/or lack of food. During diapause their respiration drops to an extremely low level leading to tolerance to any kind of fumigation. Diapausing larvae are also cold-hardy and may survive temperatures below −10 °C. Should favourable conditions return, the pest is able to multiply rapidly and cause serious damage to the commodity.	Text added to highlight that fumigation could be done with different phytosanitary products	Uruguay
[60]	10	Editorial	Khapra beetle can have between one and more than ten generations per year depending on food availability and quality, temperature and humidity. A complete life cycle may be as short as 26 days (temperature 32–35 °C) or as long as 220 days or more in a suboptimal environment. In temperate climates larvae become inactive at temperatures below 5 °C, so the pest is able to survive and breed only in protected environments. However, there are two genetic variations of larvae: those that are able to undergo facultative diapause and those that are unable to do so. Larvae of the first type are stimulated into diapause by adverse conditions such as low or high temperatures and/or lack of food. During diapause their respiration drops to an extremely low level leading to tolerance to any kind of fumigation. Diapausing larvae are also cold-hardy and may survive temperatures below −10 °C. Should favourable conditions return, the pest is able to multiply rapidly and cause serious damage to the commodity.	To highlight because fumigation could be done with different phytosanitary products.	COSAVE,Paraguay ,Chile
[61]	10	Editorial	Khapra beetle can have between one and more than ten generations per year depending on food availability and quality, temperature and humidity. A complete life cycle may be as short as 26 days (temperature 32–35 °C)	Such factual material was likely taken from one or more references, and these should be cited.	United States of America

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			or as long as 220 days or more in a suboptimal environment. In temperate climates larvae become inactive at temperatures below 5 °C, so the pest is able to survive and breed only in protected environments. However, there are two genetic variations of larvae: those that are able to undergo facultative diapause and those that are unable to do so. Larvae of the first type are stimulated into diapause by adverse conditions such as low or high temperatures and/or lack of food. During diapause their respiration drops to an extremely low level leading to tolerance to fumigation. Diapausing larvae are also cold-hardy and may survive temperatures below −10 °C. Should favourable conditions return, the pest is able to multiply rapidly and cause serious damage to the commodity.REFERENCE(S)?		
[62]	10	Editorial	Khapra beetle can have between one and more than ten generations per year depending on food availability and quality, temperature and humidity. A complete life cycle may be as short as 26 days (temperature 32–35 °C) or as long as 220 days or more in a suboptimal environment. In temperate climates larvae become inactive at temperatures below 5 °C, so the pest is able to survive and breed only in protected environments. However, there are two genetic variations of larvae: those that are able to undergo facultative diapause and those that are unable to do so. Larvae of the first type are stimulated into diapause by adverse conditions such as low or high temperatures and/or lack of food. During diapause their respiration drops to an extremely low level leading to tolerance any kind of to fumigation. Diapausing larvae are also cold-hardy and may survive temperatures below −10 °C. Should favourable conditions return, the pest is able to multiply rapidly and cause serious damage to the commodity.	To highlight because fumigation could be done with different phytosanitary products	Argentina
[63]	10	Editorial	Khapra beetle can have between one and more than ten generations per year depending on food availability and quality, temperature and humidity. A complete life cycle may be as short as 26 days (temperature 32–35 °C) or as long as 220 days or more in a suboptimal environment. In temperate climates larvae become inactive at temperatures below 5 °C, so the pest is able to survive and breed only in protected environments. Like many other species of beetles~~However,~~ there are two genetic variations of larvae: those that are able to undergo facultative diapause and those that are unable to do so. Larvae of the first type are stimulated into diapause by adverse conditions such as low or high temperatures and/or lack of food. During diapause their respiration drops to an extremely low level leading to tolerance to fumigation. Diapausing larvae are also cold-hardy and may survive temperatures below −10 °C. Should favourable conditions return, the pest is able to multiply rapidly and cause serious damage to the commodity.	Suggested changes make the paragraph gramatically correct.	Canada
[64]	10	Editorial	Khapra beetle can have between one and more than ten generations per	To highlight because fumigation could be done	OIRSA

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			year depending on food availability and quality, temperature and humidity. A complete life cycle may be as short as 26 days (temperature 32–35 °C) or as long as 220 days or more in a suboptimal environment. In temperate climates larvae become inactive at temperatures below 5 °C, so the pest is able to survive and breed only in protected environments. However, there are two genetic variations of larvae: those that are able to undergo facultative diapause and those that are unable to do so. Larvae of the first type are stimulated into diapause by adverse conditions such as low or high temperatures and/or lack of food. During diapause their respiration drops to an extremely low level leading to tolerance to any type of fumigation. Diapausing larvae are also cold-hardy and may survive temperatures below −10 °C. Should favourable conditions return, the pest is able to multiply rapidly and cause serious damage to the commodity.	with different phytosanitary products	
[65]	10	Technical	Khapra beetle can have between one and more than ten generations per year depending on food availability and quality, temperature and humidity. A complete life cycle may be as short as 26 days (temperature 32–35 °C) or as long as 220 days or more in a suboptimal environment. In temperate climates larvae become inactive at temperatures below 5 °C, so the pest is able to survive and breed only in protected environments. However, there are two genetic variations of larvae: those that are able to undergo facultative diapause and those that are unable to do so. Larvae of the first type are stimulated into diapause by adverse conditions such as low or high temperatures and/or lack of food. During diapause their respiration drops to an extremely low level leading to tolerance to fumigation. Diapausing larvae are also cold-hardy and may survive temperatures below −10 °C. Should favourable conditions return, the pest is able to multiply rapidly and cause serious damage to the commodity.	Providing the key references for this important paragraph will be useful	New Zealand
[66]	11	Editorial	*Trogoderma* species other than ~~K~~khapra beetle may also be found in stored products, but only some of these feed on such products. Among these species the biggest economic losses are caused by *T. variabile* Ballion, which is recognized as a quarantine pest in some countries. However, most *Trogoderma* species occurring in stored products appear to be scavengers, feeding on dead bodies of other insects. During a 12-year survey conducted in California, eight species of *Trogoderma* were found in stored seeds, animal feed and grocery commodities (Strong and Okumura, 1966). Mordkovich and Sokolov (1999) mention other *Trogoderma* species that may be found in stored products. Among them, *T. longisetosum* Chao et Lee has been noted as a stored product pest in China. It is very similar to *T. glabrum* (Herbst). Some tropical *Trogoderma* species may also be present in stored products (Delobel and Tran, 1993). One ~~of~~ such species is *T. cavum*, which was described by Beal (1982) after examination of specimens infesting stored rice in Bolivia. Some species occurring in	Khapra beetle: See General Comments. "One of such species": Grammar.	United States of America

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			stored products closely resemble *T. granarium*.		
[67]	11	Editorial	*Trogoderma* species other than Khapra beetle may also be found in stored products, but only some of these feed on such products. Among these species ~~the biggest economic losses are caused by.~~ *T. variabile* Ballion, may cause significant economic damage and ~~which~~ is recognized as a quarantine pest in some countries. However, most *Trogoderma* species occurring in stored products appear to be scavengers, feeding on dead bodies of other insects. During a 12-year survey conducted in California, eight species of *Trogoderma* were found in stored seeds, animal feed and grocery commodities (Strong and Okumura, 1966). Mordkovich and Sokolov (1999) mention other *Trogoderma* species that may be found in stored products. Among them, *T. longisetosum* Chao et Lee has been noted as a stored product pest in China. It is very similar to *T. glabrum* (Herbst). Some tropical *Trogoderma* species may also be present in stored products (Delobel and Tran, 1993). One of such species is *T. cavum*, which was described by Beal (1982) after examination of specimens infesting stored rice in Bolivia. Some species occurring in stored products closely resemble *T. granarium*.	Suggested changes add clarity to the text.	Canada
[68]	11	Technical	*Trogoderma* species other than Khapra beetle may also be found in stored products, but only some of these feed on such products. Among these species the biggest economic losses are caused by *T. variabile* Ballion, which is recognized as a quarantine pest in some countries. However, most *Trogoderma* species occurring in stored products appear to be scavengers, feeding on dead bodies of other insects. During a 12-year survey conducted in California, eight species of *Trogoderma* were found in stored seeds, animal feed and grocery commodities (Strong and Okumura, 1966). Mordkovich and Sokolov (1999) mention other *Trogoderma* species that may be found in stored products. Among them, *T. longisetosum* Chao et Lee has been noted as a stored product pest in China. It is very similar to *T. glabrum* (Herbst). Some tropical *Trogoderma* species may also be present in stored products (Delobel and Tran, 1993). One of such species is *T. cavum*, which was described by Beal (1982) after examination of specimens infesting stored rice in Bolivia. Some species occurring in stored products closely resemble *T. granarium*.	The reference to "Chao et Lee" in the 6th sentence does not include a year. Year should be included.	Canada
[69]	12	Editorial	For more general information on *T. granarium*, see Hinton (1945), Lindgren *et al*. (1955), Pasek (1998), EPPO/CABI (1997), Berg (1999a), CABI (2005), EPPO (2007) and Walker (2008).	Reference moved from paragraph 7.	Costa Rica ,Uruguay ,Nicaragua ,El Salvador
[70]	12	Editorial	For more general information on *T. granarium*, see the EPPO PQR database (EPPO, 2011). as well as ~~see~~ Hinton (1945), Lindgren *et al*. (1955), Pasek (1998), EPPO/CABI (1997), Berg (1999a), CABI (20~~05~~11)	Sentence moved from para 7. (Note: the new version of the EPPO PQR database is due to be published in 2011.) The latest version of the	EPPO,Russian Federation ,Ukraine

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			and Walker (2008).	CABI Crop Protection Compendium was published in 2011.	,Morocco ,Uzbekistan
[71]	12	Editorial	For more general information on *T. granarium*, see Hinton (1945), Lindgren *et al*. (1955), Pasek (1998), EPPO/CABI (1997), Berg (1999a), CABI (2005), EPPO (2007) and Walker (2008).		COSAVE,Paraguay ,Chile,Brazil
[72]	12	Editorial	For more general information on *T. granarium*, see Hinton (1945), Lindgren *et al*. (1955), ~~Pasek (1998),~~ EPPO/CABI (1997), Pasek (1998), Berg (1999a), CABI (2005) and Walker (2008).	Arranging multiple citations chronologically.	United States of America
[73]	12	Editorial	For more general information on *T. granarium*, see Hinton (1945), Lindgren *et al*. (1955), Pasek (1998), EPPO/CABI (1997), Berg (1999a), CABI (2005) EPPO (2007) Reference moved from para 7Reference moved from para 7and Walker (2008).	Reference moved from para 7	Argentina
[74]	12	Editorial	For more general information on *T. granarium*, see the EPPO PQR database (EPPO, 2011). as well as ~~see~~ Hinton (1945), Lindgren *et al*. (1955), Pasek (1998), EPPO/CABI (1997), Berg (1999a), CABI (20~~05~~11) and Walker (2008).	Sentence moved from para 7. The information can also be found in the latest version of the EPPO PQR database published in 2011. The latest version of the CABI Crop Protection Compendium was published in 2011.	European Union
[75]	12	Editorial	For more general information on *T. granarium*, see Hinton (1945), Lindgren *et al*. (1955), Pasek (1998), EPPO/CABI (1997), Berg (1999a), CABI (2005), EPPO (2007) and Walker (2008).	Reference moved from paragraph 7	OIRSA
[76]	17	Editorial	~~*Trogoderma quinquefasciata*~~ *Trogoderma quinquefasciata* Leesberg, 1906	Remove underlining of name	EPPO,Russian Federation ,Ukraine ,Morocco ,Uzbekistan
[77]	17	Editorial	~~*Trogoderma quinquefasciata*~~ Leesberg, 1906 *Trogoderma quinquefasciata* Leesberg, 1906	Normal procedure for citing scientific names, with scientific name in italics but no underline and authority and year without italics and underline .	United States of America
[78]	17	Editorial	~~*Trogoderma quinquefasciata*~~ *Trogoderma quinquefasciata* Leesberg, 1906	Remove underlining of name	European Union
[79]	17	Editorial	*Trogoderma quinquefasciata* Leesberg, 1906	Delete underline as the species name is already written in italics. Refer to paragraph 18 and 19.	South Africa
[80]	19	Editorial	*Trogoderma afrum* Pries~~t~~ner, 1951	A misspelt name	EPPO,Russian Federation ,European Union ,Ukraine

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
					,Morocco ,Uzbekistan
[81]	19	Editorial	*Trogoderma afrum* Priest~~t~~ner, 1951	Correct the author's name (see Have, 2003)	Japan
[82]	20	Technical	*Trogoderma granarium* ssp. *afrum* Attia & Kamel, 1965; *Trogoderma koningsbergeri* Pic, 1933	The synonym should be added.	EPPO,Russian Federation ,European Union ,Ukraine ,Morocco ,Uzbekistan
[83]	20	Technical	*Trogoderma granarium* ssp. *afrum* Attia & Kamel, 1965 *Trogoderma koningsbergeri* Pic, 1993	See Have(2007). New nomenclatorial changes in the family Dermestidae (Coleoptera). Acta Entomologica Slovenica 15(1):69-74	Japan
[84]	21	Editorial	**Common names:** ~~K~~khapra beetle (English)	See general comments.	United States of America
[85]	22	Editorial	Trogoderme (dermeste) du grain, Dermeste des ~~G~~grains (French)	No capital letter for the word "grain".	EPPO,Russian Federation ,European Union ,Ukraine ,Morocco ,Uzbekistan
[86]	22	Editorial	Trogoderme (dermeste) du grain, ~~D~~dermeste des ~~G~~grains (French)	No capitalization required: Larouse Unabridged French Dictionary (1998)	United States of America
[87]	23	Editorial	*Trogoderma* de los granos, ~~E~~escarabajo ~~K~~khapra, ~~G~~gorgojo khapra (Spanish)	No capitalization required; Simon and Schuster's International Spanish Dictionary, 2nd ed. (1997).	United States of America
[88]	27	Editorial	~~T.~~ *Trogoderma granarium* has the following life developmental stages: eggs on the surface of grain and other stored products; larvae (5–11 instars) in stored products (larvae may be found in packing material or within storage structures); pupae in stored products, in the last larval exuviae (cast skins); adults in stored products.	Genus names are not abbreviated at the beginning of a sentence.	United States of America
[89]	28	Editorial	Methods to detect *T. granarium* infestations include inspection (physical and visual search), ~~and~~ use of food baits ~~or, more importantly,~~and pheromone traps. Often the infested material contains only larvae. There are three reasons for this: (1) adult longevity is usually between 12 and 25 days, but can be as long as 147 days in unfavourable conditions, whereas larval longevity is usually 19–190 days (and can be up to six years should larvae go into diapause); (2) most of the dermestid larvae occurring in the stored product will partially or wholly consume dead adults; and (3) adults are most prevalent when conditions are favourable for population growth.	Suggested changes in the first and last sentence make the text more easily readable.	Canada

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			Larval exuviae are usually not consumed so their presence is a clear indication of a possible active infestation. Larvae are extremely cryptic by nature. This is particularly so in the case of diapausing larvae, that may~~which can~~ stay inactive for periods in cracks and crevices where they are very difficult or nearly impossible to locate.		
[90]	28	Substantive	Methods to detect *T. granarium* infestations include inspection ~~(physical and visual search)~~ and use of food baits or, more importantly, pheromone traps. Often the infested material contains only larvae. There are three reasons for this: (1) adult longevity is usually between 12 and 25 days, but can be as long as 147 days in unfavourable conditions, whereas larval longevity is usually 19–190 days (and can be up to six years should larvae go into diapause); (2) most of the dermestid larvae occurring in the stored product will partially or wholly consume dead adults; and (3) adults are most prevalent when conditions are favourable for population growth. Larval exuviae are usually not consumed so their presence is a clear indication of a possible active infestation. Larvae are extremely cryptic by nature. This is particularly so in the case of diapausing larvae, which can stay inactive for periods in cracks and crevices where they are very difficult or nearly impossible to locate.	It is not necessary to include physical and visual because the definition of inspection includes visual examination.	South Africa
[91]	28	Technical	Methods to detect *T. granarium* infestations include inspection (physical and visual examination ~~search~~) and use of food baits or, more importantly, pheromone traps. Often the infested material contains only larvae. There are three reasons for this: (1) adult longevity is usually between 12 and 25 days, but can be as long as 147 days in unfavourable conditions, whereas larval longevity is usually 19–190 days (and can be up to six years should larvae go into diapause); (2) most of the dermestid larvae occurring in the stored product will partially or wholly consume dead adults; and (3) adults are most prevalent when conditions are favourable for population growth. Larval exuviae are usually not consumed so their presence is a clear indication of a possible active infestation. Larvae are extremely cryptic by nature. This is particularly so in the case of diapausing larvae, which can stay inactive for periods in cracks and crevices where they are very difficult or nearly impossible to locate.	To be consistent with ISPM 5 (inspection is a visual examination)	Costa Rica ,Uruguay ,Nicaragua ,El Salvador
[92]	28	Technical	Methods to detect *T. granarium* infestations include inspection (physical and visual examination ~~search~~) and use of food baits or, more importantly, pheromone traps. Often the infested material contains only larvae. There are three reasons for this: (1) adult longevity is usually between 12 and 25 days, but can be as long as 147 days in unfavourable conditions, whereas larval longevity is usually 19–190 days (and can be up to six years should larvae go into diapause); (2) most of the dermestid larvae occurring in the stored product will partially or wholly consume dead adults; and (3) adults are most prevalent when conditions are favourable for population growth.	To be consistent with ISPM 5 (inspection is a visual examination)	COSAVE,Paraguay ,Chile,Brazil

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			Larval exuviae are usually not consumed so their presence is a clear indication of a possible active infestation. Larvae are extremely cryptic by nature. This is particularly so in the case of diapausing larvae, which can stay inactive for periods in cracks and crevices where they are very difficult or nearly impossible to locate.		
[93]	28	Technical	Methods to detect *T. granarium* infestations include inspection (physical and visual ~~search~~ examination) and use of food baits or, more importantly, pheromone traps. Often the infested material contains only larvae. There are three reasons for this: (1) adult longevity is usually between 12 and 25 days, but can be as long as 147 days in unfavourable conditions, whereas larval longevity is usually 19–190 days (and can be up to six years should larvae go into diapause); (2) most of the dermestid larvae occurring in the stored product will partially or wholly consume dead adults; and (3) adults are most prevalent when conditions are favourable for population growth. Larval exuviae are usually not consumed so their presence is a clear indication of a possible active infestation. Larvae are extremely cryptic by nature. This is particularly so in the case of diapausing larvae, which can stay inactive for periods in cracks and crevices where they are very difficult or nearly impossible to locate.	As is defined visual examination in ISPM 5.	Mexico
[94]	28	Technical	Methods to detect *T. granarium* infestations include inspection (physical and visual examination ~~search~~) and use of food baits or, more importantly, pheromone traps. Often the infested material contains only larvae. There are three reasons for this: (1) adult longevity is usually between 12 and 25 days, but can be as long as 147 days in unfavourable conditions, whereas larval longevity is usually 19–190 days (and can be up to six years should larvae go into diapause); (2) most of the dermestid larvae occurring in the stored product will partially or wholly consume dead adults; and (3) adults are most prevalent when conditions are favourable for population growth. Larval exuviae are usually not consumed so their presence is a clear indication of a possible active infestation. Larvae are extremely cryptic by nature. This is particularly so in the case of diapausing larvae, which can stay inactive for periods in cracks and crevices where they are very difficult or nearly impossible to locate.	To be consistent with ISPM 5 (inspection is a visual examination)	Argentina
[95]	28	Technical	Methods to detect *T. granarium* infestations include inspection (physical and visual examination~~search~~) and use of food baits or, more importantly, pheromone traps. Often the infested material contains only larvae. There are three reasons for this: (1) adult longevity is usually between 12 and 25 days, but can be as long as 147 days in unfavourable conditions, whereas larval longevity is usually 19–190 days (and can be up to six years should larvae go into diapause); (2) most of the dermestid larvae occurring in the stored product will partially or wholly consume dead adults; and (3) adults are most prevalent when conditions are favourable for population growth.	To be consistent with ISPM 5 (inspection is a visual examination)	OIRSA

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			Larval exuviae are usually not consumed so their presence is a clear indication of a possible active infestation. Larvae are extremely cryptic by nature. This is particularly so in the case of diapausing larvae, which can stay inactive for periods in cracks and crevices where they are very difficult or nearly impossible to locate.		
[96]	29	Editorial	It should be mentioned that many other dermestid species belonging to other genera occur in stored products. Members of *Dermestes* and *Attagenus* genera are frequently found feeding on materials of animal origin, such as dog biscuits, dried meat, dried blood, as well as rat, mice and bird carcasses (also in the stores). *Anthrenus* and *Anthrenocerus* species can be serious pests of wool and woollen products. In stored products heavily infested by other stored products pests, non-pest *Trogoderma* will usually feed on these dead insects.	The last sentence may be modified as below: In stored products heavily infested with other stored product pests, the non-pest Trogoderma are usually found feeding on dead insects.	New Zealand
[97]	29	Editorial	It should be mentioned that many other dermestid species belonging to other genera occur in stored products. Members of *Dermestes* and *Attagenus* ~~genera~~ are frequently found feeding on materials of animal origin, such as dog biscuits, dried meat, and dried blood~~,~~ They also feed on ~~as well as~~ rat, mice and bird carcasses ~~(also in the stores).~~ found in stored products. *Anthrenus* and *Anthrenocerus* species can be serious pests of wool and woollen products. In stored products heavily infested by other stored product~~s~~ pests, non-pest *Trogoderma* will usually feed on carcasses of these ~~dead insects~~pests.	Attempt to improve comprehension without changing ideas. Use of 'genera' in second sentence is redundant, for 'genera' mentioned in first sentence.	United States of America
[98]	29	Editorial	Other than the Trogodermids, m~~It should be mentioned that many other dermestid species belonging to other genera occur in stored products. M~~embers of *Dermestes* and *Attagenus* genera are frequently found feeding on materials of animal origin, such as dog biscuits, dried meat, dried blood, as well as rat, mice and bird carcasses (also in the stores). *Anthrenus* and *Anthrenocerus* species can be serious pests of wool and woollen products. In stored products heavily infested by other stored products pests, non-pest *Trogoderma* will usually feed on these dead insects.	The suggested edits to the first sentence add clarity to the text.	Canada
[99]	30	Editorial	Searches for this pest are particularly difficult in cases of low-level infestations. In contrast to most other stored products pests, the Khapra beetle prefers hot and dry areas. The larvae of *Trogoderma* species are very crepuscular, and populations can persist in small quantities of residues that may occur within a structure or mode of transport. Larvae in diapause can survive long periods without food. For diapausing larvae it is important to examine ~~search~~ under piles of dirt, flaking paint and rust, in empty packaging materials such as hessian bags, tarpaulins, and corrugated cardboard. Larvae are often hiding behind wall panelling, under internal lining, between floorboards, under insulation, on dry ledges, electrical cable trays and conduits, switch boxes etc. Larval exuviae	Better wording.	Costa Rica ,Uruguay ,Nicaragua ,OIRSA,Brazil

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			become airborne very easily, and therefore it is always important to check window sills, grilles of venting holes and spider webs for their presence. Rodent traps containing baits should be always inspected.		
[100]	30	Editorial	Searches for this pest are particularly difficult in cases of low-level infestations. In contrast to most other stored products pests, the Khapra beetle prefers hot and dry areas. The larvae of *Trogoderma* species are very crepuscular, and populations can persist in small quantities of residues that may occur within a structure or mode of transport. Larvae in diapause can survive long periods without food. For diapausing larvae it is important to ~~search~~ examine under piles of dirt, flaking paint and rust, in empty packaging materials such as hessian bags, tarpaulins, and corrugated cardboard. Larvae are often hiding behind wall panelling, under internal lining, between floorboards, under insulation, on dry ledges, electrical cable trays and conduits, switch boxes etc. Larval exuviae become airborne very easily, and therefore it is always important to check window sills, grilles of venting holes and spider webs for their presence. Rodent traps containing baits should be always inspected.	Better wording.	COSAVE,Paraguay ,Chile
[101]	30	Editorial	Searches for this pest are particularly difficult in cases of low-level infestations. In contrast to most other stored product~~s~~ pests, the ~~K~~khapra beetle prefers hot, ~~and~~ dry areas. The larvae of *Trogoderma* species are ~~very crepuscular~~most active at dawn and dusk.~~, and p~~Populations can persist in small quantities of residues that may occur within a structure or mode of transport. Larvae in diapause can survive long periods without food. For diapausing larvae it is important to search under piles of dirt, flaking paint and rust, and in empty packaging materials such as hessian bags, tarpaulins, and corrugated cardboard. Larvae are often hiding behind wall panelling, under internal lining, between floorboards, under insulation, and on dry ledges, electrical cable trays and conduits, switch boxes, etc. Larval exuviae become airborne very easily, and therefore it is always important to check window sills, grilles of venting holes and spider webs for their presence. Rodent traps containing baits should ~~be~~ always be inspected.	Clarity and avoidance of technical, potentially confusing term.	United States of America
[102]	30	Editorial	Searches for this pest are particularly difficult in cases of low-level infestations. In contrast to most other stored products pests, the Khapra beetle prefers hot and dry areas. The larvae of *Trogoderma* species are very crepuscular, and populations can persist in small quantities of residues that may occur within a structure or mode of transport. Larvae in diapause can survive long periods without food. For diapausing larvae it is important to ~~search~~ examine under piles of dirt, flaking paint and rust, in empty packaging materials such as hessian bags, tarpaulins, and corrugated cardboard. Larvae are often hiding behind wall panelling, under internal lining, between floorboards, under insulation, on dry ledges,	Better wording.	Argentina

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			electrical cable trays and conduits, switch boxes etc. Larval exuviae become airborne very easily, and therefore it is always important to check window sills, grilles of venting holes and spider webs for their presence. Rodent traps containing baits should be always inspected.		
[103]	30	Substantive	~~Searches for this pest are particularly difficult in cases of low-level infestations. In contrast to most other stored products pests, the Khapra beetle prefers hot and dry areas. The larvae of *Trogoderma* species are very crepuscular, and populations can persist in small quantities of residues that may occur within a structure or mode of transport. Larvae in diapause can survive long periods without food. For diapausing larvae it is important to search under piles of dirt, flaking paint and rust, in empty packaging materials such as hessian bags, tarpaulins, and corrugated cardboard. Larvae are often hiding behind wall panelling, under internal lining, between floorboards, under insulation, on dry ledges, electrical cable trays and conduits, switch boxes etc. Larval exuviae become airborne very easily, and therefore it is always important to check window sills, grilles of venting holes and spider webs for their presence. Rodent traps containing baits should be always inspected.~~	Move this paragraph to after paragraph 32. More logical order	EPPO,Russian Federation ,Ukraine ,Morocco ,Uzbekistan
[104]	30	Substantive	~~Searches for this pest are particularly difficult in cases of low-level infestations. In contrast to most other stored products pests, the Khapra beetle prefers hot and dry areas. The larvae of *Trogoderma* species are very crepuscular, and populations can persist in small quantities of residues that may occur within a structure or mode of transport. Larvae in diapause can survive long periods without food. For diapausing larvae it is important to search under piles of dirt, flaking paint and rust, in empty packaging materials such as hessian bags, tarpaulins, and corrugated cardboard. Larvae are often hiding behind wall panelling, under internal lining, between floorboards, under insulation, on dry ledges, electrical cable trays and conduits, switch boxes etc. Larval exuviae become airborne very easily, and therefore it is always important to check window sills, grilles of venting holes and spider webs for their presence. Rodent traps containing baits should be always inspected.~~	Move this paragraph to after paragraph 32. More logical order	European Union
[105]	30	Technical	~~Searches~~ Inspections for this pest are particularly difficult in cases of low-level infestations. In contrast to most other stored products pests, the Khapra beetle prefers hot and dry areas. The larvae of *Trogoderma* species are very crepuscular, and populations can persist in small quantities of residues that may occur within a structure or mode of transport. Larvae in diapause can survive long periods without food. For diapausing larvae it is important to search under piles of dirt, flaking paint and rust, in empty packaging materials such as hessian bags, tarpaulins, and corrugated cardboard. Larvae are often hiding behind wall panelling, under internal lining, between floorboards, under insulation, on dry ledges,	To be consistent with ISPM 5 (inspection is a visual examination)	Costa Rica ,Uruguay ,Mexico ,Nicaragua

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			electrical cable trays and conduits, switch boxes etc. Larval exuviae become airborne very easily, and therefore it is always important to check window sills, grilles of venting holes and spider webs for their presence. Rodent traps containing baits should be always inspected.		
[106]	30	Technical	~~Searches~~ Inspections for this pest are particularly difficult in cases of low-level infestations. In contrast to most other stored products pests, the Khapra beetle prefers hot and dry areas. The larvae of *Trogoderma* species are very crepuscular, and populations can persist in small quantities of residues that may occur within a structure or mode of transport. Larvae in diapause can survive long periods without food. For diapausing larvae it is important to search under piles of dirt, flaking paint and rust, in empty packaging materials such as hessian bags, tarpaulins, and corrugated cardboard. Larvae are often hiding behind wall panelling, under internal lining, between floorboards, under insulation, on dry ledges, electrical cable trays and conduits, switch boxes etc. Larval exuviae become airborne very easily, and therefore it is always important to check window sills, grilles of venting holes and spider webs for their presence. Rodent traps containing baits should be always inspected.	To be consistent with ISPM 5 (inspection is a visual examination)	COSAVE,Paraguay ,Chile,Brazil
[107]	30	Technical	~~Searches~~ Inspections for this pest are particularly difficult in cases of low-level infestations. In contrast to most other stored products pests, the Khapra beetle prefers hot and dry areas. The larvae of *Trogoderma* species are very crepuscular, and populations can persist in small quantities of residues that may occur within a structure or mode of transport. Larvae in diapause can survive long periods without food. For diapausing larvae it is important to search under piles of dirt, flaking paint and rust, in empty packaging materials such as hessian bags, tarpaulins, and corrugated cardboard. Larvae are often hiding behind wall panelling, under internal lining, between floorboards, under insulation, on dry ledges, electrical cable trays and conduits, switch boxes etc. Larval exuviae become airborne very easily, and therefore it is always important to check window sills, grilles of venting holes and spider webs for their presence. Rodent traps containing baits should be always inspected.	To be consistent with ISPM 5 (inspection is a visual examination)	Argentina
[108]	30	Technical	Searches for this pest are particularly difficult in cases of low-level infestations. ~~In contrast to most other stored products pests, the Khapra beetle prefers hot and dry areas.~~ The larvae of *Trogoderma* species are very crepuscular, and populations can persist in small quantities of residues that may occur within a structure or mode of transport. Larvae in diapause can survive long periods without food. For diapausing larvae it is important to search under piles of dirt, flaking paint and rust, in empty packaging materials such as hessian bags, tarpaulins, and corrugated cardboard. Larvae are often hiding behind wall panelling, under internal lining, between floorboards, under insulation, on dry ledges, electrical	Delete the sencond sentence. This sentence is not required as this pest, even though it originates from a hot and dry environment, can still survive in a variety of different conditions. Removal of this sentence is therefore required in order to reflect the varying environmental conditions under which the khapra beetle can survive and not limit its presence to just hot and dry environments.	Canada

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			cable trays and conduits, switch boxes etc. Larval exuviae become airborne very easily, and therefore it is always important to check window sills, grilles of venting holes and spider webs for their presence. Rodent traps containing baits should be always inspected.		
[109]	30	Technical	Inspections ~~Searches~~ for this pest are particularly difficult in cases of low-level infestations. In contrast to most other stored products pests, the Khapra beetle prefers hot and dry areas. The larvae of *Trogoderma* species are very crepuscular, and populations can persist in small quantities of residues that may occur within a structure or mode of transport. Larvae in diapause can survive long periods without food. For diapausing larvae it is important to examine ~~search~~ under piles of dirt, flaking paint and rust, in empty packaging materials such as hessian bags, tarpaulins, and corrugated cardboard. Larvae are often hiding behind wall panelling, under internal lining, between floorboards, under insulation, on dry ledges, electrical cable trays and conduits, switch boxes etc. Larval exuviae become airborne very easily, and therefore it is always important to check window sills, grilles of venting holes and spider webs for their presence. Rodent traps containing baits should be always inspected.	1) To be consistent with ISPM 5 (inspection is a visual examination) 2) Better wording.	El Salvador
[110]	30	Technical	~~Searches~~ Inspections for this pest are particularly difficult in cases of low-level infestations. In contrast to most other stored products pests, the Khapra beetle prefers hot and dry areas. The larvae of *Trogoderma* species are very crepuscular, and populations can persist in small quantities of residues that may occur within a structure or mode of transport. Larvae in diapause can survive long periods without food. For diapausing larvae it is important to examine ~~search~~ under piles of dirt, flaking paint and rust, in empty packaging materials such as hessian bags, tarpaulins, and corrugated cardboard. Larvae are often hiding behind wall panelling, under internal lining, between floorboards, under insulation, on dry ledges, electrical cable trays and conduits, switch boxes etc. Larval exuviae become airborne very easily, and therefore it is always important to check window sills, grilles of venting holes and spider webs for their presence. Rodent traps containing baits should be always inspected.	1) To be consistent with ISPM 5 (inspection is a visual examination) 2) Better wording.	OIRSA
[111]	31	Editorial	Khapra beetle infestations are usually recognized by (1) the presence of the pest (especially feeding larvae and exuviae) and (2) symptoms of infestation. The short-lived adults are sometimes not seen. Damage to the commodities can be a warning sign, but often it is a result of the feeding of other common stored product~~s~~ pests. Larvae usually feed first on the germ portion of cereal seeds and then on the endosperm. The seed coat is eaten in an irregular manner. In bulk commodities infestations usually concentrate in the surface layers, where numerous larval exuviae, broken setae and frass (excrement~~s~~) are present. However, larvae can occasionally be found as deep as 3–6 m in bulk grain.	'stored products pests': becomes 'stored product pests' throughout the document for sake of consistency (global change). 'Excrements' should be singular.	United States of America

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
[112]	31	Technical	Khapra beetle infestations are usually recognized by (1) the presence of the pest (especially feeding larvae and exuviae) and (2) symptoms of infestation. The short-lived adults are sometimes not seen. Damage to the commodities can be a warning sign, but often it is a result of the feeding of other common stored products pests. Larvae usually feed first on the germ portion of cereal seeds and then on the endosperm. The seed coat is eaten in an irregular manner. In bulk commodities infestations usually concentrate in the surface layers, where numerous larval exuviae, broken setae and frass (excrements) are present. However, larvae can occasionally be found as deep as 3–6 m in bulk grain.	It will be worth showing some photographs of feeding damage	New Zealand
[113]	31	Technical	Khapra beetle infestations are usually recognized by (1) the presence of the pest (especially feeding larvae and exuviae) and (2) symptoms of infestation. The short-lived adults are sometimes not seen. Damage to the commodities can be a warning sign, but often it is a result of the feeding of other common stored products pests. Larvae usually feed first on the germ portion of cereal seeds and then on the endosperm. The seed coat is eaten in an irregular manner. In bulk commodities infestations usually concentrate in the surface layers, where numerous larval exuviae, broken setae and frass (excrements) are present. However, larvae can occasionally be found as deep as 3–6 m in bulk grain. It is therefore important to consider biaised sampling when inspecting for these types of pests.	Add a new sentence at the end of para. 31 to provide additional guidance as to the type of sampling that should be considered when inspecting for khapra beetle	Canada
[114]	32	Editorial	Samples of suspect products have to be visually inspected in a well-lit area, using a 10× magnification hand lens. If no signs of *Trogoderma* infestation are found then larger samples of the product, whose size corresponds with the size of a given lot, should be passed over sieves with aperture sizes relevant to the particle size of the products. Usually sets of sieves of aperture sizes 1.0, 2.0 and 3.0 mm are used. The sifted material collected on particular sieves should be placed in Petri dishes and examined under at least 10× to 25× magnification through a stereoscopic microscope to detect the pest. This screening technique allows the detection of various developmental stages of the pest. However, some larvae feeding within grains may remain undetected. Therefore, it may become necessary to heat samples to 40 °C to drive pests out of the grains. Visual inspection is preferable because sieving can easily destroy or seriously damage dead adults and larval exuviae rendering the morphological identification very difficult or nearly impossible.		New Zealand
[115]	32	Editorial	Samples of suspect products have to be visually inspected in a well-lit area, using a 10× magnification hand lens. If no signs of *Trogoderma* infestation are found, then larger samples of the product, whose size corresponds with the size of a given lot, should be passed over sieves with aperture sizes relevant to the particle size of the products. Usually sets of	Sentence starting with 'if': Insert comma after clause. Decimal points and zeroes are not needed here because it is highly unlikely that there are field sieves with apertures between 1 and 2 mm, for instance. Whole numbers convey	United States of America

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			sieves of aperture sizes 1~~.0~~, 2~~.0~~ and 3~~.0~~ mm are used. The sifted material collected on particular sieves should be placed in Petri dishes and examined under at least 10× to 25× magnification through a stereoscopic microscope to detect the pest. This screening technique allows the detection of various developmental stages of the pest. However, some larvae feeding within grains may remain undetected. Therefore, it may become necessary to heat samples to 40 °C to drive pests out of the grains. When compared to sieving, ~~V~~visual inspection is preferable because sieving can easily destroy or seriously damage dead adults and larval exuviae rendering the identification very difficult or impossible.	the information adequately. 'When compared to sieving': added for sake of flow of thoughts and, thus, comprehension. We tried to place this sentence near the beginning of the paragraph, but it did not work satisfactorily.	
[116]	32	Substantive	Samples of suspect products have to be visually inspected in a well-lit area, using a 10× magnification hand lens. If no signs of *Trogoderma* infestation are found then larger samples of the product, whose size corresponds with the size of a given lot, should be passed over sieves with aperture sizes relevant to the particle size of the products. Usually sets of sieves of aperture sizes 1.0, 2.0 and 3.0 mm are used. The sifted material collected on particular sieves should be placed in Petri dishes and examined under at least 10× to 25× magnification through a stereoscopic microscope to detect the pest. This screening technique allows the detection of various developmental stages of the pest. However, some larvae feeding within grains may remain undetected. Therefore, it may become necessary to heat samples to 40 °C to drive pests out of the grains. Visual inspection is preferable because sieving can easily destroy or seriously damage dead adults and larval exuviae rendering the identification very difficult or impossible. Searches for this pest are particularly difficult in cases of low-level infestations. In contrast to most other stored products pests, T. granarium prefers hot and dry areas. The larvae of Trogoderma species are very crepuscular, and populations can persist in small quantities of residues that may occur within a structure or mode of transport. Larvae in diapause can survive long periods without food. For diapausing larvae it is important to search under piles of dirt, flaking paint and rust, in empty packaging materials such as hessian bags, tarpaulins, and corrugated cardboard. Larvae are often hiding behind wall panelling, under internal lining, between floorboards, under insulation, on dry ledges, electrical cable trays and conduits, switch boxes etc. Larval exuviae become airborne very easily, and therefore it is always important to check window sills, grilles of venting holes and spider webs for their presence. Rodent traps containing baits should be always inspected.	New paragraph after para 32 - moved from paragraph 30. More logical order	EPPO,Russian Federation ,Ukraine ,Morocco ,Uzbekistan
[117]	32	Substantive	Samples of suspect products have to be visually inspected in a well-lit area, using a 10× magnification hand lens. If no signs of *Trogoderma* infestation are found then larger samples of the product, whose size	New paragraph [32bis] after para [32] - moved from paragraph [30]. More logical order	European Union

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			corresponds with the size of a given lot, should be passed over sieves with aperture sizes relevant to the particle size of the products. Usually sets of sieves of aperture sizes 1.0, 2.0 and 3.0 mm are used. The sifted material collected on particular sieves should be placed in Petri dishes and examined under at least 10× to 25× magnification through a stereoscopic microscope to detect the pest. This screening technique allows the detection of various developmental stages of the pest. However, some larvae feeding within grains may remain undetected. Therefore, it may become necessary to heat samples to 40 °C to drive pests out of the grains. Visual inspection is preferable because sieving can easily destroy or seriously damage dead adults and larval exuviae rendering the identification very difficult or impossible. [32bis] Searches for this pest are particularly difficult in cases of low-level infestations. In contrast to most other stored products pests, T. granarium prefers hot and dry areas. The larvae of Trogoderma species are very crepuscular, and populations can persist in small quantities of residues that may occur within a structure or mode of transport. Larvae in diapause can survive long periods without food. For diapausing larvae it is important to search under piles of dirt, flaking paint and rust, in empty packaging materials such as hessian bags, tarpaulins, and corrugated cardboard. Larvae are often hiding behind wall panelling, under internal lining, between floorboards, under insulation, on dry ledges, electrical cable trays and conduits, switch boxes etc. Larval exuviae become airborne very easily, and therefore it is always important to check window sills, grilles of venting holes and spider webs for their presence. Rodent traps containing baits should be always inspected.		
[118]	32	Technical	Samples of suspect products have to be visually inspected in a well-lit area, using a 10× magnification hand lens. If appropriate, ~~no signs of *Trogoderma* infestation are found then larger~~ samples ~~of the product, whose size corresponds with the size of a given lot,~~ should be passed over sieves with aperture sizes relevant to the particle size of the products. Usually sets of sieves of aperture sizes 1.0, 2.0 and 3.0 mm are used. The sifted material collected on particular sieves should be placed in Petri dishes and examined under at least 10× to 25× magnification through a stereoscopic microscope to detect the pest. This screening technique allows the detection of various developmental stages of the pest. However, some larvae feeding within grains may remain undetected. Therefore, it may become necessary to heat samples to 40 °C to drive pests out of the grains. Visual inspection is preferable because sieving can easily destroy or seriously damage dead adults and larval exuviae rendering the identification very difficult or impossible.	Delete references to taking initial samples and then taking larger samples. This could conflict with the ISPM on sampling and create confusion.	EPPO,Russian Federation ,Ukraine ,Morocco ,Uzbekistan
[119]	32	Technical	Samples of suspect products have to be ~~visually~~ inspected in a well-lit	The sample should be representative of the lot.	Costa Rica

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			area, using a 10× magnification hand lens. If no signs of *Trogoderma* infestation are found then ~~larger~~ samples of the product, ~~whose size corresponds with the size of a given lot,~~ should be passed over sieves with aperture sizes relevant to the particle size of the products. Usually sets of sieves of aperture sizes 1.0, 2.0 and 3.0 mm are used. The sifted material collected on particular sieves should be placed in Petri dishes and examined under at least 10× to 25× magnification through a stereoscopic microscope to detect the pest. This screening technique allows the detection of various developmental stages of the pest. However, some larvae feeding within grains may remain undetected. Therefore, it may become necessary to heat samples to 40 °C to drive pests out of the grains. ~~Visual i~~Inspection is preferable because sieving can easily destroy or seriously damage dead adults and larval exuviae rendering the identification very difficult or impossible.	So it is not clear what larger samples of the products refers to. To be consistent with the Glossary term of inspection.	,Mexico ,Nicaragua ,El Salvador ,Brazil
[120]	32	Technical	Samples of suspect products have to be ~~visually~~ inspected in a well-lit area, using a 10× magnification hand lens. If no signs of *Trogoderma* infestation are found then ~~larger~~ samples of the product, ~~whose size corresponds with the size of a given lot~~, should be passed over sieves with aperture sizes relevant to the particle size of the products. Usually sets of sieves of aperture sizes 1.0, 2.0 and 3.0 mm are used. The sifted material collected on particular sieves should be placed in Petri dishes and examined under at least 10× to 25× magnification through a stereoscopic microscope to detect the pest. This screening technique allows the detection of various developmental stages of the pest. However, some larvae feeding within grains may remain undetected. Therefore, it may become necessary to heat samples to 40 °C to drive pests out of the grains. ~~Visual i~~Inspection is preferable because sieving can easily destroy or seriously damage dead adults and larval exuviae rendering the identification very difficult or impossible.	1) To be consistent with glossary term "inspection" 2) The sample should be representative of the lot, so it is not clear what larger samples of the product refer to. 3) Idem comment 1), to be consistent with glossary term "inspection"	Uruguay
[121]	32	Technical	Samples of suspect products have to be ~~visually~~ inspected in a well-lit area, using a 10× magnification hand lens. If no signs of *Trogoderma* infestation are found then ~~larger~~ samples of the product, ~~whose size corresponds with the size of a given lot,~~ should be passed over sieves with aperture sizes relevant to the particle size of the products. Usually sets of sieves of aperture sizes 1.0, 2.0 and 3.0 mm are used. The sifted material collected on particular sieves should be placed in Petri dishes and examined under at least 10× to 25× magnification through a stereoscopic microscope to detect the pest. This screening technique allows the detection of various developmental stages of the pest. However, some larvae feeding within grains may remain undetected. Therefore, it may become necessary to heat samples to 40 °C to drive pests out of the grains. ~~Visual i~~Inspection is preferable because sieving can easily destroy or seriously damage dead adults and larval exuviae rendering the	The sample should be representative of the lot so it is not clear what larger samples of the products refer to. Text "Visual" deleted to be consistent with glossary term inspection	COSAVE,Paraguay ,Chile

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			identification very difficult or impossible.		
[122]	32	Technical	Samples of suspect products have to be ~~visually~~ inspected in a well-lit area, using a 10× magnification hand lens. If no signs of *Trogoderma* infestation are found then ~~larger~~ samples of the product, ~~whose size corresponds with the size of a given lot~~, should be passed over sieves with aperture sizes relevant to the particle size of the products. Usually sets of sieves of aperture sizes 1.0, 2.0 and 3.0 mm are used. The sifted material collected on particular sieves should be placed in Petri dishes and examined under at least 10× to 25× magnification through a stereoscopic microscope to detect the pest. This screening technique allows the detection of various developmental stages of the pest. However, some larvae feeding within grains may remain undetected. Therefore, it may become necessary to heat samples to 40 °C to drive pests out of the grains. ~~Visual~~ inspection is preferable because sieving can easily destroy or seriously damage dead adults and larval exuviae rendering the identification very difficult or impossible.	The sample should be representative of the lot so it is not clear what larger samples of the products refer to. To be consistent with glossary term inspection	Argentina
[123]	32	Technical	Samples of suspect products have to be visually inspected in a well-lit area, using a 10× magnification hand lens. If appropriate, ~~no signs of *Trogoderma* infestation are found then larger~~ samples ~~of the product, whose size corresponds with the size of a given lot,~~ should be passed over sieves with aperture sizes relevant to the particle size of the products. Usually sets of sieves of aperture sizes 1.0, 2.0 and 3.0 mm are used. The sifted material collected on particular sieves should be placed in Petri dishes and examined under at least 10× to 25× magnification through a stereoscopic microscope to detect the pest. This screening technique allows the detection of various developmental stages of the pest. However, some larvae feeding within grains may remain undetected. Therefore, it may become necessary to heat samples to 40 °C to drive pests out of the grains. Visual inspection is preferable because sieving can easily destroy or seriously damage dead adults and larval exuviae rendering the identification very difficult or impossible.	Delete references to taking initial samples and then taking larger samples. This could conflict with the ISPM on sampling and create confusion.	European Union
[124]	32	Technical	Samples of suspect products have to be visually inspected in a well-lit area, using a 10× magnification hand lens. If no signs of *Trogoderma* infestation are found then larger samples of the product, whose size corresponds with the size of a given lot, should be passed over sieves with aperture sizes relevant to the particle size of the products. Usually sets of sieves of aperture sizes 1.0, 2.0 and 3.0 mm are used. The sifted material collected on particular sieves should be placed in Petri dishes and examined under at least 10× to 25× magnification through a stereoscopic microscope to detect the pest. This screening technique allows the detection of various developmental stages of the pest. However, some larvae feeding within grains may remain undetected. Therefore, it may	The suggested text in in the 4th sentence is in line with general comment formulated before. The diagnostioc protocol should refer to the use of Berlese funnels to extract adults and larvae of Trogoderma granarium.	Canada

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			become necessary to heat samples to 40 °C to drive pests out of the grains with an extraction tool such as a Berlese funnel. Visual inspection is preferable because sieving can easily destroy or seriously damage dead adults and larval exuviae rendering the identification very difficult or impossible.		
[125]	32	Technical	Samples of suspect products have to be ~~visually~~ inspected in a well-lit area, using a 10× magnification hand lens. If no signs of *Trogoderma* infestation are found then ~~larger~~ samples of the product, ~~whose size corresponds with the size of a given lot,~~ should be passed over sieves with aperture sizes relevant to the particle size of the products. Usually sets of sieves of aperture sizes 1.0, 2.0 and 3.0 mm are used. The sifted material collected on particular sieves should be placed in Petri dishes and examined under at least 10× to 25× magnification through a stereoscopic microscope to detect the pest. This screening technique allows the detection of various developmental stages of the pest. However, some larvae feeding within grains may remain undetected. Therefore, it may become necessary to heat samples to 40 °C to drive pests out of the grains. ~~Visual i~~Inspection is preferable because sieving can easily destroy or seriously damage dead adults and larval exuviae rendering the identification very difficult or impossible.	1) To be consistent with glossary term inspection 2) The sample should be representative of the lot so it is not clear what larger samples of the products refer to. 3) To be consistent with glossary term inspection.	OIRSA
[126]	33	Substantive	~~Insects found should be picked up carefully with small forceps or collected using an aspirator. It is important to collect multiple specimens of the pest. Identification of larvae is difficult, and if the dissection of a single specimen is not successful and serious damage occurs to the mouthparts, then exact identification is impossible. Specimens should be placed in 70% ethyl alcohol.~~	Move to after paragraph 34	EPPO,Russian Federation ,European Union ,Ukraine ,Morocco ,Uzbekistan
[127]	33	Substantive	Insects found should be picked up carefully with small forceps or collected using an aspirator. It is important to collect multiple specimens of the pest. Identification of larvae is difficult, and if the dissection of a single specimen is not successful and serious damage occurs to the mouthparts, then exact identification is impossible. Specimens should be placed in 70% ethyl alcohol for preservation if the identification is not immediately done.	The insertion brings out the function of the alcohol which is to preserve the specimen in case identification will be done at a later stage.	South Africa
[128]	34	Editorial	Additionally, it is possible to monitor the presence of *T. granarium* using various traps. Food-baited traps (containing oil seeds, peanuts, wheat germ etc.) or attractant traps (containing wheat germ oil) can be used to attract larvae. Simple ~~T~~traps ~~can be as simple as~~ offering hiding places for the larvae, such as pieces of corrugated cardboard or hessian bag can be placed on the floor. After finishing the monitoring, all the traps should be collected and destroyed. Adults may be detected with the use of pheromone traps where the pheromone capsule is combined with a non-drying sticky trap. However, the *Trogoderma* pheromone traps are not	Simpler language.	EPPO,Russian Federation ,Ukraine ,Morocco ,Uzbekistan

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			species-specific and attract many species of dermestid beetles (Saplina, 1984; Barak, 1989; Barak *et al.*, 1990; Mordkovich and Sokolov, 2000). Traps baited with pheromone and food bait are commercially available for these species.		
[129]	34	Editorial	Additionall~~y~~to initial inspections, it is possible to monitor the presence of *T. granarium* using various traps. Food-baited traps (containing oil seeds, peanuts, wheat germ etc.) or attractant traps (containing wheat germ oil) can be used to attract larvae. Traps can be as simple as offering hiding places for the larvae, such as pieces of corrugated cardboard or hessian bag placed on the floor. After finishing the monitoring, all the traps should be collected and destroyed. Adults may be detected with the use of pheromone traps where the pheromone capsule is combined with a non-drying sticky trap. However, the *Trogoderma* pheromone traps are not species-specific and attract many species of dermestid beetles (Saplina, 1984; Barak, 1989; Barak *et al.*, 1990; Mordkovich and Sokolov, 2000). Traps baited with both pheromone and food ~~bait~~ are commercially available for these species.	To improve the flow of ideas from paragraph 33 to paragraph 34. 'Traps baited with .....bait' is redundant. Addition of 'both' sets this type of trap apart from the traps in the previous sentence that rely solely on pheromones.	United States of America
[130]	34	Editorial	Additionally, it is possible to monitor the presence of *T. granarium* using various traps. Food-baited traps (containing oil seeds, peanuts, wheat germ etc.) or attractant traps (containing wheat germ oil) can be used to attract larvae. Simple ~~T~~traps ~~can be as simple as~~ offering hiding places for the larvae, such as pieces of corrugated cardboard or hessian bag can be placed on the floor. After finishing the monitoring, all the traps should be collected and destroyed. Adults may be detected with the use of pheromone traps where the pheromone capsule is combined with a non-drying sticky trap. However, the *Trogoderma* pheromone traps are not species-specific and attract many species of dermestid beetles (Saplina, 1984; Barak, 1989; Barak *et al.*, 1990; Mordkovich and Sokolov, 2000). Traps baited with pheromone and food bait are commercially available for these species.	Simpler language.	European Union
[131]	34	Substantive	Additionally, it is possible to monitor the presence of *T. granarium* using various traps. Food-baited traps (containing oil seeds, peanuts, wheat germ etc.) or attractant traps (containing wheat germ oil) can be used to attract larvae. Traps can be as simple as offering hiding places for the larvae, such as pieces of corrugated cardboard or hessian bag placed on the floor. After finishing the monitoring, all the traps should be collected and destroyed. Adults may be detected with the use of pheromone traps where the pheromone capsule is combined with a non-drying sticky trap. However, the *Trogoderma* pheromone traps are not species-specific and attract many species of dermestid beetles (Saplina, 1984; Barak, 1989; Barak *et al.*, 1990; Mordkovich and Sokolov, 2000). Traps baited with pheromone and food bait are commercially available for these species.	New paragraph after 34 - moved from 33. More logical order.	EPPO,Russian Federation ,Ukraine ,Morocco ,Uzbekistan

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			Insects found should be picked up carefully with small forceps or collected using an aspirator. It is important to collect multiple specimens of the pest. Identification of larvae is difficult, and if the dissection of a single specimen is not successful and serious damage occurs to the mouthparts, then exact identification is impossible. Specimens should be placed in 70% ethyl alcohol.		
[132]	34	Substantive	Additionally, it is possible to monitor the presence of *T. granarium* using various traps. Food-baited traps (containing oil seeds, peanuts, wheat germ etc.) or attractant traps (containing wheat germ oil) can be used to attract larvae. Traps can be as simple as offering hiding places for the larvae, such as pieces of corrugated cardboard or hessian bag placed on the floor. After finishing the monitoring, all the traps should be collected and destroyed. Adults may be detected with the use of pheromone traps where the pheromone capsule is combined with a non-drying sticky trap. However, the *Trogoderma* pheromone traps are not species-specific and attract many species of dermestid beetles (Saplina, 1984; Barak, 1989; Barak *et al.*, 1990; Mordkovich and Sokolov, 2000). Traps baited with pheromone and food bait are commercially available for these species. [34bis] Insects found should be picked up carefully with small forceps or collected using an aspirator. It is important to collect multiple specimens of the pest. Identification of larvae is difficult, and if the dissection of a single specimen is not successful and serious damage occurs to the mouthparts, then exact identification is impossible. Specimens should be placed in 70% ethyl alcohol.	New paragraph [34bis] after [34] - moved from [33]. More logical order.	European Union
[133]	36	Editorial	The genus *Trogoderma* in recent years has been reported to includ~~es~~e 117 species (Mroczkowski, 1968),~~according to Mroczkowski (1968) 117 species;~~ 115 species (Beal, 1982),~~according to Beal (1982) 115 species;~~ and 130 species (Hava, 2003) ~~according to Háva (2003) 130 species~~. There are many other species of *Trogoderma* yet to be described. Great caution needs to be exercised with the synonymies established because few of them are based on detailed comparison of the type specimens.	Rearranged to improve comprehension.	United States of America
[134]	36	Substantive	~~T~~According to Mroczowski (1968), the genus *Trogoderma* includes ~~according to Mroczkowski (1968)~~ 117 species; according to Beal (1982) 115 species; and according to Háva (2003) 130 species. There are many other species of *Trogoderma* yet to be described. Great caution needs to be exercised with the synonymies established because few of them are based on detailed comparison of the type specimens.	The reconfiguration of the words emphasizes the difference in authors' opinions.	South Africa
[135]	36	Technical	The genus *Trogoderma* includes according to Mroczkowski (1968) 117 species; according to Beal (1982) 115 species; and according to Háva (2003) 130 species and Háva (2011) - 134 species. There are many other species of *Trogoderma* yet to be described. Great caution needs to be	A new paper with updated numbers.	EPPO,Russian Federation ,Ukraine ,Morocco

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			exercised with the synonymies established because few of them are based on detailed comparison of the type specimens.		,Uzbekistan
[136]	36	Technical	The genus *Trogoderma* includes according to Mroczkowski (1968) 117 species; according to Beal (1982) 115 species; and according to Háva (2003) 130 species and Háva (2011) - 134 species. There are many other species of *Trogoderma* yet to be described. Great caution needs to be exercised with the synonymies established because few of them are based on detailed comparison of the type specimens.	A new paper with updated numbers.	European Union
[137]	37	Substantive	Identification of *Trogoderma* eggs and pupae based on external features is currently not possible. Insect eggs and pupae possess very few external features and therefore are poorly studied. Larval identification is difficult. It requires experience in identification and also good skills in dissection of small insects. Pupation takes place in the last larval cast. The larval exuviae can be used for identification, but one needs to be more cautious because the material is brittle. Adults are the easiest to identify, though misidentification ~~by less experienced entomologists~~ is still common.	The words “by less experienced entomologists” should be deleted because anyone can misidentify even experienced entomologists.	South Africa
[138]	37	Technical	Identification of *Trogoderma* eggs and pupae based on external features is currently not possible. Insect eggs and pupae possess very few external features and therefore are poorly studied. Larval identification is difficult. It requires experience in identification and also good skills in dissection of small insects. Pupation takes place in the last larval cast. The larval exuviae can be used for identification, but one needs to be more cautious because the material is brittle. Adults are the easiest to identify, though misidentification by less experienced entomologists is still common so, training in insect preparation and mounting procedures is required to apply this protocol.	Training is necessary to apply this protocol.	Costa Rica ,Nicaragua ,El Salvador
[139]	37	Technical	Identification of *Trogoderma* eggs and pupae based on external features is currently not possible. Insect eggs and pupae possess very few external features and therefore are poorly studied. Larval identification is difficult. It requires experience in identification and also good skills in dissection of small insects. Pupation takes place in the last larval cast. The larval exuviae can be used for identification, but one needs to be more cautious because the material is brittle. Adults are the easiest to identify, though misidentification by less experienced entomologists is still common, so training in insect preparation and mounting procedures is required to apply this protocol.	Training is necessary to apply this protocol.	Uruguay
[140]	37	Technical	Identification of *Trogoderma* eggs and pupae based on external features is currently not possible. Insect eggs and pupae possess very few external features and therefore are poorly studied. Larval identification is difficult. It	Training is necessary to apply this protocol.	COSAVE,Paraguay ,Chile,Brazil

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			requires experience in identification and also good skills in dissection of small insects. Pupation takes place in the last larval cast. The larval exuviae can be used for identification, but one needs to be more cautious because the material is brittle. Adults are the easiest to identify, though misidentification by less experienced entomologists is still common so training in insect preparation and montage procedures is required to apply this protocol..		
[141]	37	Technical	Identification of *Trogoderma* eggs and pupae based on external features is currently not possible. Insect eggs and pupae possess very few external features and therefore are poorly studied. Larval identification is difficult. It requires experience in identification and also good skills in dissection of small insects. Pupation takes place in the last larval cast. The larval exuviae can be used for identification, but one needs to be more cautious because the material is brittle. Adults are the easiest to identify, though misidentification by less experienced entomologists is still common.so training in insect preparation and montage procedures is required to apply this protocol.	Training is necessary to apply this protocol.	Argentina
[142]	37	Technical	Identification of *Trogoderma* eggs and pupae based on external features is currently not possible. Insect eggs and pupae possess very few external features and therefore are poorly studied. Larval identification is difficult. It requires experience in identification and also good skills in dissection of small insects. Pupation takes place in the last larval cast. The larval exuviae can be used for identification, but one needs to be more cautious because the material is brittle. Adults are the easiest to identify, though misidentification by less experienced entomologists is still common so, training in insect preparation and mounting procedures is required to apply this protocol.	Training is necessary to apply this protocol.	OIRSA
[143]	38	Editorial	Adults in good condition can be identified under a stereomicroscope using 10× to 100× magnification. However, movement of the stored product, particularly cereals, will damage the dead adults. In most cases the legs and antennae will break off and also the setae on elytra and pronotum will be rubbed off. In the case of a damaged specimen with missing body parts or morphological features not visible, it is necessary to examine the genitalia. For reliable identification the genitalia should ~~be~~ always be examined. Genitalia should be removed (section 4.2) and mounted on a cavity microscope slide temporarily using glycerol or (for a longer time) using Hoyer's mounting medium (50 ml water, 30 g gum arabic, 200 g chloral hydrate, 20 ml glycerine).	Grammar.	United States of America
[144]	38	Substantive	Adults in good condition can be identified under a stereomicroscope using 10× to 100× magnification. However, movement of the stored product, particularly cereals, will damage the dead adults. In most cases the legs	The standard recipe should be 16 ml glycerine. The mounting medium is too thin if 20 ml glycerine added to make the medium.	Singapore

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			and antennae will break off and also the setae on elytra and pronotum will be rubbed off. In the case of a damaged specimen with missing body parts or morphological features not visible, it is necessary to examine the genitalia. For reliable identification the genitalia should be always examined. Genitalia should be removed (section 4.2) and mounted on a cavity microscope slide temporarily using glycerol or (for a longer time) using Hoyer's mounting medium (50 ml water, 30 g gum arabic, 200 g chloral hydrate, ~~20~~ 16 ml glycerine).		
[145]	38	Technical	Adults in good condition can be identified under a stereomicroscope using 10× to 100× magnification. However, for reliable identification it is recommended that the genitalia are always examined. ~~m~~Movement of the stored product, particularly cereals, will damage the dead adults. In most cases the legs and antennae will break off and also the setae on elytra and pronotum will be rubbed off. In the case of a damaged specimen with missing body parts or morphological features not visible, identification should always be based on ~~it is necessary to~~ examination of~~e~~ the genitalia. ~~For reliable identification the genitalia should be always examined.~~ Genitalia should be removed (section 4.2) and mounted on a cavity microscope slide temporarily using glycerol or (for a longer time) using Hoyer's mounting medium (50 ml water, 30 g gum arabic, 200 g chloral hydrate, 20 ml glycerine).	As currently drafted, there is a conflict between 1st and 5th sentences. The first sentence indicates that adults in good condition can be identified (presumably reliably), but the 5th sentence states that for reliable identification the genitalia should always be examined. An amendment of the text is proposed in order to make it clearer.	EPPO,Russian Federation ,Ukraine ,Morocco ,Uzbekistan
[146]	38	Technical	Adults in good condition can be identified under a stereomicroscope using 10× to 100× magnification. However, movement of the stored product, particularly cereals, will damage the dead adults. In most cases the legs and antennae will break off and also the setae on elytra and pronotum will be rubbed off. In the case of a damaged specimen with missing body parts or morphological features not visible, it is necessary to examine the genitalia. However, ~~F~~for reliable identification, the genitalia should be always examined. Genitalia should be removed (section 4.2) and mounted on a cavity microscope slide temporarily using glycerol or (for a longer time) using Hoyer's mounting medium (50 ml water, 30 g gum arabic, 200 g chloral hydrate, 20 ml glycerine).	Better wording to highlight that for reliable identification, genitalia should always be examined.	Uruguay
[147]	38	Technical	Adults in good condition can be identified under a stereomicroscope using 10× to 100× magnification. However, movement of the stored product, particularly cereals, will damage the dead adults. In most cases the legs and antennae will break off and also the setae on elytra and pronotum will be rubbed off. In the case of a damaged specimen with missing body parts or morphological features not visible, it is necessary to examine the genitalia. However, ~~F~~for the reliable identification the genitalia should be always examined. Genitalia should be removed (section 4.2) and mounted on a cavity microscope slide temporarily using glycerol or (for a longer time) using Hoyer's mounting medium (50 ml water, 30 g gum arabic,	Better wording to highlight that for identification, genitalia should always be examined.	COSAVE,Paraguay ,Chile,Brazil

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			200 g chloral hydrate, 20 ml glycerine).		
[148]	38	Technical	Adults in good condition can be identified under a stereomicroscope using 10× to 100× magnification. However, for reliable identification it is recommended that the genitalia are always examined. ~~m~~Movement of the stored product, particularly cereals, will damage the dead adults. In most cases the legs and antennae will break off and also the setae on elytra and pronotum will be rubbed off. In the case of a damaged specimen with missing body parts or morphological features not visible, identification should always be based on ~~it is necessary to~~ examination of~~e~~ the genitalia. ~~For reliable identification the genitalia should be always examined.~~ Genitalia should be removed (section 4.2) and mounted on a cavity microscope slide temporarily using glycerol or (for a longer time) using Hoyer's mounting medium (50 ml water, 30 g gum arabic, 200 g chloral hydrate, 20 ml glycerine).	As originally drafted, there is a conflict between 1st and 5th sentences. The first sentence indicates that adults in good condition can be identified (presumably reliably), but the 5th sentence states that for reliable identification the genitalia should always be examined. An amendment of the text is proposed in order to make it clearer.	European Union
[149]	39	Substantive	For larval identifications the mouthparts should be dissected out (section 4.1). The larval exuviae and dissected mouthparts (catskin)should be mounted on a cavity microscope slide using Hoyer's medium (Beal, 1960) or PVA (Polyvinyl alcohol). Details of mounting procedures are included in section 4.1.		Indonesia
[150]	41	Substantive	Methods have been developed for the identification of a limited number of pest *Trogoderma* species using both immunological (ELISA test) and molecular techniques. These cannot be used yet as quarantine diagnostic techniques for the determination of species within the *Trogoderma* genus.	SA would like to propose that a clear statement be given on why ELISA and molecular techniques cannot be used as quarantine diagnostic techniques for Trogoderma genus.	South Africa
[151]	43	Editorial	Before dissection the larva should be examined under a stereomicroscope. Size, body colour, arrangement and colour of setae should be recorded. Photomicrographs area a useful method of recording features.	Photography as a method to record patterning on the elytra etc… as the use of microscope photography is widespread and frequently available now to provide a record of material prior to disturbance via manipulation and handling- provides an independent interpretation of the material	Australia
[152]	44	Editorial	For identification larvae should be mounted on Hoyer's medium on a microscope slide using the following method: ~~:~~	The colon was added to show the contuinity with the changes made in paragraph 45.	Costa Rica ,Uruguay ,Nicaragua ,El Salvador
[153]	44	Editorial	For identification the larva~~e~~ should be mounted ~~on~~in Hoyer's medium on a microscope slide using the following method.	Paragraphs 43 and 45 use the singular 'larva', so the singular should be used in paragraph 44 as well.	United States of America
[154]	44	Editorial	For identification larvae should be mounted on Hoyer's medium on a microscope slide using the following method: ~~:~~	Add the colon at the end of the paragraph 44 to give the appropiate sequence to the next	Mexico

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
				paragraph.	
[155]	44	Editorial	For identification larvae should be mounted ~~on~~ in Hoyer's medium on a microscope slide using the following method.	Correct preposition.	European Union
[156]	44	Editorial	For identification larvae should be mounted on Hoyer's medium on a microscope slide using the following method:~~-~~	The colon was added to show continuity with the changes made in paragraph 45	OIRSA
[157]	44	Substantive	For identification larvae should be mounted on Hoyer's medium or PVA (Polyvinyl alcohol) on a microscope slide using the following method.		Indonesia
[158]	45	Editorial	At first, place the specimen ventral side up on a microscope slide. Cut open the whole body along the mid-line from under the head capsule to the last abdominal segment using eye surgery scissors. Next put the larva into a test-tube containing 10% potassium hydroxide (KOH) solution and heat in a boiling water bath until it has become clear. Rinse thoroughly in warm distilled water. Remove all internal tissues using a very fine, short hair brush or the convex surface of a hooked tip of a no. 1 insect pin, or a loop formed from a micropin. All setae should be removed from one side of the 7th and 8th abdominal segment. Remove the head capsule and put it back in the hot KOH solution for 5 minutes. Rinse the head capsule in warm distilled water. Dissection of the head can be performed in a few drops of Hoyer's or glycerol on a microscope slide or in water in an excavated glass block. Turn the head ventral side up and hold it to the glass with a blunt no. 1 insect pin. Remove the mandibles, maxillae and labial palpi using jeweller's forceps and micropins. Remove the epipharynx and antennae. Mount the head capsule and the mandibles in the cavity of the slide using Hoyer's. Mount the cleared skin, fully opened on the flat part of the microscope slide, next to the cavity. It is usually best done ventral side up. Epipharynx, antennae, maxillae and labial palpi should be mounted with the skin under the same cover slip. Mount all body parts on the same microscope slide. -In the case of larval exuviae, before proceeding with the dissection soak the specimen in a 5% solution of any laboratory detergent (e.g. Decon 90) for about two hours and thoroughly rinse in distilled water. Cut the specimen open anteriorly and dissect out the mouthparts. They can be mounted directly in Hoyer's without clearing. Label slides immediately after mounting specimens and place them in an oven for a few days at 40 °C. After drying, ring the slides using any lacquer recommended for sealing of microscopic slides (e.g. Glyptol, Brunseal), or at least two layers of nail polish in order to prevent the Hoyer's from drying and possibly damaging the specimen.	First sentence: Without adding a comma, non native speakers can read "at first place" as "at first location". 17th sentence: It would be clearer to begin with a new paragraph (i.e. one paragraph for larvae, and one paragraph for larval exuviae).	EPPO,Russian Federation ,Ukraine ,Morocco ,Uzbekistan
[159]	45	Editorial	a) At first place the specimen ventral side up on a microscope slide.	To clarify the procedure for preparation of the	Costa Rica

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			b) Cut open the whole body along the mid-line from under the head capsule to the last abdominal segment using eye surgery scissors. c) Next put the larva into a test-tube containing 10% potassium hydroxide (KOH) solution and heat in a boiling water bath until it has become clear. d) Rinse thoroughly in warm distilled water. e) Remove all internal tissues using a very fine, short hair brush or the convex surface of a hooked tip of a no. 1 insect pin, or a loop formed from a micropin. All setae should be removed from one side of the 7th and 8th abdominal segment. f) Remove the head capsule and put it back in the hot KOH solution for 5 minutes. Rinse the head capsule in warm distilled water.–Dissection of the head can be performed in a few drops of Hoyer's or glycerol on a microscope slide or in water in an excavated glass block. Turn the head ventral side up and hold it to the glass with a blunt no. 1 insect pin. g) Remove the mandibles, maxillae and labial palpi using jeweller's forceps and micropins. Remove the epipharynx and antennae. Mount the head capsule and the mandibles in the cavity of the slide using Hoyer's. Mount the cleared skin, fully opened on the flat part of the microscope slide, next to the cavity. It is usually best done ventral side up. Epipharynx, antennae, maxillae and labial palpi should be mounted with the skin under the same cover slip. Mount all body parts on the same microscope slide. h) In the case of larval exuviae, before proceeding with the dissection soak the specimen in a 5% solution of any laboratory detergent (e.g. Decon 90) for about two hours and thoroughly rinse in distilled water. Cut the specimen open anteriorly and dissect out the mouthparts. They can be mounted directly in Hoyer's without clearing. Label slides immediately after mounting specimens and place them in an oven for a few days at 40 °C. After drying, ring the slides using any lacquer recommended for sealing of microscopic slides (e.g. Glyptol, Brunseal), or at least two layers of nail polish in order to prevent the Hoyer's from drying and possibly damaging the specimen.	larvae, the paragraph was separated using bullets.	,Uruguay ,Mexico ,Nicaragua ,El Salvador
[160]	45	Editorial	At first place the specimen ventral side up on a microscope slide. a) Cut open the whole body along the mid-line from under the head capsule to the last abdominal segment using eye surgery scissors. b) Next put the larva into a test-tube containing 10% potassium hydroxide (KOH) solution and heat in a boiling water bath until it has become clear. c) Rinse thoroughly in warm distilled water. d) Remove all internal tissues using a very fine, short hair brush or the convex surface of a hooked tip of a no. 1 insect pin, or a loop formed from a micropin. All setae should be removed from one side of the 7th and 8th	To clarify bullets were added	COSAVE,Paraguay ,Chile,Brazil

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			abdominal segment. e) Remove the head capsule and put it back in the hot KOH solution for 5 minutes. Rinse the head capsule in warm distilled water. Dissection of the head can be performed in a few drops of Hoyer's or glycerol on a microscope slide or in water in an excavated glass block. Turn the head ventral side up and hold it to the glass with a blunt no. 1 insect pin. f) Remove the mandibles, maxillae and labial palpi using jeweller's forceps and micropins. Remove the epipharynx and antennae. Mount the head capsule and the mandibles in the cavity of the slide using Hoyer's. Mount the cleared skin, fully opened on the flat part of the microscope slide, next to the cavity. It is usually best done ventral side up. Epipharynx, antennae, maxillae and labial palpi should be mounted with the skin under the same cover slip. Mount all body parts on the same microscope slide. g) In the case of larval exuviae, before proceeding with the dissection soak the specimen in a 5% solution of any laboratory detergent (e.g. Decon 90) for about two hours and thoroughly rinse in distilled water. Cut the specimen open anteriorly and dissect out the mouthparts. They can be mounted directly in Hoyer's without clearing. Label slides immediately after mounting specimens and place them in an oven for a few days at 40 °C. After drying, ring the slides using any lacquer recommended for sealing of microscopic slides (e.g. Glyptol, Brunseal), or at least two layers of nail polish in order to prevent the Hoyer's from drying and possibly damaging the specimen.		
[161]	45	Editorial	~~At first~~ First, place the specimen ventral side up on a microscope slide. Cut open the whole body along the mid-line from under the head capsule to the last abdominal segment using eye surgery scissors. Next put the larva into a tes~~t~~ tube containing 10% potassium hydroxide (KOH) solution and heat in a boiling water bath until ~~it has become clear~~larval tissues loosen and begin to separate from the cuticle. Rinse thoroughly in warm distilled water. Remove all internal tissues using a very fine, short- haired brush or the convex surface of a hooked tip of a no. 1 insect pin, or a loop formed from a micropin. All setae should be removed from one side of the 7th and 8th abdominal segment. Remove the head capsule and put it back in the hot KOH solution for 5 minutes. Rinse the head capsule in warm distilled water. Dissection of the head can be performed in a few drops of Hoyer's or glycerol on a microscope slide or in water in an excavated glass block. Turn the head ventral side up and hold it to the glass with a blunt no. 1 insect pin. Remove the mandibles, maxillae and labial palpi using jeweller's forceps and micropins. Remove the epipharynx and antennae. Mount the head capsule and the mandibles in the cavity of the slide using Hoyer's . Mount the cleared skin, fully opened on the flat part of the microscope slide, next to the cavity. It is usually best	'First': word usage. 'Test-tube': Test tube is a noun and test-tube is an adjective. Here it is used as a noun. 'It has become clear': What has become clear? The KOH solution? A 10% solution is already clear. The test tube itself is clear from the start. We think the author is referring to the larval itself. 'Short hair brush': Short-hair is an adjective, and short-haired is more grammatically correct. Commas usually follow 'for example', 'for instance', and other such prepositional phrases.	United States of America

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			done ventral side up. Epipharynx, antennae, maxillae and labial palpi should be mounted with the skin under the same cover slip. Mount all body parts on the same microscope slide. In the case of larval exuviae, before proceeding with the dissection, soak the specimen in a 5% solution of any laboratory detergent (e.g., Decon 90) for about two hours and thoroughly rinse in distilled water. Cut the specimen open anteriorly and dissect out the mouthparts. They can be mounted directly in Hoyer's without clearing. Label slides immediately after mounting specimens and place them in an oven for a few days at 40 °C. After drying, ring the slides using any lacquer recommended for sealing of microscopic slides (e.g., Glyptol, Brunseal), or at least two layers of nail polish in order to prevent the Hoyer's from drying and possibly damaging the specimen.		
[162]	45	Editorial	At first, place the specimen ventral side up on a microscope slide. Cut open the whole body along the mid-line from under the head capsule to the last abdominal segment using eye surgery scissors. Next put the larva into a test-tube containing 10% potassium hydroxide (KOH) solution and heat in a boiling water bath until it has become clear. Rinse thoroughly in warm distilled water. Remove all internal tissues using a very fine, short hair brush or the convex surface of a hooked tip of a no. 1 insect pin, or a loop formed from a micropin. All setae should be removed from one side of the 7th and 8th abdominal segment. Remove the head capsule and put it back in the hot KOH solution for 5 minutes. Rinse the head capsule in warm distilled water. Dissection of the head can be performed in a few drops of Hoyer's or glycerol on a microscope slide or in water in an excavated glass block. Turn the head ventral side up and hold it to the glass with a blunt no. 1 insect pin. Remove the mandibles, maxillae and labial palpi using jeweller's forceps and micropins. Remove the epipharynx and antennae. Mount the head capsule and the mandibles in the cavity of the slide using Hoyer's. Mount the cleared skin, fully opened on the flat part of the microscope slide, next to the cavity. It is usually best done ventral side up. Epipharynx, antennae, maxillae and labial palpi should be mounted with the skin under the same cover slip. Mount all body parts on the same microscope slide. [New paragraph]-In the case of larval exuviae, before proceeding with the dissection soak the specimen in a 5% solution of any laboratory detergent (e.g. Decon 90) for about two hours and thoroughly rinse in distilled water. Cut the specimen open anteriorly and dissect out the mouthparts. They can be mounted directly in Hoyer's without clearing. Label slides immediately after mounting specimens and place them in an oven for a few days at 40 °C. After drying, ring the slides using any lacquer recommended for sealing of microscopic slides (e.g. ~~Glyptol~~Glyptal, Brunseal), or at least two layers of nail polish in order to prevent the Hoyer's from drying and possibly damaging the specimen.	First sentence: Without adding a comma, non native speakers can read "at first place" as "at first location". 17th sentence: It would be clearer to begin with a new paragraph (i.e. one paragraph for larvae, and one paragraph for larval exuviae). Last sentence: correct name.	European Union

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
[163]	45	Editorial	At first place the specimen ventral side up on a microscope slide. a) Cut open the whole body along the mid-line from under the head capsule to the last abdominal segment using eye surgery scissors. b) Next put the larva into a test-tube containing 10% potassium hydroxide (KOH) solution and heat in a boiling water bath until it has become clear. c) Rinse thoroughly in warm distilled water. d) Remove all internal tissues using a very fine, short hair brush or the convex surface of a hooked tip of a no. 1 insect pin, or a loop formed from a micropin. All setae should be removed from one side of the 7th and 8th abdominal segment. e) Remove the head capsule and put it back in the hot KOH solution for 5 minutes. Rinse the head capsule in warm distilled water. Dissection of the head can be performed in a few drops of Hoyer's or glycerol on a microscope slide or in water in an excavated glass block. Turn the head ventral side up and hold it to the glass with a blunt no. 1 insect pin. f) Remove the mandibles, maxillae and labial palpi using jeweller's forceps and micropins. Remove the epipharynx and. Mount the head capsule and the mandibles in the cavity of the slide using Hoyer's. Mount the cleared skin, fully opened on the flat part of the microscope slide, next to the cavity. It is usually best done ventral side up. Epipharynx, antennae, maxillae and labial palpi should be mounted with the skin under the same cover slip. Mount all body parts on the same microscope slide. g) In the case of larval exuviae, before proceeding with the dissection soak the specimen in a 5% solution of any laboratory detergent (e.g. Decon 90) for about two hours and thoroughly rinse in distilled water. Cut the specimen open anteriorly and dissect out the mouthparts. They can be mounted directly in Hoyer's without clearing. Label slides immediately after mounting specimens and place them in an oven for a few days at 40 °C. After drying, ring the slides using any lacquer recommended for sealing of microscopic slides (e.g. Glyptol, Brunseal), or at least two layers of nail polish in order to prevent the Hoyer's from drying and possibly damaging the specimen.	To clarify bullets were added	Argentina
[164]	45	Substantive	At first place the specimen ventral side up on a microscope slide. Cut open the whole body along the mid-line from under the head capsule to the last abdominal segment using eye surgery scissors. Next put the larva into a test-tube containing 10% potassium hydroxide (KOH) solution and heat in	"It is usually best done ventral side up.": We think the author should add why this is the case because it is not evident. Perhaps the antecostal sutures are more visible from beneath than from	United States of America

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			a boiling water bath until it has become clear. Rinse thoroughly in warm distilled water. Remove all internal tissues using a very fine, short hair brush or the convex surface of a hooked tip of a no. 1 insect pin, or a loop formed from a micropin. All setae should be removed from one side of the 7th and 8th abdominal segment. Remove the head capsule and put it back in the hot KOH solution for 5 minutes. Rinse the head capsule in warm distilled water. Dissection of the head can be performed in a few drops of Hoyer's or glycerol on a microscope slide or in water in an excavated glass block. Turn the head ventral side up and hold it to the glass with a blunt no. 1 insect pin. Remove the mandibles, maxillae and labial palpi using jeweller's forceps and micropins. Remove the epipharynx and antennae. Mount the head capsule and the mandibles in the cavity of the slide using Hoyer's. Mount the cleared skin, fully opened on the flat part of the microscope slide, next to the cavity. It is usually best done ventral side up. Epipharynx, antennae, maxillae and labial palpi should be mounted with the skin under the same cover slip. Mount all body parts on the same microscope slide. In the case of larval exuviae, before proceeding with the dissection soak the specimen in a 5% solution of any laboratory detergent (e.g. Decon 90) for about two hours and thoroughly rinse in distilled water. Cut the specimen open anteriorly and dissect out the mouthparts. They can be mounted directly in Hoyer's without clearing. Label slides immediately after mounting specimens and place them in an oven for a few days at 40 °C. After drying, ring the slides using any lacquer recommended for sealing of microscopic slides (e.g. Glyptol, Brunseal), or at least two layers of nail polish in order to prevent the Hoyer's from drying and possibly damaging the specimen.	above, but that is speculative on our part. If the author has no specific reason, then perhaps this sentence should be omitted.	
[165]	45	Technical	At first place the specimen ventral side up on a microscope slide. Cut open the whole body along the mid-line from under the head capsule to the last abdominal segment using eye surgery scissors. Next put the larva into a test-tube containing 10% potassium hydroxide (KOH) solution and heat in a boiling water bath until it has become clear. Rinse thoroughly in warm distilled water. Remove all internal tissues using a very fine, short hair brush or the convex surface of a hooked tip of a no. 1 insect pin, or a loop formed from a micropin. All setae should be removed from one side of the 7th and 8th abdominal segment and stained with fuchsin acid or chlorazol black. Remove the head capsule and put it back in the hot KOH solution for 5 minutes. Rinse the head capsule in warm distilled water. Dissection of the head can be performed in a few drops of Hoyer's or glycerol on a microscope slide or in water in an excavated glass block. Turn the head ventral side up and hold it to the glass with a blunt no. 1 insect pin. Remove the mandibles, maxillae and labial palpi using jeweller's forceps and micropins. Remove the epipharynx and antennae and stained with fuchsin acid or chlorazol black.	An stain was recommended because it makes the structures more visible. The use of brand names should be avoided as agreed by CPM.	Costa Rica ,Nicaragua ,El Salvador

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			Mount the head capsule and the mandibles in the cavity of the slide using Hoyer's. Mount the cleared skin, fully opened on the flat part of the microscope slide, next to the cavity. It is usually best done ventral side up. Epipharynx, antennae, maxillae and labial palpi should be mounted with the skin under the same cover slip. Mount all body parts on the same microscope slide. In the case of larval exuviae, before proceeding with the dissection soak the specimen in a 5% solution of any laboratory detergent ~~(e.g. Decon 90)~~ for about two hours and thoroughly rinse in distilled water. Cut the specimen open anteriorly and dissect out the mouthparts. They can be mounted directly in Hoyer's without clearing. Label slides immediately after mounting specimens and place them in an oven for a few days at 40 °C. After drying, ring the slides using any lacquer recommended for sealing of microscopic slides (e.g. Glyptol, Brunseal), or at least two layers of nail polish in order to prevent the Hoyer's from drying and possibly damaging the specimen.		
[166]	45	Technical	At first place the specimen ventral side up on a microscope slide. Cut open the whole body along the mid-line from under the head capsule to the last abdominal segment using eye surgery scissors. Next put the larva into a test-tube containing 10% potassium hydroxide (KOH) solution and heat in a boiling water bath until it has become clear. Rinse thoroughly in warm distilled water. Remove all internal tissues using a very fine, short hair brush or the convex surface of a hooked tip of a no. 1 insect pin, or a loop formed from a micropin. All setae should be removed from one side of the 7th and 8th abdominal segment and stained with fuchsin acid or chlorazol black. Remove the head capsule and put it back in the hot KOH solution for 5 minutes. Rinse the head capsule in warm distilled water. Dissection of the head can be performed in a few drops of Hoyer's or glycerol on a microscope slide or in water in an excavated glass block. Turn the head ventral side up and hold it to the glass with a blunt no. 1 insect pin. Remove the mandibles, maxillae and labial palpi using jeweller's forceps and micropins. Remove the epipharynx and antennae and stain with fuchsin acid or chlorazol black. Mount the head capsule and the mandibles in the cavity of the slide using Hoyer's. Mount the cleared skin, fully opened on the flat part of the microscope slide, next to the cavity. It is usually best done ventral side up. Epipharynx, antennae, maxillae and labial palpi should be mounted with the skin under the same cover slip. Mount all body parts on the same microscope slide. In the case of larval exuviae, before proceeding with the dissection soak the specimen in a 5% solution of any laboratory detergent ~~(e.g. Decon 90)~~ for about two hours and thoroughly rinse in distilled water. Cut the specimen open anteriorly and dissect out the mouthparts. They can be mounted directly in Hoyer's without clearing. Label slides immediately after mounting specimens and place them in an oven for a few	1) A stain was recommended because it makes the structures more visible. 2) The use of brand names should be avoided in diagnostic protocols as agreed by CPM.	Uruguay

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			days at 40 °C. After drying, ring the slides using any lacquer recommended for sealing of microscopic slides (e.g. Glyptol, Brunseal), or at least two layers of nail polish in order to prevent the Hoyer's from drying and possibly damaging the specimen.		
[167]	45	Technical	At first place the specimen ventral side up on a microscope slide. Cut open the whole body along the mid-line from under the head capsule to the last abdominal segment using eye surgery scissors. Next put the larva into a test-tube containing 10% potassium hydroxide (KOH) solution and heat in a boiling water bath until it has become clear. Rinse thoroughly in warm distilled water. Remove all internal tissues using a very fine, short hair brush or the convex surface of a hooked tip of a no. 1 insect pin, or a loop formed from a micropin. All setae should be removed from one side of the 7th and 8th abdominal segment and stained with acid fuchsin and or chlorazol black. Remove the head capsule and put it back in the hot KOH solution for 5 minutes. Rinse the head capsule in warm distilled water. Dissection of the head can be performed in a few drops of Hoyer's or glycerol on a microscope slide or in water in an excavated glass block. Turn the head ventral side up and hold it to the glass with a blunt no. 1 insect pin. Remove the mandibles, maxillae and labial palpi using jeweller's forceps and micropins. Remove the epipharynx and antennae and stain with acid fuchsin and or chlorazol black. Mount the head capsule and the mandibles in the cavity of the slide using Hoyer's. Mount the cleared skin, fully opened on the flat part of the microscope slide, next to the cavity. It is usually best done ventral side up. Epipharynx, antennae, maxillae and labial palpi should be mounted with the skin under the same cover slip. Mount all body parts on the same microscope slide. In the case of larval exuviae, before proceeding with the dissection soak the specimen in a 5% solution of any laboratory detergent ~~(e.g. Decon 90)~~ for about two hours and thoroughly rinse in distilled water. Cut the specimen open anteriorly and dissect out the mouthparts. They can be mounted directly in Hoyer's without clearing. Label slides immediately after mounting specimens and place them in an oven for ~~a few~~ at least three days at 40 °C. After drying, ring the slides using any lacquer recommended for sealing of microscopic slides (e.g. Glyptol, Brunseal), or at least two layers of nail polish in order to prevent the Hoyer's from drying and possibly damaging the specimen.	Added text "and stained with acid fuchsin and or chlorazol black" and "and stain with acid fuchsin and or chlorazol black": An stain was recommended because it makes the structures more visible than without stain. Deleted text "(e.g. Decon 90)": The use of brand name should be avoided as agreed. Added text "at least three": To specify the minimum time required.	COSAVE,Paraguay ,Chile,Brazil
[168]	45	Technical	At first place the specimen ventral side up on a microscope slide. Cut open the whole body along the mid-line from under the head capsule to the last abdominal segment using eye surgery scissors. Next put the larva into a test-tube containing 10% potassium hydroxide (KOH) solution and heat in a boiling water bath until it has become clear. Rinse thoroughly in warm distilled water. Remove all internal tissues using a very fine, short hair brush or the convex surface of a hooked tip of a no. 1 insect pin, or a loop formed from a micropin. All setae should be removed from one side of the	The use of brand names should be avoided as agree by CPM.	Mexico

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			7th and 8th abdominal segment. Remove the head capsule and put it back in the hot KOH solution for 5 minutes. Rinse the head capsule in warm distilled water. Dissection of the head can be performed in a few drops of Hoyer's or glycerol on a microscope slide or in water in an excavated glass block. Turn the head ventral side up and hold it to the glass with a blunt no. 1 insect pin. Remove the mandibles, maxillae and labial palpi using jeweller's forceps and micropins. Remove the epipharynx and antennae. Mount the head capsule and the mandibles in the cavity of the slide using Hoyer's. Mount the cleared skin, fully opened on the flat part of the microscope slide, next to the cavity. It is usually best done ventral side up. Epipharynx, antennae, maxillae and labial palpi should be mounted with the skin under the same cover slip. Mount all body parts on the same microscope slide. In the case of larval exuviae, before proceeding with the dissection soak the specimen in a 5% solution of any laboratory detergent ~~(e.g. Decon 90)~~ for about two hours and thoroughly rinse in distilled water. Cut the specimen open anteriorly and dissect out the mouthparts. They can be mounted directly in Hoyer's without clearing. Label slides immediately after mounting specimens and place them in an oven for a few days at 40 °C. After drying, ring the slides using any lacquer recommended for sealing of microscopic slides (e.g. Glyptol, Brunseal), or at least two layers of nail polish in order to prevent the Hoyer's from drying and possibly damaging the specimen.		
[169]	45	Technical	At first place the specimen ventral side up on a microscope slide. a) Cut open the whole body along the mid-line from under the head capsule to the last abdominal segment using eye surgery scissors. b) Next put the larva into a test-tube containing 10% potassium hydroxide (KOH) solution and heat in a boiling water bath until it has become clear. c) Rinse thoroughly in warm distilled water. d) Remove all internal tissues using a very fine, short hair brush or the convex surface of a hooked tip of a no. 1 insect pin, or a loop formed from a micropin. All setae should be removed from one side of the 7th and 8th abdominal segment and stained with acid fuchsin and or chlorazol black. e) Remove the head capsule and put it back in the hot KOH solution for 5 minutes. Rinse the head capsule in warm distilled water. Dissection of the head can be performed in a few drops of Hoyer's or glycerol on a microscope slide or in water in an excavated glass block. Turn the head ventral side up and hold it to the glass with a blunt no. 1 insect pin. f) Remove the mandibles, maxillae and labial palpi using jeweller's	An stain was recommended because it makes the structures more visible than without stain	Argentina

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			forceps and micropins. Remove the epipharynx and antennae and stain with acid fuchsin and or chlorazol black. Mount the head capsule and the mandibles in the cavity of the slide using Hoyer's. Mount the cleared skin, fully opened on the flat part of the microscope slide, next to the cavity. It is usually best done ventral side up. Epipharynx, antennae, maxillae and labial palpi should be mounted with the skin under the same cover slip. Mount all body parts on the same microscope slide. g) In the case of larval exuviae, before proceeding with the dissection soak the specimen in a 5% solution of any laboratory detergent ~~(e.g. Decon 90)~~ for about two hours and thoroughly rinse in distilled water. Cut the specimen open anteriorly and dissect out the mouthparts. They can be mounted directly in Hoyer's without clearing. Label slides immediately after mounting specimens and place them in an oven for ~~a few~~ at least three days at 40 °C. After drying, ring the slides using any lacquer recommended for sealing of microscopic slides (e.g. Glyptol, Brunseal), or at least two layers of nail polish in order to prevent the Hoyer's from drying and possibly damaging the specimen.		
[170]	45	Technical	a) At first place the specimen ventral side up on a microscope slide~~.~~; b) Cut open the whole body along the mid-line from under the head capsule to the last abdominal segment using eye surgery scissors~~.~~; c) Next put the larva into a test-tube containing 10% potassium hydroxide (KOH) solution and heat in a boiling water bath until it has become clear~~.~~; d) Rinse thoroughly in warm distilled water~~.~~; e) Remove all internal tissues using a very fine, short hair brush or the convex surface of a hooked tip of a no. 1 insect pin, or a loop formed from a micropin. All setae should be removed from one side of the 7th and 8th abdominal segment and stained with fuchsin acid and or chlorazol black~~.~~; f) Remove the head capsule and put it back in the hot KOH solution for 5 minutes. Rinse the head capsule in warm distilled water. Dissection of the head can be performed in a few drops of Hoyer's or glycerol on a microscope slide or in water in an excavated glass block. Turn the head ventral side up and hold it to the glass with a blunt no. 1 insect pin~~.~~; g) Remove the mandibles, maxillae and labial palpi using jeweller's forceps and micropins. Remove the epipharynx and antennae and stain with fuchsin acid or chlorazol black. Mount the head capsule and the mandibles in the cavity of the slide using Hoyer's. Mount the cleared skin, fully opened on the flat part of the microscope slide, next to the cavity. It is usually best done ventral side up. Epipharynx, antennae, maxillae and labial palpi should be mounted with the skin under the same cover slip.	1) To clarify the procedure for preparation of the larvae, the paragraph was separated using bullets 2) A stain was recommended because it makes the structures more visible 3) A stain was recommended because it makes the structures more visible 4) The use of brand names should be avoided as agreed by CPM.	OIRSA

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			Mount all body parts on the same microscope slide~~.~~; h) In the case of larval exuviae, before proceeding with the dissection soak the specimen in a 5% solution of any laboratory detergent ~~(e.g. Decon 90)~~ for about two hours and thoroughly rinse in distilled water. Cut the specimen open anteriorly and dissect out the mouthparts. They can be mounted directly in Hoyer's without clearing. Label slides immediately after mounting specimens and place them in an oven for a few days at 40 °C. After drying, ring the slides using any lacquer recommended for sealing of microscopic slides (e.g. Glyptol, Brunseal), or at least two layers of nail polish in order to prevent the Hoyer's from drying and possibly damaging the specimen.		
[171]	47	Editorial	~~The identification should be performed using a high-powered (at least 400×) compound microscope. Depending on the quality of the microscope, oil immersion may need to be used to achieve satisfactory resolution.~~	Text is repeated in pargraph 40.	Costa Rica ,Uruguay ,Mexico ,Nicaragua ,El Salvador
[172]	47	Editorial	~~The identification should be performed using a high-powered (at least 400×) compound microscope. Depending on the quality of the microscope, oil immersion may need to be used to achieve satisfactory resolution.~~	It is repeated in para 40.	COSAVE,Paraguay ,Chile,Brazil
[173]	47	Editorial	~~The identification should be performed using a high-powered (at least 400×) compound microscope. Depending on the quality of the microscope, oil immersion may need to be used to achieve satisfactory resolution.~~	Is repeated in para 40	Argentina
[174]	47	Editorial	~~The identification should be performed using a high-powered (at least 400×) compound microscope. Depending on the quality of the microscope, oil immersion may need to be used to achieve satisfactory resolution.~~	Text is repeated in paragraph 40	OIRSA
[175]	49	Editorial	Adult *Trogoderma* specimens may need to be cleaned before identification, with any laboratory detergent or by using an ultrasonic cleaner. If the specimen was caught in a sticky trap the glue can be dissolved using a number of solvents (e.g., kerosene). These solvents can be removed from the specimen by any laboratory detergent.	'e.g. kerosene': Placing parentheses around this will make it consistent with prior usage in this document.	United States of America
[176]	50	Editorial	Before beginning the preparation soak the adult in warm distilled water for about an hour. a) ~~Next~~ First remove abdomen while the specimen is still in the water using fine forceps. Dry the specimen and mount it on a cardboard rectangle, preferably laterally. (Gluing it on the side makes the specimen less exposed to damage and accessible for both dorsal and ventral examination.) Next cut the abdomen laterally open, leaving the last abdominal segment untouched. Place it in 10% KOH or sodium hydroxide (NaOH) solution in a hot water bath for about 10 minutes.	To clarify the procedure the paragraph was separated using bullets.	Uruguay

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			b) Rinse the specimen in water and carefully remove the genitalia using hooked micropins. The abdomen should be glued onto the same cardboard rectangle with the insect, ventral side facing up. c) Usually the genitalia need to be macerated further in the caustic solution. Separate the aedeagus from the periphallic tergum and the 9th abdominal segment using micropins.		
[177]	50	Editorial	Before beginning the preparation soak the adult in warm distilled water for about an hour. a) ~~Next~~ First remove abdomen while the specimen is still in the water using fine forceps. Dry the specimen and mount it on a cardboard rectangle, preferably laterally. (Gluing it on the side makes the specimen less exposed to damage and accessible for both dorsal and ventral examination.) Next cut the abdomen laterally open, leaving the last abdominal segment untouched. Place it in 10% KOH or sodium hydroxide (NaOH) solution in a hot water bath for about 10 minutes. b) Rinse the specimen in water and carefully remove the genitalia using hooked micropins. The abdomen should be glued onto the same cardboard rectangle with the insect, ventral side facing up. c) Usually the genitalia need to be macerated further in the caustic solution. Separate the aedeagus from the periphallic tergum and the 9th abdominal segment using micropins.	To clarify bullets were added.	COSAVE,Paraguay ,Argentina ,Chile,Brazil
[178]	50	Editorial	Before beginning the preparation soak the adult in warm distilled water for about an hour. Next remove abdomen while the specimen is still in the water using fine forceps. Dry the specimen (minus abdomen) and mount it on a cardboard rectangle, preferably laterally. (Gluing it on the side makes the specimen less exposed to damage and accessible for both dorsal and ventral examination.) Next cut the abdomen laterally open, leaving the last abdominal segment untouched. Place it in 10% KOH or sodium hydroxide (NaOH) solution in a hot water bath for about 10 minutes. Rinse the specimen in water and carefully remove the genitalia using hooked micropins. After removing the genitalia, ~~T~~the abdomen should be glued onto the same cardboard rectangle with the insect, ventral side facing up. Usually the genitalia need to be macerated further in the caustic solution. Separate the aedeagus from the periphallic tergum and the 9th abdominal segment using micropins.		New Zealand
[179]	50	Editorial	Before beginning the preparation soak the adult in warm distilled water for about an hour. ~~Next~~Using fine forceps, remove the abdomen while the specimen is still in the water ~~using fine forceps~~. Dry the specimen and mount it on a cardboard rectangle, preferably laterally. (Gluing it on the side makes the specimen less exposed to damage and accessible for both dorsal and ventral examination.) Next cut the abdomen ~~laterally~~	In both cases, changes are recommended to improve comprehension.	United States of America

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			open laterally, leaving the last abdominal segment untouched. Place it in 10% KOH or sodium hydroxide (NaOH) solution in a hot water bath for about 10 minutes. Rinse the specimen in water and carefully remove the genitalia using hooked micropins. The abdomen should be glued onto the same cardboard rectangle with the insect, ventral side facing up. Usually the genitalia need to be macerated further in the caustic solution. Separate the aedeagus from the periphallic tergum and the 9th abdominal segment using micropins.		
[180]	50	Editorial	Before beginning the preparation soak the adult in warm distilled water for about an hour. Next remove abdomen while the specimen is still in the water using fine forceps. Dry the specimen and mount it on a cardboard rectangle, preferably laterally. ~~(~~Gluing it on the side makes the specimen less exposed to damage and accessible for both dorsal and ventral examination.~~)~~ Next cut the abdomen laterally open, leaving the last abdominal segment untouched. Place it in 10% KOH or sodium hydroxide (NaOH) solution in a hot water bath for about 10 minutes. Rinse the specimen in water and carefully remove the genitalia using hooked micropins. The abdomen should be glued onto the same cardboard rectangle with the insect, ventral side facing up. Usually the genitalia need to be macerated further in the caustic solution. Separate the aedeagus from the periphallic tergum and the 9th abdominal segment using micropins.	The use of brackets is unnecessary and the sentence should stand on its own	South Africa
[181]	50	Technical	Before beginning the preparation soak the adult in warm distilled water for about an hour. Next remove abdomen while the specimen is still in the water using fine forceps. Dry the specimen and mount it on a cardboard rectangle, preferably laterally. (Gluing it on the side makes the specimen less exposed to damage and accessible for both dorsal and ventral examination.) Next cut the abdomen laterally open, leaving the last abdominal segment untouched. Place it in 10% KOH or sodium hydroxide (NaOH) solution in a hot water bath for about 10 minutes. Rinse the specimen in water and carefully remove the genitalia using hooked micropins. The abdomen should be glued onto the same cardboard rectangle with the insect, ventral side facing up. ~~Usually t~~The genitalia need to be macerated further in the caustic solution. Separate the aedeagus from the periphallic tergum and the 9th abdominal segment using micropins. Stain with fuchsin acid or chorazol black.	Genitalia always needs to be macerated. A stain was recomended because it makes the structures more visible.	Costa Rica ,Nicaragua ,El Salvador
[182]	50	Technical	Before beginning the preparation soak the adult in warm distilled water for about an hour. Next remove abdomen while the specimen is still in the water using fine forceps. Dry the specimen and mount it on a cardboard rectangle, preferably laterally. (Gluing it on the side makes the specimen less exposed to damage and accessible for both dorsal and ventral examination.) Next cut the abdomen laterally open, leaving the last	1) The word usually was deleted because the genitalia needs always to be macerated. 2) Astain was recommended because it makes the structures more visibles.	Uruguay

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			abdominal segment untouched. Place it in 10% KOH or sodium hydroxide (NaOH) solution in a hot water bath for about 10 minutes. Rinse the specimen in water and carefully remove the genitalia using hooked micropins. The abdomen should be glued onto the same cardboard rectangle with the insect, ventral side facing up. ~~Usually t~~The genitalia need to be macerated further in the caustic solution. Separate the aedeagus from the periphallic tergum and the 9th abdominal segment using micropins. Stain with fuchsin acid or chlorazol black. ~~.~~		
[183]	50	Technical	Before beginning the preparation soak the adult in warm distilled water for about an hour. Next remove abdomen while the specimen is still in the water using fine forceps. Dry the specimen and mount it on a cardboard rectangle, preferably laterally. (Gluing it on the side makes the specimen less exposed to damage and accessible for both dorsal and ventral examination.) Next cut the abdomen laterally open, leaving the last abdominal segment untouched. Place it in 10% KOH or sodium hydroxide (NaOH) solution in a hot water bath for about 10 minutes. Rinse the specimen in water and carefully remove the genitalia using hooked micropins. The abdomen should be glued onto the same cardboard rectangle with the insect, ventral side facing up. ~~Usually t~~The genitalia need to be macerated further in the caustic solution. Separate the aedeagus from the periphallic tergum and the 9th abdominal segment using micropins. Stain with acid fuchsin and or chlorazol black.	The genitalia needs always be macerated. Added text "Stain with acid fuchsin and or chlorazol black": An stain was recommended because it makes the structures more visible	COSAVE,Paraguay ,Argentina ,Chile,Brazil
[184]	50	Technical	Before beginning the preparation soak the adult in warm distilled water for about an hour. Next remove abdomen while the specimen is still in the water using fine forceps. Dry the specimen and mount it on a cardboard rectangle, preferably laterally. (Gluing it on the side makes the specimen less exposed to damage and accessible for both dorsal and ventral examination.) Next cut the abdomen laterally open, leaving the last abdominal segment untouched. Place it in 10% KOH or sodium hydroxide (NaOH) solution in a hot water bath for about 10 minutes. Rinse the specimen in water and carefully remove the genitalia using hooked micropins. The abdomen should be glued onto the same cardboard rectangle with the insect, ventral side facing up. ~~Usually t~~The genitalia need to be macerated further in the caustic solution. Separate the aedeagus from the periphallic tergum and the 9th abdominal segment using micropins. Stain with fuchsin acid or chlorazol black.	1) The genitalia always need to be macerated. 2) A stain was recommended because it makes the structures more visible	OIRSA
[185]	51	Substantive	Genitalia can be mounted on a microscope slide using Hoyer's mounting medium or PVA (Polyvinyl alcohol). The aedeagus should be mounted on a cavity microscope slide so it is able to retain its shape. Female genitalia can be mounted on a flat microscope slide.		Indonesia
[186]	52	Editorial	Slides and pinned insects should be labelled immediately after mounting	To clarify.	Costa Rica

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			the specimens. The slides should be placed in an oven for a few days at 40ºC. After drying all slides should be ringed (see 4.1h).		,Nicaragua ,El Salvador
[187]	52	Editorial	Slides and pinned insects should be labelled immediately after mounting the specimens. The slides should be placed in an oven for a few days at 40ºC. After drying all slides should be ringed (see 4.1 h).	To clarify, according changes proposed in paragraph 45.	Uruguay
[188]	52	Editorial	Slides and pinned insects should be labelled immediately after mounting the specimens. The slides should be placed in an oven for a few days at 40ºC. After drying all slides should be ringed (see 4.1g).	To clarify	COSAVE,Paraguay ,Chile,Brazil
[189]	52	Editorial	Slides and pinned insects should be labelled immediately after mounting the specimens. The slides should be placed in an oven for a few days at 40ºC. After drying all slides should be ringed (see 4.1g).	To clarify	Argentina
[190]	52	Editorial	Slides and pinned insects should be labelled immediately after mounting the specimens. The slides should be placed in an oven for a few days at 40ºC. After drying all slides should be ringed (see 4.1 h).	To clarify.	OIRSA
[191]	52	Technical	Slides and pinned insects should be labelled immediately after mounting the specimens. The slides should be placed in an oven for ~~a few~~ at least three days at 40ºC. After drying all slides should be ringed (see 4.1).	To specify the minimum time required.	COSAVE,Paraguay ,Chile,Brazil
[192]	52	Technical	Slides and pinned insects should be labelled immediately after mounting the specimens. The slides should be placed in an oven for a few days at 40ºC. After drying all slides should be ringed (see 4.1).	Second sentence in paragraph 52 refer that .. "The slides should be placed in an oven for a few days at 40ºC. .. Would be useful for all countries to know how many days will be necesary to obtain better results.	Mexico
[193]	52	Technical	Slides and pinned insects should be labelled immediately after mounting the specimens. The slides should be placed in an oven for a ~~few~~ at least three days at 40ºC. After drying all slides should be ringed (see 4.1).	To specify the minimum time required .	Argentina
[194]	53	Editorial	If there is no need for mounting the genitalia using a permanent or semi-permanent mounting agent, they can be examined in a drop of glycerol on a microscope slide. After the identification the organs can be placed in a microvial in a drop of glycerol or glued onto the cardboard rectangle next to the abdomen.	Comma after introductory clause.	United States of America
[195]	54	Editorial	**4.3 Genera of the family Dermestidae frequently occurring in stored commodities** If the diagnostic key being used was not specifically written to include the area of origin (and interception) of the specimen/s, the key should be used with caution as there are many undescribed species of Dermestidae worldwide.	clarification	Australia

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
[196]	55	Editorial	Besides *Trogoderma*, other dermestid genera may also be found in stored products, such as *Anthrenus*, *Anthrenocerus*, *Attagenus* and *Dermestes*. The first step of diagnosis of collected specimens is identification to genus. Adults of these beetles, and in some cases larvae, ~~of these beetles~~ can be identified using at least one of the keys of Mound (1989), Kingsolver (1991), Haines (1991), Banks (1994), Rees (2004) and~~/or~~ Háva (2004). Genera of the North American Dermestidae can be identified using the key of Kingsolver (2002).	Prepositional phrase describing the noun 'adults' needs to follow the noun. The use of 'at least one of the keys of' obviates the need for '/or' later on.	United States of America
[197]	56	Editorial	The following simple keys quickly enable *Trogoderma* to be distinguished from four other dermestid genera commonly occurring in stored commodities. Distinguishing characters are described in section 4.5.3 and illustrated in section 9, Figures 1 to 16.	For better reading and follow up of the protocol.	Costa Rica ,Uruguay ,Nicaragua ,El Salvador
[198]	56	Editorial	The following simple keys quickly enable *Trogoderma* to be distinguished from four other dermestid genera commonly occurring in stored commodities. Distinguishing characters are described in section 4.5.3 and illustrated in section 9, Figures 1 to 16.	For better reading and follow the protocol.	COSAVE,Paraguay ,Chile,Brazil
[199]	56	Editorial	The following simple keys quickly enable *Trogoderma* to be distinguished from four other dermestid genera commonly occurring in stored commodities. Distinguishing characters are described in section 4.5.3 and illustrated in section 9, Figures 1 to 16.	For better reading and follow the protocol	Argentina
[200]	56	Editorial	The following simple keys quickly enable *Trogoderma* to be distinguished from four other dermestid genera commonly occurring in stored commodities. Distinguishing characters are described in section 4.5.3 and illustrated in section 9, Figures 1 to 16.	For better reading and follow up of the protocol	OIRSA
[201]	56	Substantive	The ~~following~~ simple keys in 4.3.1 and 4.6.1 quickly enable ~~*Trogoderma* to be distinguished from~~ four other dermestid genera commonly occurring in stored commodities to be distinguished from *Trogoderma*. Distinguishing characters are illustrated in section 9, Figures 1 to 16.	1. Modified because there is a need to refer to where the simple keys are (consequent change from moving all information on larvae and adults together) 2. Wording is confusing as it implies that the key is all that is required to identify Trogoderma spp, whereas the distinguishing features in (former) 4.4.2 are required	EPPO,Russian Federation ,Ukraine ,Morocco ,Uzbekistan
[202]	56	Substantive	The ~~following~~ simple keys in 4.3.1 and 4.6.1 quickly enable ~~*Trogoderma* to be distinguished from~~ four other dermestid genera commonly occurring in stored commodities to be distinguished from *Trogoderma*. Distinguishing characters are illustrated in section 9, Figures 1 to 16.	1. Modified because there is a need to refer to where the simple keys are (consequent change from moving all information on larvae and adults together) 2. Wording is confusing as it implies that the key is all that is required to identify Trogoderma spp, whereas the distinguishing features in (former) 4.4.2 are required	European Union

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
[203]	56	Technical	The following simple keys quickly enable *Trogoderma* to be distinguished from four other dermestid genera commonly occurring in stored commodities. Distinguishing characters are illustrated in section 9, Figures 1 to 16.	Question to author regarding differentiation of adults: the key of Delobel and Tran includes three genus closed to Trogoderma, i.e. Thaumaglossa, Orphinus and Phradonoma, which can be found in stores. Is is sure they are eliminated in the key? The main character allowing to separate them is at antennal level (antenna which frequently disappear in stored conditions).	EPPO,Russian Federation ,Ukraine ,Morocco ,Uzbekistan
[204]	56	Technical	The following simple keys quickly enable *Trogoderma* to be distinguished from four other dermestid genera commonly occurring in stored commodities. Distinguishing characters are illustrated in section 9, Figures 1 to 16.	Question to author regarding differentiation of adults: the key of Delobel and Tran includes three genus closed to Trogoderma, i.e. Thaumaglossa, Orphinus and Phradonoma, which can be found in stores. Is is sure they are eliminated in the key? The main character allowing to separate them is at antennal level (antenna which frequently disappear in stored conditions).	European Union
[205]	57	Editorial	**4.3.1 Larvae**	It is preferable to present the adult key first followed by larval key, though larvae are more commonly encountered.	New Zealand
[206]	57	Substantive	**4.3.1 Simple key for differentiation of dermestid ~~L~~larvae**	Consider changing title to reflect contents (consequence of moving former section 4.3.2)	EPPO,Russian Federation ,European Union ,Ukraine ,Morocco ,Uzbekistan
[207]	58	Editorial	(1) Urogomphi present on 9th abdominal segment, 10th segment sclerotized, cylindrical **Dermestes spp.**	Dermestes should be in italics.	EPPO,Russian Federation ,Ukraine ,Morocco ,Uzbekistan
[208]	58	Editorial	(1) Urogomphi present on 9th abdominal segment, 10th segment sclerotized, cylindrical **Dermestes spp.**	Italize Dermestes	Kenya
[209]	58	Editorial	(1) Urogomphi present on 9th abdominal segment, 10th segment sclerotized, cylindrical ~~**Dermestes**~~ ***Dermestes*** **spp.**	All genera in the keys are italicized and bolded.	United States of America
[210]	58	Editorial	(1) Urogomphi present on 9th abdominal segment, 10th segment sclerotized, cylindrical **Dermestes spp.**	Dermestes should be in italics.	European Union

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
[211]	60	Editorial	(2) Dorsal surface without hastisetae, maxillary palp 4-segmented ***Attagenus* spp.**	It is good to add more diagnostic images through out the keys for adult & larval characters.	New Zealand
[212]	65	Substantive	Second and last antennal segments subequal, head of hastisetae less than three times as long as wide at widest point ***Trogoderma* spp.** Larval or exuvial specimens having the above characteristics are very likely to be Trogoderma species and therefore it is warranted to check the detailed list of features listed in section 4.4.1.	New paragraph to ensure that diagnosticians do not stop at paragraph 65. Wording adapted from paragraph 78. (Numbering changed to reflect proposal to delete paragraphs 77-79)	EPPO,Russian Federation ,Ukraine ,Morocco ,Uzbekistan
[213]	65	Substantive	Second and last antennal segments subequal, head of hastisetae less than three times as long as wide at widest point ***Trogoderma* spp.** Larval or exuvial specimens having the above characteristics are very likely to be Trogoderma species and therefore it is warranted to check the detailed list of features listed in section 4.4.1.	New paragraph to ensure that diagnosticians do not stop at paragraph 65. Wording adapted from paragraph 78. (Numbering changed to reflect proposal to delete paragraphs 77-79). (Note: Trogoderma should be in italics).	European Union
[214]	66	Editorial	**4.3.2 Adults** insert explanation of different types of setae	Suggest that somewhere there is an explanation of the different types of setae; along with a diagram for each setae explaining the components for clarification and assistance with interpreting the key – there is a high level of assumed knowledge	Australia
[215]	66	Editorial	**4.3.2 Adults** [67]-[73] all genus Latin name should be Italic in format	Format consistence in text of standard.	China
[216]	66	Substantive	~~**4.3.2 Adults**~~	Move to after paragraph 97 so all information on identification of adults is together	EPPO,Russian Federation ,European Union ,Ukraine ,Morocco ,Uzbekistan
[217]	67	Editorial	(1) Median ocellus absent **Dermestes spp.** (Figure 10)	Scientific name should be in italics.	Thailand ,Korea, Republic of ,Lao People's Democratic Republic,Japan ,India
[218]	67	Editorial	(1) Median ocellus absent **Dermestes spp.** (Figure 10)	Italize Dermestes.	Kenya
[219]	67	Editorial	(1) Median ocellus absent ~~**Dermestes**~~ ***Dermestes*** **spp.** (Figure 10)	All genera in keys are italicized and written in bold font.	United States of America
[220]	67	Substantive	~~(1) Median ocellus absent **Dermestes spp.** (Figure 10)~~	Move to after paragraph 97 so all information on	EPPO,Russian

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
				identification of adults is together	Federation ,European Union ,Ukraine ,Morocco ,Uzbekistan
[221]	68	Substantive	~~Median ocellus present.....2~~	Move to after paragraph 97 so all information on identification of adults is together	EPPO,Russian Federation ,European Union ,Ukraine ,Morocco ,Uzbekistan
[222]	69	Editorial	(2) Body covered with scale-like setae; antennal cavity filled by antennae, fully visible from anterior view (Figure 9(A)) **Anthrenus spp.** (Figure 12)	Scientific name should be in italics.	Thailand ,Korea, Republic of ,Lao People's Democratic Republic,Japan ,India
[223]	69	Editorial	(2) Body covered with scale-like setae; antennal cavity filled by antennae, fully visible from anterior view (Figure 9(A)) **Anthrenus spp.** (Figure 12)	Italize Anthrenus.	Kenya
[224]	69	Editorial	(2) Body covered with scale-like setae; antennal cavity filled by antennae, fully visible from anterior view (Figure 9(A)) ~~Anthrenus~~*Anthrenus* **spp.** (Figure 12)	All genera in keys are italicized and written in bold font.	United States of America
[225]	69	Substantive	~~(2) Body covered with scale-like setae; antennal cavity filled by antennae, fully visible from anterior view (Figure 9(A)) **Anthrenus spp.** (Figure 12)~~	Move to after paragraph 97 so all information on identification of adults is together	EPPO,Russian Federation ,European Union ,Ukraine ,Morocco ,Uzbekistan
[226]	70	Substantive	~~Body covered with simple setae, some of them whitish, flattened (ensiform) but never scale-like....3~~	Move to after paragraph 97 so all information on identification of adults is together	EPPO,Russian Federation ,European Union ,Ukraine ,Morocco ,Uzbekistan
[227]	71	Editorial	(3) Antennal cavity completely closed behind, antennal club 3-segmented and well defined **Anthrenocerus spp.**	Scientific name should be in italics.	Thailand ,Korea, Republic of ,Lao People's Democratic Republic,Japan

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
					,India
[228]	71	Editorial	(3) Antennal cavity completely closed behind, antennal club 3-segmented and well defined **Anthrenocerus spp.**	Italize Anthrenocerus.	Kenya
[229]	71	Editorial	(3) Antennal cavity completely closed behind, antennal club 3-segmented and well defined ~~**Anthrenocerus**~~***Anthrenocerus*** **spp.**	All genera in keys are italicized and written in bold font.	United States of America
[230]	71	Substantive	~~(3) Antennal cavity completely closed behind, antennal club 3-segmented and well defined **Anthrenocerus spp.**~~	Move to after paragraph 97 so all information on identification of adults is together	EPPO,Russian Federation ,European Union ,Ukraine ,Morocco ,Uzbekistan
[231]	72	Editorial	Antennal cavity open behind or partially delimited by a posterior carina, ~~antennae~~antennal cavity much wider than antennae, not visible in anterior view (Figure 9(B))……..**4**	Adjective required, and to achieve consistency with the same phrase in the same key (e.g., paragraphs 73 and 74).	United States of America
[232]	72	Substantive	~~Antennal cavity open behind or partially delimited by a posterior carina, antennae cavity much wider than antennae, not visible in anterior view (Figure 9(B))……..**4**~~	Move to after paragraph 97 so all information on identification of adults is together	EPPO,Russian Federation ,European Union ,Ukraine ,Morocco ,Uzbekistan
[233]	73	Editorial	(4) Antennal cavity open behind, posterior margin of hind coxa angulate, first segment of posterior tarsus shorter than second segment **Attagenus spp.** (Figure 11)	Scientific name should be in italics.	Thailand ,Korea, Republic of ,Lao People's Democratic Republic,Japan ,India
[234]	73	Editorial	(4) Antennal cavity open behind, posterior margin of hind coxa angulate, first segment of posterior tarsus shorter than second segment **Attagenus spp.** (Figure 11)	Italize Attagenus	Kenya
[235]	73	Editorial	(4) Antennal cavity open behind, posterior margin of hind coxa angulate, first segment of posterior tarsus shorter than second segment ~~**Attagenus**~~***Attagenus*** **spp.** (Figure 11)	All genera in keys are italicized and written in bold font.	United States of America
[236]	73	Substantive	~~(4) Antennal cavity open behind, posterior margin of hind coxa angulate, first segment of posterior tarsus shorter than second segment **Attagenus spp.** (Figure 11)~~	Move to after paragraph 97 so all information on identification of adults is together	EPPO,Russian Federation ,European Union ,Ukraine ,Morocco

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
					,Uzbekistan
[237]	74	Substantive	~~Antennal cavity carinate posteriorly, posterior margin of hind coxa straight, arcuate or sinuate, first segment of posterior tarsus longer than second segment ***Trogoderma*** **spp.** (Figures 1(A), 3, 9(B)).~~	Move to after paragraph 97 so all information on identification of adults is together	EPPO,Russian Federation ,European Union ,Ukraine ,Morocco ,Uzbekistan
[238]	76	Editorial	~~Unfortunately, so far no key has been published for all known *Trogoderma* species.~~ There is no key as yet published that covers all known Trogoderma species. In part this is because there are still many undescribed species. Several keys have been published for the economically important species. Banks (1994) published a key to adults and larvae of the genus *Trogoderma* associated with stored products, as well as keys to larvae and adults of some species found in warehouses. Beal (1960) constructed an identification key to larvae of 14 species of *Trogoderma* from different parts of the world, including stored products pests. Mitsui (1967) published illustrated keys for identification of larvae and adults of some Japanese *Trogoderma* species. Kingsolver (1991) and Barak (1995) published keys to adults and larvae of some dermestid beetles, including a few *Trogoderma* species.	clarification	Australia
[239]	76	Editorial	Unfortunately, ~~so far~~ no key has been published for all known *Trogoderma* species. Several keys have been published for the economically important species. Banks (1994) published a key to adults and larvae of the genus *Trogoderma* associated with stored products, as well as keys to larvae and adults of some species found in warehouses. Beal (1960) constructed an identification key to larvae of 14 species of *Trogoderma* from different parts of the world, including stored product~~s~~ pests. Mitsui (1967) published illustrated keys for identification of larvae and adults of some Japanese *Trogoderma* species. Kingsolver (1991) and Barak (1995) published keys to adults and larvae of some dermestid beetles, including a few *Trogoderma* species.	'So far' is redundant. 'Stored product pests' is used throughout the document.	United States of America
[240]	77	Substantive	~~**4.4.1 Preliminary identification of *Trogoderma* larvae**~~	Suggest deletion of this section. Repeated in section 4.3.1 and paragraphs 80-82. Merge any relevant requirements with content in 80-82.	EPPO,Russian Federation ,European Union ,Ukraine ,Morocco ,Uzbekistan
[241]	78	Substantive	~~If all of the following features can be observed on the larva or exuviae it is very likely that the specimen is a *Trogoderma* species, and therefore it is warranted to check the detailed list of features listed in section 4.4.2:~~	Delete. Propose to cover the concept in section 4.3.1 (see comment on paragraph 65)	EPPO,Russian Federation ,Ukraine

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
					,Morocco ,Uzbekistan
[242]	78	Substantive	~~If all of the following features can be observed on the larva or exuviae it is very likely that the specimen is a *Trogoderma* species, and therefore it is warranted to check the detailed list of features listed in section 4.4.2:~~	Delete. Proposed to cover the concept in section 4.3.1 (see comment on paragraph 65)	European Union
[243]	79	Substantive	~~elongate, cylindrical, hairy larvae~~ ~~hastisetae present on sclerotized part of terga~~ ~~pretarsal setae on the ventral side of claws unequal~~ ~~antennal segments subequal.~~	Delete paragraph. Already covered in section 4.3.1 and paragraph 82. Merge any relevant requirements with content in 82	EPPO,Russian Federation ,Ukraine ,Morocco ,Uzbekistan
[244]	79	Substantive	elongate, ~~cylindrical,~~spindle, hairy larvae hastisetae present on sclerotized part of terga pretarsal setae on the ventral side of claws unequal antennal segments subequal.	According to observation and records, as well as refer to the larvae picture(B, C) in [160]. The shape oof larvae should be spindle. Reference paper: Zhang sheng Fang et al. The Identification of 8 important larva species in Genus Trogoderma, Plant Quarantine, 2007,Vol 21 (5):284-287	China
[245]	79	Substantive	~~elongate, cylindrical, hairy larvae~~ ~~hastisetae present on sclerotized part of terga~~ ~~pretarsal setae on the ventral side of claws unequal~~ ~~antennal segments subequal.~~	Delete paragraph. Already covered in section 4.3.1 and paragraph 82. Merge any relevant requirements with content in 82	European Union
[246]	80	Editorial	**4.4.~~2~~ 1 Discriminating features of *Trogoderma* larvae**	Numbering adjusted to take into account proposed deletion of paragraphs 77-79	EPPO,Russian Federation ,European Union ,Ukraine ,Morocco ,Uzbekistan
[247]	81	Editorial	Discriminating features of *Trogoderma* larvae below are adapted from Banks (1994), Beal (1954, 1960), Haines (1991), Hinton (1945), Kingsolver (1991), Lawrence (1991), Lawrence *et al.* (1999a), Okumura and Blanc (1955), Peacock (1993) and Rees (1943):	Order of citations needs to be consistent within the entire document.	United States of America
[248]	82	Editorial	(1) body elongated, cylindrical, somewhat flattened, roughly six times as long as wide, nearly parallel-sided but gradually tapering toward rear (2) head well developed, sclerotized, and hypognathous (3) three pairs of jointed legs present (4) pretarsal setae on the ventral side of claws unequal	clarification	Australia

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			(5) very hairy, being covered with different types of setae: hastisetae, spicisetae and/or fiscisetae (Figures 13 and 15) (6) head of setae: hastisetae not more than three times longer than wide (Figure 15) (7) numerous hastisetae on all nota and terga, with prominent tufts of erect hastisetae inserted on the posterolateral part of the tergal plates of abdominal segments 6 to 8 (in *Anthrenus* genus the tufts of hastisetae are inserted on the membrane behind the sclerotized part of terga 5, 6 and 7) (8) urogomphi absent.		
[249]	82	Editorial	(1) body elongated, cylindrical, somewhat flattened, roughly six times as long as wide, nearly parallel-sided but gradually tapering toward rear (2) head well developed, sclerotized, and hypognathous (3) three pairs of jointed legs present (4) pretarsal setae on the ventral side of claws unequal (5) very hairy, being covered with different types of setae: hastisetae, spicisetae and/or fiscisetae (Figures 13 and 15) (6) head of hastisetae not more than three times longer than wide (Figure 15) (7) numerous hastisetae on all nota and terga, with prominent tufts of erect hastisetae inserted on the posterolateral part of the tergal plates of abdominal segments 6 to 8 (in *Anthrenus* genus the tufts of hastisetae are inserted on the membrane behind the sclerotized part of terga 5, 6 and 7) (8) urogomphi absent.		New Zealand
[250]	82	Editorial	(1) body elongated, cylindrical, somewhat flattened, roughly six times as long as wide, nearly parallel-sided but gradually tapering toward rear (2) head well developed, sclerotized, and hypognathous (3) three pairs of jointed legs present (4) pretarsal setae on the ventral side of claws unequal (5) very hairy, being covered with different types of setae: hastisetae, spicisetae and/or fiscisetae (Figures 13 and 15) (6) head of hastisetae not more than three times longer than wide (Figure 15) (7) numerous hastisetae on all nota and terga, with prominent tufts of erect hastisetae inserted on the posterolateral part of the tergal plates of abdominal segments 6 to 8 (in *Anthrenus* genus the tufts of hastisetae are inserted on the membrane behind the sclerotized part of terga 5, 6 and 7) (8) urogomphi absent.	Insert colon in number (5)= Punctuation rule.	United States of America

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
[251]	82	Substantive	~~(1) body elongated, cylindrical, somewhat flattened, roughly six times as long as wide, nearly parallel sided but gradually tapering toward rear(2) head well developed, sclerotized, and hypognathous(3) three pairs of jointed legs present(4) pretarsal setae on the ventral side of claws unequal(5) very hairy, being covered with different types of setae hastisetae, spicisetae and/or fiscisetae (Figures 13 and 15)(6) head of hastisetae not more than three times longer than wide (Figure 15)(7) numerous hastisetae on all nota and terga, with prominent tufts of erect hastisetae inserted on the posterolateral part of the tergal plates of abdominal segments 6 to 8 (in *Anthrenus* genus the tufts of hastisetae are inserted on the membrane behind the sclerotized part of terga 5, 6 and 7)(8) urogomphi absent.~~ (1) body elongated, spindle, somewhat flattened, roughly six times as long as wide, nearly parallel-sided but gradually tapering toward rear (2) head well developed, sclerotized, and hypognathous (3) three pairs of jointed legs present (4) pretarsal setae on the ventral side of claws unequal (5) very hairy, being covered with different types of setae hastisetae, spicisetae and/or fiscisetae (Figures 13 and 15) (6) head of hastisetae not more than three times longer than wide (Figure 15) (7) numerous hastisetae on all nota and terga, with prominent tufts of erect hastisetae inserted on the posterolateral part of the tergal plates of abdominal segments 6 to 8 (in Anthrenus genus the tufts of hastisetae are inserted on the membrane behind the sclerotized part of terga 5, 6 and 7) (8) urogomphi absent.	According to observation and records, as well as refer to the larvae picture(B, C) in [160]. The shape oof larvae should be spindle.	China
[252]	82	Technical	(1) body elongated, cylindrical, somewhat flattened, roughly six times as long as wide, nearly parallel-sided but gradually tapering toward rear (2) head well developed, sclerotized, and hypognathous (3) three pairs of jointed legs present (4) pretarsal setae on the ventral side of claws unequal (5) very hairy, being covered with different types of setae hastisetae, spicisetae and/or fiscisetae (Figures 13 and 15) (6) head of hastisetae not more than three times longer than wide (Figure 15) (7) numerous hastisetae on all nota and terga, with prominent tufts of erect hastisetae inserted on the posterolateral part of the tergal plates of abdominal segments 6 to 8 (in *Anthrenus* genus the tufts of hastisetae are inserted on the membrane behind the sclerotized part of terga 5, 6 and 7 Fig.) (8) urogomphi absent.		New Zealand
[253]	83	Editorial	**4.5 Identification of *Trogoderma granarium* late instar larvae**		New Zealand

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
[254]	83	Substantive	**4.5 Identification of *Trogoderma granarium* larvae** Add more species as reference such as T.angustum, T.teukton, T.inclusum, T. versicolor, T. anthrenoides	Because the listed three species are too less to help indentify the specie Trogoderma granarium. See reference paper:Zhang sheng Fang et al. The Identification of 8 important larva species in Genus Trogoderma, Plant Quarantine, 2007,Vol 21 (5):284-287	China
[255]	84	Editorial	**4.5.1 Identification key ~~of~~to *Trogoderma granarium* larvae**	To be consistent with title for Trogoderma granarium adults.	United States of America
[256]	84	Technical	**4.5.1 Identification key of *Trogoderma ~~granarium~~* larvae**	This is because the key includes others species of Trogoderma	Mexico
[257]	86	Editorial	(1) Epipharynx with 4 distal papillae, usually in a single sensory cup……Fig ……**2**		New Zealand
[258]	87	Technical	Epipharynx with 6 distal papillae…? in a sensory cup……..**3**	Clarify if these papillae are in a sensory cup or not, to align with first point	Australia
[259]	88	Substantive	(2) Terga uniformly yellowish-brown, without greyish pigmentation at base of large spicisetae; acrotergites weakly sclerotized; antecostal suture on 8th abdominal segment almost always absent (if present, faint and usually broken); setae ~~almost completely encircling~~occupy 50 to 75 percent of the basal antennal segment, second segment usually with a single seta, apical segment with sensory pores in basal quarter; hastisetae morphology as in Figure 15(A), (B) ***Trogoderma granarium* Everts**	We have never seen a specimen of khapra beetle that has had "setae almost completely encircling basal antennal segment". Perhaps many do display this condition, but we have only observed specimens whose setae occupy 50 to 75 percent of the basal antennal segment.	United States of America
[260]	88	Technical	(2) Terga uniformly yellowish-brown, without greyish pigmentation at base of large spicisetae; acrotergites weakly sclerotized; antecostal suture on 8th abdominal segment almost always absent (if present, faint and usually broken); setae almost completely encircling basal antennal segment, second segment usually with a single seta or no seta, apical segment with sensory pores in basal quarter; hastisetae morphology as in Figure 15(A), (B) ***Trogoderma granarium* Everts**	Reference: Zhang, Sh. F, 2007	Korea, Republic of ,Lao People's Democratic Republic,India
[261]	88	Technical	~~(2) Terga uniformly yellowish-brown, without greyish pigmentation at base of large spicisetae; acrotergites weakly sclerotized; antecostal suture on 8th abdominal segment almost always absent (if present, faint and usually broken); setae almost completely encircling basal antennal segment, second segment usually with a single seta, apical segment with sensory pores in basal quarter; hastisetae morphology as in Figure 15(A), (B) *Trogoderma granarium* Everts~~ (2) Terga uniformly yellowish-brown, without greyish pigmentation at base of large spicisetae; acrotergites weakly sclerotized; antecostal suture on	According to the observation during work and the public paper:Zhang sheng Fang et al. The Identification of 8 important larva species in Genus Trogoderma, Plant Quarantine, 2007,Vol 21 (5):284-287	China

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			8th abdominal segment almost always absent (if present, faint and usually broken); setae almost completely encircling basal antennal segment, second segment usually with a single seta or no seta, apical segment with sensory pores in basal quarter; hastisetae morphology as in Figure 15(A), (B) Trogoderma granarium Everts		
[262]	91	Substantive	Specimen without above combination of characters **other *Trogoderma* spp.** Add more species as reference such as T.angustum, T.teukton, T.inclusum, T. versicolor, T. anthrenoides.	Because the listed three species are too less to help indentify the specie Trogoderma granarium. See reference paper:Zhang sheng Fang et al. The Identification of 8 important larva species in Genus Trogoderma, Plant Quarantine, 2007,Vol 21 (5):284-287	China
[263]	92	Editorial	Larval identification should be considered unreliable if it is based only on one specimen, or exuviae or worn specimens. This is because in many species the intraspecific variation is such that in individual specimens features considered specific to the species ~~cannot~~ may not be seen, while features specific to other species ~~can~~ may be. In addition, large numbers of non-pest *Trogoderma* species occur in stored commodities and many of their characteristics are not well studied.	"Can" is usually avoided in ISPMs; we therefore suggest "may"	EPPO,Russian Federation ,Ukraine ,Morocco ,Uzbekistan
[264]	92	Editorial	Larval identification should be considered unreliable if it is based only on one specimen, or exuviae or worn specimens. This is because in many species the intraspecific variation is such that in individual specimens features considered specific to the species ~~cannot~~ may not be seen, while features specific to other species ~~can~~ may be. In addition, large numbers of non-pest *Trogoderma* species occur in stored commodities and many of their characteristics are not well studied.	"Can" is usually avoided in ISPMs; we therefore suggest "may"	European Union
[265]	95	Editorial	(1) antennal segments subequal (2) setae of basal antennal segment almost completely encircling the segment, reaching or surpassing apex of second segment, at least three-fourths as long as the second antennal segment (3) second antennal segment of last instar usually with one seta (4) last antennal segment with at least one sensory pore in basal quarter (5) epipharynx with four papillae in distal sensory cup, usually in a single unit (6) fiscisetae absent (7) mesally directed tergal setae absent (8) at least six small spicisetae on first abdominal tergum, posterior to antecostal suture, anterior to large spicisetae (9) anterior-median small spicisetae anterior to antecostal suture not long enough to reach over the suture	clarity	Australia

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			(10) large median spicisetae on first abdominal segment smooth or covered with inconspicuous scales with tips smooth for at least four times the diameter of seta (11) antecostal suture of 8th abdominal tergum almost always absent, but if present, faint and interrupted (12) antecostal suture on 7th abdominal tergum faint or interrupted (13) no greyish pigmentation on sides of thoracic and other segments, not even at the base of large lateral spicisetae.		
[266]	95	Substantive	(1) antennal segments subequal (2) setae of basal antennal segment ~~almost completely encircling~~occupying 50 to 75 percent of the circumference of the segment, reaching or surpassing apex of second segment, at least three-fourths as long as the second antennal segment (3) second antennal segment of last instar usually with one seta (4) last antennal segment with at least one sensory pore in basal quarter (5) epipharynx with four papillae in distal sensory cup, usually in a single unit (6) fiscisetae absent (7) mesally directed tergal setae absent (8) at least six small spicisetae on first abdominal tergum posterior to antecostal suture anterior to large spicisetae (9) anterior-median small spicisetae anterior to antecostal suture not long enough to reach over the suture (10) large median spicisetae on first abdominal segment smooth or covered with inconspicuous scales with tips smooth for at least four times the diameter of seta (11) antecostal suture of 8th abdominal tergum almost always absent, but if present, faint and interrupted (12) antecostal suture on 7th abdominal tergum faint or interrupted (13) no greyish pigmentation on sides of thoracic and other segments, not even at the base of large lateral spicisetae.	Same explanation as in paragraph 88. Even if author has seen specimens where setae encircle the basal antennal segment, he/she should modify statements in paragraphs 88 and 95 to take into account variability in this character.	United States of America
[267]	95	Technical	(1) antennal segments subequal (2) setae of basal antennal segment almost completely encircling the segment, reaching or surpassing apex of second segment, at least three-fourths as long as the second antennal segment (3) second antennal segment of last instar usually with one seta or no seta (4) last antennal segment with at least one sensory pore in basal quarter	Reference: Zhang, Sh. F, 2007	Korea, Republic of ,Lao People's Democratic Republic,India

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			(5) epipharynx with four papillae in distal sensory cup, usually in a single unit (6) fiscisetae absent (7) mesally directed tergal setae absent (8) at least six small spicisetae on first abdominal tergum posterior to antecostal suture anterior to large spicisetae (9) anterior-median small spicisetae anterior to antecostal suture not long enough to reach over the suture (10) large median spicisetae on first abdominal segment smooth or covered with inconspicuous scales with tips smooth for at least four times the diameter of seta (11) antecostal suture of 8th abdominal tergum almost always absent, but if present, faint and interrupted (12) antecostal suture on 7th abdominal tergum faint or interrupted (13) no greyish pigmentation on sides of thoracic and other segments, not even at the base of large lateral spicisetae.		
[268]	95	Technical	~~(1) antennal segments subequal(2) setae of basal antennal segment almost completely encircling the segment, reaching or surpassing apex of second segment, at least three fourths as long as the second antennal segment(3) second antennal segment of last instar usually with one seta(4) last antennal segment with at least one sensory pore in basal quarter(5) epipharynx with four papillae in distal sensory cup, usually in a single unit(6) fiscisetae absent(7) mesally directed tergal setae absent(8) at least six small spicisetae on first abdominal tergum posterior to antecostal suture anterior to large spicisetae(9) anterior-median small spicisetae anterior to antecostal suture not long enough to reach over the suture(10) large median spicisetae on first abdominal segment smooth or covered with inconspicuous scales with tips smooth for at least four times the diameter of seta(11) antecostal suture of 8th abdominal tergum almost always absent, but if present, faint and interrupted(12) antecostal suture on 7th abdominal tergum faint or interrupted(13) no greyish pigmentation on sides of thoracic and other segments, not even at the base of large lateral~~ (1) antennal segments subequal (2) setae of basal antennal segment almost completely encircling the segment, reaching or surpassing apex of second segment, at least three-fourths as long as the second antennal segment (3) second antennal segment of last instar usually with one seta, sometimes no seta. (4) last antennal segment with at least one sensory pore in basal quarter (5) epipharynx with four papillae in distal sensory cup, usually in a single unit (6) fiscisetae absent (7) mesally directed tergal	According to the observation during work and the public paper:Zhang sheng Fang et al. The Identification of 8 important larva species in Genus Trogoderma, Plant Quarantine, 2007,Vol 21 (5):284-287	China

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			setae absent (8) at least six small spicisetae on first abdominal tergum posterior to antecostal suture anterior to large spicisetae (9) anterior-median small spicisetae anterior to antecostal suture not long enough to reach over the suture (10) large median spicisetae on first abdominal segment smooth or covered with inconspicuous scales with tips smooth for at least four times the diameter of seta (11) antecostal suture of 8th abdominal tergum almost always absent, but if present, faint and interrupted (12) antecostal suture on 7th abdominal tergum faint or interrupted (13) no greyish pigmentation on sides of thoracic and other segments, not even at the base of large lateral spicisetae.		
[269]	97	Substantive	The first-instar larva (Figure 1(B)) is 1.6–1.8 mm long and 0.25–0.3 mm wide. Body is uniformly yellowish-white, head and hairs are reddish-brown. The mature larva (Figure 1(C)) is 4.5–6 mm long and 1.5 mm wide and body is reddish-brown. The larval body is covered with two kinds of hairs: spicisetae (Figure 13(B)), in which the shaft is covered with tiny, stiff, upwardly directed, pointed scales; and hastisetae (Figure 13(A)), in which the shaft is multi-segmented with spear-headed apex. Spicisetae are scattered over the dorsal surface of the head and body segments. Two groups of long spicisetae on the 9th abdominal segment form the tail. Hastisetae are found on all notal and abdominal segments, but on the last three or four segments they form distinctive, paired, erect tufts (Beal, 1960, 1991; EPPO/CABI, 1997). 4.6 Adults 4.6.1 Simple key for differentiation of dermestid adults (1) Median ocellus absent Dermestes spp. (Figure 10) Median ocellus present……2 (2) Body covered with scale-like setae; antennal cavity filled by antennae, fully visible from anterior view (Figure 9(A)) Anthrenus spp. (Figure 12) Body covered with simple setae, some of them whitish, flattened (ensiform) but never scale-like....3 (3) Antennal cavity completely closed behind, antennal club 3-segmented and well defined Anthrenocerus spp. Antennal cavity open behind or partially delimited by a posterior carina, antennae cavity much wider than antennae, not visible in anterior view (Figure 9(B))……..4 (4) Antennal cavity open behind, posterior margin of hind coxa angulate, first segment of posterior tarsus shorter than second segment Attagenus spp. (Figure 11)	New paragraphs after 97. - Moved from paragraph 66 plus a new subheading to introduce the simple key for adults - Former paragraphs 67-73 moved here so that all information on adults is in the same place - Former para 74 added, plus a new paragraph to ensure that diagnosticians do not stop the diagnosis here. Wording adapted from paragraph 100.	EPPO,Russian Federation ,Ukraine ,Morocco ,Uzbekistan

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			Antennal cavity carinate posteriorly, posterior margin of hind coxa straight, arcuate or sinuate, first segment of posterior tarsus longer than second segment Trogoderma spp. (Figures 1(A), 3, 9(B)). Adult specimens having the above characteristics are very likely to be Trogoderma species and therefore it is warranted to check the detailed list of features listed in section 4.6.2		
[270]	97	Substantive	The first-instar larva (Figure 1(B)) is 1.6–1.8 mm long and 0.25–0.3 mm wide. Body is uniformly yellowish-white, head and hairs are reddish-brown. The mature larva (Figure 1(C)) is 4.5–6 mm long and 1.5 mm wide and body is reddish-brown. The larval body is covered with two kinds of hairs: spicisetae (Figure 13(B)), in which the shaft is covered with tiny, stiff, upwardly directed, pointed scales; and hastisetae (Figure 13(A)), in which the shaft is multi-segmented with spear-headed apex. Spicisetae are scattered over the dorsal surface of the head and body segments. Two groups of long spicisetae on the 9th abdominal segment form the tail. Hastisetae are found on all notal and abdominal segments, but on the last three or four segments they form distinctive, paired, erect tufts (Beal, 1960, 1991; EPPO/CABI, 1997). 4.6 Adults 4.6.1 Simple key for differentiation of dermestid adults (1) Median ocellus absent Dermestes spp. (Figure 10) Median ocellus present……2 (2) Body covered with scale-like setae; antennal cavity filled by antennae, fully visible from anterior view (Figure 9(A)) Anthrenus spp. (Figure 12) Body covered with simple setae, some of them whitish, flattened (ensiform) but never scale-like....3 (3) Antennal cavity completely closed behind, antennal club 3-segmented and well defined Anthrenocerus spp. Antennal cavity open behind or partially delimited by a posterior carina, antennae cavity much wider than antennae, not visible in anterior view (Figure 9(B))……..4 (4) Antennal cavity open behind, posterior margin of hind coxa angulate, first segment of posterior tarsus shorter than second segment Attagenus spp. (Figure 11) Antennal cavity carinate posteriorly, posterior margin of hind coxa straight, arcuate or sinuate, first segment of posterior tarsus longer than second segment Trogoderma spp. (Figures 1(A), 3, 9(B)). Adult specimens having the above characteristics are very likely to be	New paragraphs after [97]. - Moved from paragraph [66] plus a new subheading to introduce the simple key for adults - Former paragraphs [67]-[73] moved here so that all information on adults is in the same place - Former para [74] added, plus a new paragraph to ensure that diagnosticians do not stop the diagnosis here. Wording adapted from paragraph [100].	European Union

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			Trogoderma species and therefore it is warranted to check the detailed list of features listed in section 4.6.2		
[271]	98	Editorial	**4.~~6~~7 Identification of *Trogoderma* adults**	Renumbered to take into account new section on differentiation of dermestid adults	EPPO,Russian Federation ,European Union ,Ukraine ,Morocco ,Uzbekistan
[272]	99	Substantive	~~**4.6.1 Preliminary identification of *Trogoderma* adults**~~	Suggest deletion of this section. Most of the information is repeated in paragraphs 102-104. Merge any relevant requirements with paragraphs 102-104.	EPPO,Russian Federation ,Ukraine ,Morocco ,Uzbekistan
[273]	99	Substantive	~~**4.6.1 Preliminary identification of *Trogoderma* adults**~~	Suggest deletion of this section. Most of the information is repeated in paragraphs [102]-[104]. Merge any relevant requirements with paragraphs [102]-[104].	European Union
[274]	100	Substantive	~~If all of the following features can be observed it is very likely that the specimen is a *Trogoderma* species; therefore it is warranted to check the detailed list of features listed in section 4.6.2:~~	Delete. Propose to cover the concept in new paragraph after 97 (formerly paragraph 74)	EPPO,Russian Federation ,Ukraine ,Morocco ,Uzbekistan
[275]	100	Substantive	~~If all of the following features can be observed it is very likely that the specimen is a *Trogoderma* species; therefore it is warranted to check the detailed list of features listed in section 4.6.2:~~	Delete. Propose to cover the concept in new paragraph after [97] (formerly paragraph [74])	European Union
[276]	101	Editorial	median ocellus present antennal cavity well defined by a posterior carina and open laterally antennal outline smooth (Figure 5(A)), antennal club at least three-segmented body hairy, elytra usually with three transverse bands of pale (ensiform) setae (setae of dead adults often absent as they are rubbed off).		Australia
[277]	101	Editorial	median ocellus present antennal cavity well defined by a posterior carina and open laterally antennal outline smooth (Figure 5(A)), antennal club at least three-	Ensiform is a term that relates to shape. If this character is important, then use "bands of pale, ensiform setae" or "bands of pale, sword-shaped setae", without the parentheses.	United States of America

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			segmented body hairy, elytra usually with three transverse bands of pale ~~(ensiform)~~, sword-shaped setae (setae of dead adults often rubbed off).		
[278]	101	Substantive	~~median ocellus present~~ ~~antennal cavity well defined by a posterior carina and open laterally~~ ~~antennal outline smooth (Figure 5(A)), antennal club at least three-segmented~~ ~~body hairy, elytra usually with three transverse bands of pale (ensiform) setae (setae of dead adults often rubbed off).~~	Delete paragraph. Most is already covered in paragraphs 102-104. Merge any relevant requirements with paragraph 104.	EPPO,Russian Federation ,Ukraine ,Morocco ,Uzbekistan
[279]	101	Substantive	~~median ocellus present~~ ~~antennal cavity well defined by a posterior carina and open laterally~~ ~~antennal outline smooth (Figure 5(A)), antennal club at least three-segmented~~ ~~body hairy, elytra usually with three transverse bands of pale (ensiform) setae (setae of dead adults often rubbed off).~~	Delete paragraph. Most is already covered in paragraphs [102]-[104]. Merge any relevant requirements with paragraph [104].	European Union
[280]	101	Technical	median ocellus present antennal cavity well defined by a posterior carina and open laterally antennal outline smooth (Figure 5(A)), antennal club at least three-segmented: female - 4 to 5 segmented; male - 5-8 segmented body hairy, elytra usually with three transverse bands of pale (ensiform) setae (setae of dead adults often rubbed off).	Reference: Zhang, Sh. F, 2007	Korea, Republic of ,Lao People's Democratic Republic,India
[281]	101	Technical	~~median ocellus presentantennal cavity well defined by a posterior carina and open laterallyantennal outline smooth (Figure 5(A)), antennal club at least three-segmentedbody hairy, elytra usually with three transverse bands of pale (ensiform) setae (setae of dead adults often rubbed of~~ • median ocellus present • antennal cavity well defined by a posterior carina and open laterally • antennal outline smooth (Figure 5(A)), antennal club at least three-segmented. Female usually 4-5 segmented, male usually 5-8 segmented. • body hairy, elytra usually with three transverse bands of pale (ensiform)	According to the observation during work and the public paper:Zhang sheng Fang et al. The Identification of 8 important larva species in Genus Trogoderma, Plant Quarantine, 2007,Vol 21 (5):284-287	China

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			setae (setae of dead adults often rubbed off).		
[282]	102	Editorial	**4.~~6.2~~7.1 Discriminating features of *Trogoderma* adults**	Renumbered to take into account new section on differentiation of dermestid adults	EPPO,Russian Federation ,European Union ,Ukraine ,Morocco ,Uzbekistan
[283]	103	Editorial	The features below are adapted from Banks (1994), Beal (1954, 1960), Haines (1991), Háva (2004), Hinton (1945), Kingsolver (1991), Lawrence and Britton (1991, 1994), Lawrence *et al.* (1999b), Okumura and Blanc (1955) and Peacock (1993):	The manner by which multiple citations are ordered needs to be consistent within the entire document.	United States of America
[284]	104	Editorial	(1) body ovate, densely setose, setae simple, usually 2–3 different types, recumbent, yellowish-white slightly flattened ~~(ensiform)~~ , sword-shaped setae (2) presence of median ocellus (3) pronotum without lateral carina (4) antennal cavity of anteroventral surface not, or only slightly, visible in anterior view (Figure 9(B)) (5) antennal cavity carinate posteriorly at least to half of length and open laterally (6) prosternum forming a “collar” anteriorly (7) mesosternum deeply divided by sulcus (8) posterior margin of hind coxal plate curved or sinuate, never angulate (9) first segment of hind tarsus longer than second segment (10) antennae short, 9–11-segmented, with a 3–8-segmented club, antennal outline usually smooth or rarely flabellate, terminal segment never disproportionately enlarged (11) tarsi of all legs 5-segmented.	See paragraph 101. Ensiform is a term relating to shape. If setal shape is important, then use "flattened, sword-shaped" or "flattened, ensiform" setae without parentheses. Comma inserted in (4). Rule of punctuation.	United States of America
[285]	105	Editorial	**4.8~~7~~ Identification of *Trogoderma granarium* adults**	Renumbered to take into account new section on differentiation of dermestid adults	EPPO,Russian Federation ,European Union ,Ukraine ,Morocco ,Uzbekistan
[286]	106	Editorial	The following short key ~~elaborated by Andras Szito~~ should be used to distinguish adult *T. granarium* from some other *Trogoderma* species frequently occurring in stored commodities. If necessary, other species can be identified with the keys of Beal (1954, 1956), Banks (1994), Kingsolver	Text not needed as the document is supplied by IPPC	Australia

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			(1991) and Mordkovich and Sokolov (1999). These keys include species occurring in stored products and therefore may be used for identification of *Trogoderma* adults. It should be noted, that identification of adult sex of various *Trogoderma* species is practically possible only after dissecting of their genitalia (for morphology of male and female genitalia, see Figures 7 and 8). Checking of external distinguishing features as antennal club morphology should be performed on specimens surely identified to sex.		
[287]	106	Editorial	The following short key elaborated by Andras Szito should be used to distinguish adult *T. granarium* from some other *Trogoderma* species frequently occurring in stored commodities. If necessary, other species can be identified with the keys of Beal (1954, 1956), Banks (1994), Kingsolver (1991) and Mordkovich and Sokolov (1999). These keys include species occurring in stored products and therefore may be used for identification of *Trogoderma* adults. It should be note~~d,~~ that identification of adult sex of various *Trogoderma* species is practically possible only after dissecting ~~of~~ their genitalia (for morphology of male and female genitalia, see Figures 7 and 8). Checking of external distinguishing features as antennal club morphology should be performed on specimens surely identified to sex.	Punctuation and grammatical points.	United States of America
[288]	107	Editorial	**4.~~7~~8.1 Identification key to *Trogoderma granarium* adults**	Renumbered to take into account new section on differentiation of dermestid adults	EPPO,Russian Federation ,European Union ,Ukraine ,Morocco ,Uzbekistan
[289]	107	Technical	**4.7.1 Identification key to *Trogoderma granarium* adults**	Question to author: Why are Trogoderma longisetosum and Trogoderma cavum, which are cited at the begining of the diagnostic protocol, not considered in this key?	EPPO,Russian Federation ,Ukraine ,Morocco ,Uzbekistan
[290]	107	Technical	**4.7.1 Identification key to *Trogoderma ~~granarium~~* adults**	To distinguish T. granarium of the others species of Trogoderma. The key is useful not only to distinguish T granarium adults also include some other Trogoderma species	Mexico
[291]	107	Technical	**4.7.1 Identification key to *Trogoderma granarium* adults**	Question to author: Why are Trogoderma longisetosum and Trogoderma cavum, which are cited at the begining of the diagnostic protocol, not considered in this key?	European Union
[292]	118	Editorial	(6) Basal loop never connected to the antemedian band ***Trogoderma variabile*** **Ballion** (Figure 4)	Figure 4 illustrates this character well.	United States of America

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
[293]	119	Editorial	Basal loop of elytral maculation connected to the antemedian band by a longitudinal band or bands (*T. inclusum* with less obvious emargination of eyes may key out here) ***Trogoderma ornatum* (Say),** ***T. simplex* Jayne, *T. sternale* Jayne, *T. versicolor* (Creutzer)**	Trogoderma ornatum and T. simplex should be brought into proximity of one another. Typist used a hard stop after "(Say)", preventing T. simplex from following directly after it.	United States of America
[294]	120	Editorial	(7) Elytral integument with three well-defined (basal, submedian and apical) fasciae, setae on fasciae largely white, ensiform with ~~very little~~ sparse, yellowish, recumbent setae ***Trogoderma angustum* (Solier)**	'Sparse' is better word to denote number of yellow setae, not size of yellow setae. Beal (1956) speaks to this in in key: T. angustum has "hairs of elytral fasciae almost entirely white: less than three percent light golden brown."	United States of America
[295]	121	Editorial	Elytral integument with well-defined basal band and median spot *Trogoderma variabile* (reduced pattern) (Figure 4) ~~***Trogoderma variabile* reduced pattern** (Figure 4).~~	Is this the meaning?	EPPO,Russian Federation ,European Union ,Ukraine ,Morocco ,Uzbekistan
[296]	122	Editorial	In general, ~~E~~elytral fasciae of *Trogoderma* species usually form a more or less complete basal loop, ante-median and median bands and apical spots. Some specimens have a reduced elytral pattern where the basal loop is indicated by curved anterior band, antemedian and/or median bands by small spots, and apical spots are usually missing.	To set apart a thought different from the last paragraph of the key---- and to improve comprehension.	United States of America
[297]	125	Editorial	Matveeva (2001) provides additional features for separat~~ion~~ing ~~of~~ adults of *Trogoderma granarium* from *T. variabile* (Figures 3, 4) and *T. glabrum*. Size and morphology of hind wings can be useful for identifying damaged specimens and, although considering ~~of~~ these two characteristics is not mandatory, it helps to increase the certainty of identification based on other features (Figure 6). During dissection, hind wings must be removed and mounted in glycerol or Hoyer's medium.	Grammatical points. Use of commas to separate a clause and a prepositional phrase.	United States of America
[298]	126	Editorial	Hind wings of the Khapra beetle are smaller (mean length is 1.9 mm as compared with 2.5 mm for *T. variabile* and *T. glabrum*); they are paler in colour with less visible venation; number of setae S1 on costal vein (mean = 10) is half that on *T. variabile* and *T. glabrum* (mean = 20–23); number of small setae S2 between costal vein and pterostigma (mean = 2, sometimes absent) is less than that for *T. variabile* and *T. glabrum* (mean = 8) (Figure 6).		New Zealand
[299]	126	Editorial	Hind wings of the ~~K~~khapra beetle are smaller (mean length is 1.9 mm as compared with 2.5 mm for *T. variabile* and *T. glabrum*); they are paler in colour with less visible venation; number of setae S1 on costal vein (mean = 10) is half that on *T. variabile* and *T. glabrum* (mean = 20–23); number of small setae S2 between costal vein and pterostigma (mean = 2, sometimes	Khapra beetle: See general comments. "less than": grammatical rule.	United States of America

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			absent) is less than that for *T. variabile* and *T. glabrum* (mean = 8) (Figure 6).		
[300]	127	Editorial	**4.~~7~~8.2 Discriminating features of *Trogoderma granarium* adults**	Renumbered to take into account new section on differentiation of dermestid adults	EPPO,Russian Federation ,European Union ,Ukraine ,Morocco ,Uzbekistan
[301]	130	Editorial	(1) elytral cuticle unicoloured, usually light brown or reddish-brown, or vaguely mottled without a clearly defined pattern (2) elytral setae predominantly brown (~~Y~~yellowish or white hairs forming no clearly defined banded pattern may also be present; these hairs are gradually rubbed off as the beetle moves around and the adult develops a shiny appearance.) (3) antennae with 9–11 segments; male antennal club with 4–5 segments; female antennal club with 3–4 segments (Figure 5) (4) inner eye margin straight or sinuate (5) male abdominal tergum 8 more or less evenly sclerotized, with setae along its margin sometimes tending to be grouped medially; tergum 9 with proximal margin of broader section almost U-shaped; tergum 10 with many long setae (6) serrate sclerites of bursa copulatrix of female small, not longer than corrugated part of spermatheca, with 10–15 teeth (Figure 8) (7) male genitalia with bridge straight, and evenly wide, broader at connections to the parameres (Figure 7(A)).	The initial "Y" in brackets should not be capital.	EPPO,Russian Federation ,Ukraine ,Morocco ,Uzbekistan
[302]	130	Editorial	(1) elytral cuticle unicoloured, usually light brown or reddis~~h~~-brown, or vaguely mottled without a clearly defined pattern (2) elytral setae predominantly brown; (~~Y~~yellowish or white ~~hairs~~setae forming no clearly defined banded pattern may also be present; these ~~hairs~~setae are gradually rubbed off as the beetle moves around, and the adult thus develops a shiny appearance~~.~~) (3) antennae with 9–11 segments; male antennal club with 4–5 segments; female antennal club with 3–4 segments (Figure 5) (4) inner eye margin straight or sinuate (5) male abdominal tergum 8 more or less evenly sclerotized, with setae along its margin sometimes tending to be grouped medially; tergum 9 with proximal margin of broader section almost U-shaped; tergum 10 with many long setae (6) serrate sclerites of bursa copulatrix of female small, not longer than	(1) No hyphen necessary if not preceding a noun. (2) Improve comprehension by removing parentheses and keeping terminology consistent with 'setae', not 'hairs'. Comma needed to separate two ideas. (7) Delete period to produce consistency with (1) through (6).	United States of America

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			corrugated part of spermatheca, with 10–15 teeth (Figure 8) (7) male genitalia with bridge straight, and evenly wide, broader at connections to the parameres (Figure 7(A)~~)~~.		
[303]	130	Editorial	(1) elytral cuticle unicoloured, usually light brown or reddish-brown, or vaguely mottled without a clearly defined pattern (2) elytral setae predominantly brown (~~Y~~yellowish or white hairs forming no clearly defined banded pattern may also be present; these hairs are gradually rubbed off as the beetle moves around and the adult develops a shiny appearance.) (3) antennae with 9–11 segments; male antennal club with 4–5 segments; female antennal club with 3–4 segments (Figure 5) (4) inner eye margin straight or sinuate (5) male abdominal tergum 8 more or less evenly sclerotized, with setae along its margin sometimes tending to be grouped medially; tergum 9 with proximal margin of broader section almost U-shaped; tergum 10 with many long setae (6) serrate sclerites of bursa copulatrix of female small, not longer than corrugated part of spermatheca, with 10–15 teeth (Figure 8) (7) male genitalia with bridge straight, and evenly wide, broader at connections to the parameres (Figure 7(A)).	The initial "Y" in brackets should not be capital.	European Union
[304]	131	Editorial	**4.~~7~~8.3 Description of *Trogoderma granarium* adults**	Renumbered to take into account new section on differentiation of dermestid adults	EPPO,Russian Federation ,European Union ,Ukraine ,Morocco ,Uzbekistan
[305]	132	Editorial	~~A~~ The adult stage of *T. granarium* ~~adult~~ is illustrated in Figure 1(A).	To improve comprehension.	United States of America
[306]	134	Editorial	Body: Length 1.4–2.3 mm (mean 1.99 mm), width 0.75–1.1 (mean 0.95 mm~~,~~) mm, ratio of length to width about 2.1:1. Head and pronotum dark reddish-brown; elytra reddish-brown, usually with indistinct lighter reddish-brown fasciae. Venter of thorax and abdomen reddish-brown; legs yellowish-brown.	In the second bracket, the comma is to be deleted.	EPPO,Russian Federation ,European Union ,Ukraine ,Morocco ,Uzbekistan
[307]	134	Editorial	Body: Length 1.4–2.3 mm (mean 1.99 mm), width 0.75–1.1 mm(mean 0.95 mm,) ~~mm,~~ ratio of length to width about 2.1:1. Head and pronotum dark reddish-brown; elytra reddish-brown, usually with indistinct lighter reddish-brown fasciae. Venter of thorax and abdomen reddish-brown; legs	Consistency	Malaysia

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			yellowish-brown.		
[308]	134	Editorial	Body: Length 1.4–2.3 mm (mean 1.99) ~~mm)~~, width 0.75–1.1 mm (mean 0.95) ~~mm,) mm~~, ratio of length to width about 2.1:1. Head and pronotum dark reddish-brown; elytra reddish-brown, usually with indistinct lighter reddish-brown fasciae. Venter of thorax and abdomen reddish-brown; legs yellowish-brown.		Philippines
[309]	134	Editorial	Body: Length 1.4–2.3 mm (mean 1.99) ~~mm)~~, width 0.75–1.1 (mean 0.95 mm,) mm, ratio of length to width about 2.1:1. Head and pronotum dark reddish-brown; elytra reddish-brown, usually with indistinct lighter reddish-brown fasciae. Venter of thorax and abdomen reddish-brown; legs yellowish-brown.		Thailand ,Korea, Republic of ,Lao People's Democratic Republic,Japan ,India
[310]	134	Editorial	Body: Length 1.4–2.3 mm (mean 1.99 mm), width 0.75–1.1 mm (mean 0.95 ~~mm,~~mm) ~~mm~~, ratio of length to width about 2.1:1. Head and pronotum dark reddish-brown; elytra reddish-brown, usually with indistinct lighter reddish-brown fasciae. Venter of thorax and abdomen reddish-brown; legs yellowish-brown.	'mm' in wrong position placed after 'width 0.75-1.1'. An extra comma deleted after 'mean 0.95 mm'.	United States of America
[311]	140	Editorial	Hind wings with vague venation; mean number of larger setae S1 on costal vein is 10, mean number of small setae S2 between costal vein and pterostigma is 2, but sometimes these are missing.	Insertion of a verb.	United States of America
[312]	152	Editorial	In cases where other contracting parties may be adversely affected by the results of the diagnosis, the records and evidence (in particular, preserved larvae and adults, slide-mounted specimens, photographs) should be kept for at least one year.	To be consistent with ISPM 27:2006.	Costa Rica ,Uruguay ,Mexico ,Nicaragua ,El Salvador
[313]	152	Editorial	In cases where other contracting parties may be adversely affected by the results of the diagnosis, the records and evidence (in particular, preserved larvae and adults, slide-mounted specimens, photographs) should be kept for at least one year.	To be consistent with ISPM 27.	COSAVE,Paraguay ,Argentina ,Chile,Brazil
[314]	152	Editorial	In cases where other contracting parties may be adversely affected by the results of the diagnosis, the records and evidence (in particular, preserved larvae and adults, slide-mounted specimens, photographs) should be kept for at least one year.	To be consistent with ISPM 27	OIRSA
[315]	155	Technical	Department of Agriculture and Food Western Australia, Biosecurity & Research Division, Plant Biosecurity Branch, Entomology Unit, 3 Baron-Hay Court, South Perth, WA 6151, Australia (tel: +61 8 9368 3248, +61 8 9368 3965; fax: +61 8 9368 3223, +61 8 9474 2840; e-mail: aszito@agric.wa.gov.au).	Corrections: 1st - of the telephone number 2nd - of the e-mail addresses.	EPPO,Russian Federation ,Ukraine ,Morocco ,Uzbekistan

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			Main Inspectorate of Plant Health and Seed Service, Central Laboratory, Żwirki i Wigury 73, 87-100 Toruń, Poland (tel: +48 56 639 1110~~0~~1, +48 56 639 1115; fax: +48 56 639 1115; e-mail: wkarnkowski@piorin.gov.pl). Laboratorio de Plagas y Enfermedades de las Plantas. Servicio Nacional de Sanidad y Calidad Agroalimentaria (SENASA), Av. Ing. Huergo 1001, C1107AOK Buenos Aires, Argentina (tel: +54 11 4362 1177, extns 117, 118, 129 and 132; fax: +54 11 4362 1177, extn 171; e-mail: ~~alb~~abriano@senasa.gov.ar,).		
[316]	155	Technical	Department of Agriculture and Food Western Australia, Biosecurity & Research Division, Plant Biosecurity Branch, Entomology Unit, 3 Baron-Hay Court, South Perth, WA 6151, Australia (tel: +61 8 9368 3248, +61 8 9368 3965; fax: +61 8 9368 3223, +61 8 9474 2840; e-mail: aszito@agric.wa.gov.au). Main Inspectorate of Plant Health and Seed Service, Central Laboratory, Żwirki i Wigury 73, 87-100 Toruń, Poland (tel: +48 56 639 1110, +48 56 639 1115; fax: +48 56 639 1115; e-mail: wkarnkowski@piorin.gov.pl). Laboratorio de Plagas y Enfermedades de las Plantas. Servicio Nacional de Sanidad y Calidad Agroalimentaria (SENASA), Av. Ing. Huergo 1001, C1107AOK Buenos Aires, Argentina (tel: +54 11 4362 1177, extns 117, 118, 129 and 132; fax: +54 11 4362 1177, extn 171; e-mail: albabriano@senasa.gov.ar).	Experts of the institutions should be identified to facilitate the contact. We suggest including their name in addition to the institution in this section.	Costa Rica ,Uruguay ,Nicaragua ,El Salvador
[317]	155	Technical	Department of Agriculture and Food Western Australia, Biosecurity & Research Division, Plant Biosecurity Branch, Entomology Unit, 3 Baron-Hay Court, South Perth, WA 6151, Australia (tel: +61 8 9368 3248, +61 8 9368 3965; fax: +61 8 9368 3223, +61 8 9474 2840; e-mail: aszito@agric.wa.gov.au). Main Inspectorate of Plant Health and Seed Service, Central Laboratory, Żwirki i Wigury 73, 87-100 Toruń, Poland (tel: +48 56 639 1110, +48 56 639 1115; fax: +48 56 639 1115; e-mail: wkarnkowski@piorin.gov.pl). Laboratorio de Plagas y Enfermedades de las Plantas. Servicio Nacional de Sanidad y Calidad Agroalimentaria (SENASA), Av. Ing. Huergo 1001, C1107AOK Buenos Aires, Argentina (tel: +54 11 4362 1177, extns 117, 118, 129 and 132; fax: +54 11 4362 1177, extn 171; e-mail: albabriano@senasa.gov.ar).	Experts within institutions should be identified to facilitate the contact. We suggest including their names in addition to the institutions in this section.	COSAVE,Paraguay ,Argentina ,Chile,Brazil
[318]	155	Technical	Department of Agriculture and Food Western Australia, Biosecurity & Research Division, Plant Biosecurity Branch, Entomology Unit, 3 Baron-Hay Court, South Perth, WA 6151, Australia (tel: +61 8 9368 3248, +61 8 9368 3965; fax: +61 8 9368 3223, +61 8 9474 2840; e-mail: aszito@agric.wa.gov.au). Main Inspectorate of Plant Health and Seed Service, Central Laboratory,	Experts of the institutions should be identified to facilitate the contact. We suggest to include the name of the contact point.	Mexico

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			Żwirki i Wigury 73, 87-100 Toruń, Poland (tel: +48 56 639 1110, +48 56 639 1115; fax: +48 56 639 1115; e-mail: wkarnkowski@piorin.gov.pl). Laboratorio de Plagas y Enfermedades de las Plantas. Servicio Nacional de Sanidad y Calidad Agroalimentaria (SENASA), Av. Ing. Huergo 1001, C1107AOK Buenos Aires, Argentina (tel: +54 11 4362 1177, extns 117, 118, 129 and 132; fax: +54 11 4362 1177, extn 171; e-mail: albabriano@senasa.gov.ar).		
[319]	155	Technical	Department of Agriculture and Food Western Australia, Biosecurity & Research Division, Plant Biosecurity Branch, Entomology Unit, 3 Baron-Hay Court, South Perth, WA 6151, Australia (tel: +61 8 9368 3248, +61 8 9368 3965; fax: +61 8 9368 3223, +61 8 9474 2840; e-mail: aszito@agric.wa.gov.au). Main Inspectorate of Plant Health and Seed Service, Central Laboratory, Żwirki i Wigury 73, 87-100 Toruń, Poland (tel: +48 56 639 11119, +48 56 639 1115; fax: +48 56 639 1115; e-mail: wkarnkowski@piorin.gov.pl). Laboratorio de Plagas y Enfermedades de las Plantas. Servicio Nacional de Sanidad y Calidad Agroalimentaria (SENASA), Av. Ing. Huergo 1001, C1107AOK Buenos Aires, Argentina (tel: +54 11 4362 1177, extns 117, 118, 129 and 132; fax: +54 11 4362 1177, extn 171; e-mail: albabriano@senasa.gov.ar,tmail.com).	Corrections: - the telephone number - the e-mail address + alternative e-mail address	European Union
[320]	155	Technical	Department of Agriculture and Food Western Australia, Biosecurity & Research Division, Plant Biosecurity Branch, Entomology Unit, 3 Baron-Hay Court, South Perth, WA 6151, Australia (tel: +61 8 9368 3248, +61 8 9368 3965; fax: +61 8 9368 3223, +61 8 9474 2840; e-mail: aszito@agric.wa.gov.au). Main Inspectorate of Plant Health and Seed Service, Central Laboratory, Żwirki i Wigury 73, 87-100 Toruń, Poland (tel: +48 56 639 1110, +48 56 639 1115; fax: +48 56 639 1115; e-mail: wkarnkowski@piorin.gov.pl). Laboratorio de Plagas y Enfermedades de las Plantas. Servicio Nacional de Sanidad y Calidad Agroalimentaria (SENASA), Av. Ing. Huergo 1001, C1107AOK Buenos Aires, Argentina (tel: +54 11 4362 1177, extns 117, 118, 129 and 132; fax: +54 11 4362 1177, extn 171; e-mail: albabriano@senasa.gov.ar).	Experts within institutions should be identified to facilitate the contact. We suggest including their names in addition to the institutions in this section.	OIRSA
[321]	158	Editorial	**8. References** **Banks, H.J.** 1994. *Illustrated identification keys for* Trogoderma granarium, T. glabrum, T. inclusum *and* T. variabile *(Coleoptera: Dermestidae) and other* Trogoderma *associated with stored products*. CSIRO Division of Entomology Technical Paper, No. 32. Commonwealth Scientific and Industrial Research Organisation, Canberra. 66 pp. **Barak, A.V.** 1989. Development of new trap to detect and monitor Khapra	Incorrect citiation, the material if from PaDIL not K Walker. consistency , see Annex 1 to ISPM 27 Thrips palmi	Australia

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			beetle (Coleoptera: Dermestidae). *Journal of Economic Entomology*, 82: 1470–1477. —— 1995. Chapter 25: Identification of common dermestids.*In*V. Krischik, G. Cuperus & D. Galliart, eds.*Stored product management,*pp. 187–196. Oklahoma State University, Cooperative Extension Service Circular No. E-912 (revised). **Barak, A.V., Burkholder, W.E. & Faustini, D.L.** 1990. Factors affecting the design of traps for stored-products insects. *Journal of the Kansas Entomological Society*, 63(4): 466–485. **Beal, R.S. Jr.** 1954. Biology and taxonomy of nearctic species of *Trogoderma. University of California Publications in Entomology,* 10(2): 35–102. —— 1956. Synopsis of the economic species of*Trogoderma*occurring in the United States with description of new species (Coleoptera: Dermestidae). *Annals of the Entomological Society of America*, 49: 559–566. —— 1960.*Descriptions, biology and notes on the identification of some*Trogoderma*larvae (Coleoptera, Dermestidae)*. Technical Bulletin, United States Department of Agriculture, No. 1226. 26 pp. —— 1982. A new stored product species of*Trogoderma*(Coleoptera: Dermestidae) from Bolivia.*The Coleopterists Bulletin*, 36(2): 211–215. —— 1991. Dermestidae (Bostrychoidea) (including Thorictidae, Thylodriidae).*In*F.W. Stehr, ed. *Immature Insects*, pp. 434–439. Duboque, Iowa, Michigan State University, Kendall/Hunt. Vol. 2, xvi+ 975 pp. **Berg, G.H.** 1999a. *Trogoderma granarium* Everts. *In* OIRSA, *Hojas de Datos sobre Plagas y Enfermedades de Productos Almacenados de Importancia Cuarentenaria y/o Económica para los Países Miembros del OIRSA*, pp. 120–145. El Salvador, OIRSA. Vol. 6. 164 pp. —— 1999b.*Trogoderma variabile*Ballion.*In*OIRSA,*Hojas de Datos sobre Plagas y Enfermedades de Productos Almacenados de Importancia Cuarentenaria y/o Económica para los Países Miembros del OIRSA*, pp. 146–161. El Salvador, OIRSA. Vol. 6. 164 pp. **CABI**. 2005. *Trogoderma granarium*. *In* Crop Protection Compendium, CD-ROM, Wallingford, UK, CAB International. ISBN 085199086x, http://cabicompendium.org/cpc/home.asp. **COSAVE**. 1999. *Trogoderma granarium* Everts, Ficha de procedimientos y métodos analiticos. COSAVE. 10 pp. (unpublished manuscript) **Delobel, A. & Tran, M.** 1993. *Les coléopterés des denrées alimentaires entreposeés dans les régions chaudes*. Faune tropicale XXXII. Paris, ORSTOM. 424 pp.		

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			Berg, G.H. 1999a. *Trogoderma granarium* Everts. *In* OIRSA, *Hojas de Datos sobre Plagas y Enfermedades de Productos Almacenados de Importancia Cuarentenaria y/o Económica para los Países Miembros del OIRSA*, pp. 120–145. El Salvador, OIRSA. Vol. 6. 164 pp. —— 1999b.*Trogoderma variabile*Ballion.*In*OIRSA,*Hojas de Datos sobre Plagas y Enfermedades de Productos Almacenados de Importancia Cuarentenaria y/o Económica para los Países Miembros del OIRSA*, pp. 146–161. El Salvador, OIRSA. Vol. 6. 164 pp. **CABI**. 2005. *Trogoderma granarium*. *In* Crop Protection Compendium, CD-ROM, Wallingford, UK, CAB International. ISBN 085199086x, http://cabicompendium.org/cpc/home.asp. **COSAVE**. 1999. *Trogoderma granarium* Everts, Ficha de procedimientos y métodos analiticos. COSAVE. 10 pp. (unpublished manuscript) **Delobel, A. & Tran, M.** 1993. *Les coléopterés des denrées alimentaires entreposeés dans les régions chaudes*. Faune tropicale XXXII. Paris, ORSTOM. 424 pp. **EPPO/CABI.** 1997. *Trogoderma granarium*. *In* I.M. Smith, D.G. McNamara, P.R. Scott, & M. Holderness, eds. *Quarantine pests for Europe*, 2nd edition. Wallingford, UK. CAB International. 1425 pp. **EPPO**. 2002. Diagnostic protocols for regulated pests, *Trogoderma granarium*. *Bulletin OEPP/EPPO Bulletin*, 32: 299–310. —— 2007. Plant Quarantine Data Retrieval System (PQR version 4.6). Paris, EPPO. **Green, M.** 1979. The identification of *Trogoderma variable* Ballion, *T. inclusum* and *T. granarium* Everts (Coleoptera, Dermestidae), using characters provided by their genitalia. *Entomologists Gazette*, 30: 199–204. **Haines, C.P.** (ed.) 1991. *Insects and arachnids of tropical stored products: their biology and identification (a training manual)*. Chatham Maritime, UK, Natural Resources Institute. 246 pp. **Háva, J.** 2003. *World catalogue of the Dermestidae (Coleoptera)*. Studie a zprávy Okresního muzea Praha-Východ, 1:196 pp. —— 2004. World keys to the genera and subgenera of Dermestidae (Coleoptera) with descriptions, nomenclature and distributional records.*Acta Musei Nationalis Pragae, Series B, Natural History*, 60(3–4): 149–164. **Hinton, H.E.** 1945. *A monograph of the beetles associated with stored products*, Vol. 1. London, British Museum (Natural History). 443 pp. **Kingsolver, J.M.** 1991. Dermestid beetles (Dermestidae, Coleoptera). *In* J.R. Gorham, ed. *Insect and mite pests in food. An illustrated key*, pp. 113–		

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			Importancia Cuarentenaria y/o Económica para los Países Miembros del OIRSA, pp. 120–145. El Salvador, OIRSA. Vol. 6. 164 pp. —— 1999b.*Trogoderma variabile*Ballion.*In*OIRSA,*Hojas de Datos sobre Plagas y Enfermedades de Productos Almacenados de Importancia Cuarentenaria y/o Económica para los Países Miembros del OIRSA*, pp. 146–161. El Salvador, OIRSA. Vol. 6. 164 pp. **CABI**. 2005. *Trogoderma granarium*. *In* Crop Protection Compendium, CD-ROM, Wallingford, UK, CAB International. ISBN 085199086x, http://cabicompendium.org/cpc/home.asp. **COSAVE**. 1999. *Trogoderma granarium* Everts, Ficha de procedimientos y métodos analiticos. COSAVE. 10 pp. (unpublished manuscript) **Delobel, A. & Tran, M.** 1993. *Les coléopterés des denrées alimentaires entreposeés dans les régions chaudes*. Faune tropicale XXXII. Paris, ORSTOM. 424 pp. **EPPO/CABI.** 1997. *Trogoderma granarium*. *In* I.M. Smith, D.G. McNamara, P.R. Scott, & M. Holderness, eds. *Quarantine pests for Europe*, 2nd edition. Wallingford, UK. CAB International. 1425 pp. **EPPO**. 2002. Diagnostic protocols for regulated pests, *Trogoderma granarium*. *Bulletin OEPP/EPPO Bulletin*, 32: 299–310. —— 2007. Plant Quarantine Data Retrieval System (PQR version 4.6). Paris, EPPO. **Green, M.** 1979. The identification of *Trogoderma variable* Ballion, *T. inclusum* and *T. granarium* Everts (Coleoptera, Dermestidae), using characters provided by their genitalia. *Entomologists Gazette*, 30: 199–204. **Haines, C.P.** (ed.) 1991. *Insects and arachnids of tropical stored products: their biology and identification (a training manual)*. Chatham Maritime, UK, Natural Resources Institute. 246 pp. **Háva, J.** 2003. *World catalogue of the Dermestidae (Coleoptera)*. Studie a zprávy Okresního muzea Praha-Východ, 1:196 pp. —— 2004. World keys to the genera and subgenera of Dermestidae (Coleoptera) with descriptions, nomenclature and distributional records.*Acta Musei Nationalis Pragae, Series B, Natural History*, 60(3–4): 149–164. **Hinton, H.E.** 1945. *A monograph of the beetles associated with stored products*, Vol. 1. London, British Museum (Natural History). 443 pp. **Kingsolver, J.M.** 1991. Dermestid beetles (Dermestidae, Coleoptera). *In* J.R. Gorham, ed. *Insect and mite pests in food. An illustrated key*, pp. 113–136. Washington, DC, USDA ARS and USDHHS, PHS, Agriculture Handbook No. 655, Vol. 1: 324 pp.		

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			—— 2002. Dermestidae. *In*R.H. Arnett Jr., M.C. Thomas, P.E. Skelley, & J.H. Frank, eds.*American beetles*, Vol. 2, pp. 228–232. Boca Raton, Florida, CRC Press. 861 pp. **Lawrence, J.F.** 1991. Order Coleoptera. *In* F.W. Stehr, ed. *Immature Insects,* 2 vols. 1137 pp. Dubuque, Iowa, Kendall/Hunt, Vol. 2. xvi + 975 pp. **Lawrence, J.F. & Britton, E.B.** 1991. Coleoptera (beetles). *In* C.S.I.R.O. ed. *Insects of Australia*, 2nd edition, Vol. 2, pp. 543–683. Carlton: Melbourne University Press. 2 vols. 1137 pp. ——. 1994. *Australian beetles.*Melbourne: Melbourne University Press. 192 pp. **Lawrence, J.F., Hastings, A.M., Dallwitz, M.J., Paine, T.A. & Zurcher E.J.** 1999a. Beetle larvae of the world: descriptions, illustrations, and information retrieval for families and subfamilies. CD-ROM, Version 1.1 for MS-Windows. Melbourne: CSIRO Publishing. —— 1999b. Beetles of the world: a key and information system for families and subfamilies. CD-ROM, Version 1.0 for MS-Windows. Melbourne: CSIRO Publishing. **Lindgren, D.L., Vincent, L.E. & Krohne, H.E.** 1955. The Khapra beetle, *Trogoderma granarium* Everts. *Hilgardia*, 24(1): 1–36. **Matveeva, V.I.** 2001. Идентификациа капрового жука, *Защита и карантин растиений*, 4: 31. **Mitsui, E.** 1967. [On the identification of the Khapra beetle.] *Reports of the Japan Food Research Institute, Tokyo*, 22: 8–13. (in Japanese) **Mordkovich, Ya.B. & Sokolov, E.A.** 1999. Определитель карантинных и других опасных вредителей сырья, продуктов запаса и посевного материала, Колос, Москва: 384. —— 2000. Выявление капрового жука в складских помещниях, *Защита и карантин растиений*, 12: 26–27. **Mound, L.** (ed.) 1989. *Common insect pests of stored food products. A guide to their identification*. London, British Museum (Natural History). 68 pp. **Mroczkowski, M.** 1968. Distribution of the Dermestidae (Coleoptera) of the world with a catalogue of all known species, *Annales Zoologici,* 26(3): 1–191. **Okumura, G.T. & Blanc, F.L.** 1955. Key to species of *Trogoderma* and to related genera of Dermestidae commonly encountered in stored grain in California. *In* California Legislature Joint Interim Committee on Agricultural and Livestock Problems*, Special Report on the Khapra Beetle,* Trogoderma granarium, pp. 87–89. Sacramento, California.		

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			Trogoderma. University of California Publications in Entomology, 10(2): 35–102. —— 1956. Synopsis of the economic species of *Trogoderma* occurring in the United States with description of new species (Coleoptera: Dermestidae). *Annals of the Entomological Society of America*, 49: 559–566. —— 1960.*Descriptions, biology and notes on the identification of some* Trogoderma*larvae (Coleoptera, Dermestidae)*. Technical Bulletin, United States Department of Agriculture, No. 1226. 26 pp. —— 1982. A new stored product species of *Trogoderma* (Coleoptera: Dermestidae) from Bolivia.*The Coleopterists Bulletin*, 36(2): 211–215. —— 1991. Dermestidae (Bostrychoidea) (including Thorictidae, Thylodriidae).*In*F.W. Stehr, ed. *Immature Insects*, pp. 434–439. Duboque, Iowa, Michigan State University, Kendall/Hunt. Vol. 2, xvi+ 975 pp. **Berg, G.H.** 1999a. *Trogoderma granarium* Everts. *In* OIRSA, *Hojas de Datos sobre Plagas y Enfermedades de Productos Almacenados de Importancia Cuarentenaria y/o Económica para los Países Miembros del OIRSA*, pp. 120–145. El Salvador, OIRSA. Vol. 6. 164 pp. —— 1999b.*Trogoderma variabile*Ballion. *In* OIRSA,*Hojas de Datos sobre Plagas y Enfermedades de Productos Almacenados de Importancia Cuarentenaria y/o Económica para los Países Miembros del OIRSA*, pp. 146–161. El Salvador, OIRSA. Vol. 6. 164 pp. **CABI**. 2005. *Trogoderma granarium*. *In* Crop Protection Compendium, CD-ROM, Wallingford, UK, CAB International. ISBN 085199086x, http://cabicompendium.org/cpc/home.asp. **COSAVE**. 1999. *Trogoderma granarium* Everts, Ficha de procedimientos y métodos analiticos. COSAVE. 10 pp. (unpublished manuscript) **Delobel, A. & Tran, M.** 1993. *Les coléopterés des denrées alimentaires entreposeés dans les régions chaudes*. Faune tropicale XXXII. Paris, ORSTOM. 424 pp. **EPPO/CABI.** 1997. *Trogoderma granarium*. *In* I.M. Smith, D.G. McNamara, P.R. Scott, & M. Holderness, eds. *Quarantine pests for Europe*, 2nd edition. Wallingford, UK. CAB International. 1425 pp. **EPPO**. 2002. Diagnostic protocols for regulated pests, *Trogoderma granarium*. *Bulletin OEPP/EPPO Bulletin*, 32: 299–310. —— 2007. Plant Quarantine Data Retrieval System (PQR version 4.6). Paris, EPPO. **Green, M.** 1979. The identification of *Trogoderma variable* Ballion, *T. inclusum* and *T. granarium* Everts (Coleoptera, Dermestidae), using characters provided by their genitalia. *Entomologists Gazette*, 30: 199–		

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			204. **Haines, C.P.** (ed.) 1991. *Insects and arachnids of tropical stored products: their biology and identification (a training manual)*. Chatham Maritime, UK, Natural Resources Institute. 246 pp. **Háva, J.** 2003. *World catalogue of the Dermestidae (Coleoptera)*. Studie a zprávy Okresního muzea Praha-Východ, 1:196 pp. —— 2004. World keys to the genera and subgenera of Dermestidae (Coleoptera) with descriptions, nomenclature and distributional records.*Acta Musei Nationalis Pragae, Series B, Natural History*, 60(3–4): 149–164. **Hinton, H.E.** 1945. *A monograph of the beetles associated with stored products*, Vol. 1. London, British Museum (Natural History). 443 pp. **Kingsolver, J.M.** 1991. Dermestid beetles (Dermestidae, Coleoptera). *In* J.R. Gorham, ed. *Insect and mite pests in food. An illustrated key*, pp. 113–136. Washington, DC, USDA ARS and USDHHS, PHS, Agriculture Handbook No. 655, Vol. 1: 324 pp. —— 2002. Dermestidae. *In*R.H. Arnett Jr., M.C. Thomas, P.E. Skelley, & J.H. Frank, eds.*American beetles*, Vol. 2, pp. 228–232. Boca Raton, Florida, CRC Press. 861 pp. **Lawrence, J.F.** 1991. Order Coleoptera. *In* F.W. Stehr, ed. *Immature Insects,* 2 vols. 1137 pp. Dubuque, Iowa, Kendall/Hunt, Vol. 2. xvi + 975 pp. **Lawrence, J.F. & Britton, E.B.** 1991. Coleoptera (beetles). *In* C.S.I.R.O. ed. *Insects of Australia*, 2nd edition, Vol. 2, pp. 543–683. Carlton: Melbourne University Press. 2 vols. 1137 pp. ——. 1994. *Australian beetles.*Melbourne: Melbourne University Press. 192 pp. **Lawrence, J.F., Hastings, A.M., Dallwitz, M.J., Paine, T.A. & Zurcher E.J.** 1999a. Beetle larvae of the world: descriptions, illustrations, and information retrieval for families and subfamilies. CD-ROM, Version 1.1 for MS-Windows. Melbourne: CSIRO Publishing. —— 1999b. Beetles of the world: a key and information system for families and subfamilies. CD-ROM, Version 1.0 for MS-Windows. Melbourne: CSIRO Publishing. **Lindgren, D.L., Vincent, L.E. & Krohne, H.E.** 1955. The Khapra beetle, *Trogoderma granarium* Everts. *Hilgardia*, 24(1): 1–36. **Matveeva, V.I.** 2001. *Идентификациа капрового́ жука, Защита и карантин растиений*, 4: 31. **Mitsui, E.** 1967. [On the identification of the Khapra beetle.] *Reports of the Japan Food Research Institute, Tokyo*, 22: 8–13. (in Japanese)		

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			Mordkovich, Ya.B. & Sokolov, E.A. 1999. Определитель карантинных и других опасных вредителей сырья, продуктов запаса и посевного материала, Колос, Москва: 384. —— 2000. Выявление капрового жука в складских помещниях, *Защита и карантин растиений*, 12: 26–27. **Mound, L.** (ed.) 1989. *Common insect pests of stored food products. A guide to their identification*. London, British Museum (Natural History). 68 pp. **Mroczkowski, M.** 1968. Distribution of the Dermestidae (Coleoptera) of the world with a catalogue of all known species, *Annales Zoologici,* 26(3): 1–191. **Okumura, G.T. & Blanc, F.L.** 1955. Key to species of *Trogoderma* and to related genera of Dermestidae commonly encountered in stored grain in California. *In* California Legislature Joint Interim Committee on Agricultural and Livestock Problems*, Special Report on the Khapra Beetle,* Trogoderma granarium, pp. 87–89*.* Sacramento, California. **Pasek, J.E.** 1998. *Khapra beetle (*Trogoderma granarium *Everts): pest-initiated pest risk assessment*. Raleigh, NC, USDA. 46 pp. **Peacock E.R.** 1993. *Adults and larvae of hide, larder and carpet beetles and their relatives (Coleoptera: Dermestidae) and of derontid beetles (Coleoptera: Derontidae)*. Handbooks for the identification of British insects No. 5, Royal Entomological Society, London. 144 pp. **Rees, B.E.** 1943. *Classification of the Dermestidae (larder, hide, and carpet beetles) based on larval characters, with a key to the North American genera*. USDA Miscellaneous Publication No. 511. 18 pp. **Rees, D.P.** 2004. *Insects of stored products*. Melbourne, Australia, CSIRO Publishing; London, UK, Manson Publishing. viii +181 pp. **Saplina, G.S.** 1984. Обследование складских помещений с помощью ловушек. *Защита растиений***,** 9: 38. **Strong, R.G. & Okumura, G.T.** 1966. *Trogoderma* species found in California, distribution, relative abundance and food habits. *Bulletin, Department of Agriculture, State of California,* 55: 23–30. **Varshalovich, A.A.** 1963. Капровый жук – опаснейший вредитель пищевых запасов. Сельхоиздат, Москва: 1–52. **Walker, K.**2008. Khapra beetle (Trogoderma granarium).Pest and Diseases Image Library (PaDIL), available online:http://www.padil.gov.au/viewPestDiagnosticImages.aspx?id=124, accessed 10 July 2008.		
[326]	158	Editorial	**8. References** **Banks, H.J.** 1994. *Illustrated identification keys for* Trogoderma granarium,	New references were added to be in accordance with the modified text.	Argentina

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			T. glabrum, T. inclusum *and* T. variabile *(Coleoptera: Dermestidae) and other* Trogoderma *associated with stored products*. CSIRO Division of Entomology Technical Paper, No. 32. Commonwealth Scientific and Industrial Research Organisation, Canberra. 66 pp. **Barak, A.V.** 1989. Development of new trap to detect and monitor Khapra beetle (Coleoptera: Dermestidae). *Journal of Economic Entomology*, 82: 1470–1477. —— 1995. Chapter 25: Identification of common dermestids.*In*V. Krischik, G. Cuperus & D. Galliart, eds.*Stored product management,*pp. 187–196. Oklahoma State University, Cooperative Extension Service Circular No. E-912 (revised). **Barak, A.V., Burkholder, W.E. & Faustini, D.L.** 1990. Factors affecting the design of traps for stored-products insects. *Journal of the Kansas Entomological Society*, 63(4): 466–485. **Beal, R.S. Jr.** 1954. Biology and taxonomy of nearctic species of *Trogoderma. University of California Publications in Entomology,* 10(2): 35–102. —— 1956. Synopsis of the economic species of*Trogoderma*occurring in the United States with description of new species (Coleoptera: Dermestidae). *Annals of the Entomological Society of America*, 49: 559–566. —— 1960.*Descriptions, biology and notes on the identification of some*Trogoderma*larvae (Coleoptera, Dermestidae)*. Technical Bulletin, United States Department of Agriculture, No. 1226. 26 pp. —— 1982. A new stored product species of*Trogoderma*(Coleoptera: Dermestidae) from Bolivia.*The Coleopterists Bulletin*, 36(2): 211–215. —— 1991. Dermestidae (Bostrychoidea) (including Thorictidae, Thylodriidae).*In*F.W. Stehr, ed. *Immature Insects*, pp. 434–439. Duboque, Iowa, Michigan State University, Kendall/Hunt. Vol. 2, xvi+ 975 pp. **Berg, G.H.** 1999a. *Trogoderma granarium* Everts. *In* OIRSA, *Hojas de Datos sobre Plagas y Enfermedades de Productos Almacenados de Importancia Cuarentenaria y/o Económica para los Países Miembros del OIRSA*, pp. 120–145. El Salvador, OIRSA. Vol. 6. 164 pp. —— 1999b.*Trogoderma variabile*Ballion.*In*OIRSA,*Hojas de Datos sobre Plagas y Enfermedades de Productos Almacenados de Importancia Cuarentenaria y/o Económica para los Países Miembros del OIRSA*, pp. 146–161. El Salvador, OIRSA. Vol. 6. 164 pp. **CABI**. 2005. *Trogoderma granarium*. *In* Crop Protection Compendium, CD-ROM, Wallingford, UK, CAB International. ISBN 085199086x, http://cabicompendium.org/cpc/home.asp. **COSAVE**. 1999. *Trogoderma granarium* Everts, Ficha de procedimientos y		

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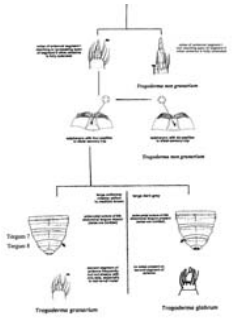
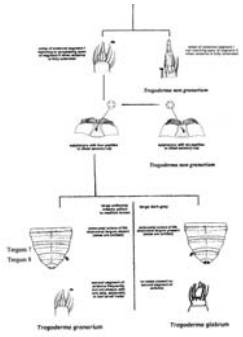
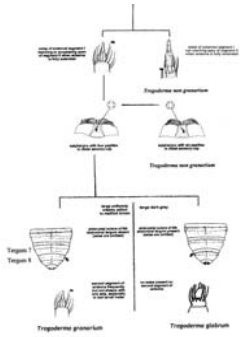
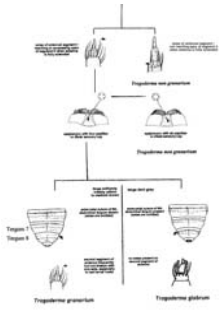
			carpet beetles) based on larval characters, with a key to the North American genera. USDA Miscellaneous Publication No. 511. 18 pp. **Rees, D.P.** 2004. *Insects of stored products*. Melbourne, Australia, CSIRO Publishing; London, UK, Manson Publishing. viii +181 pp. **Saplina, G.S.** 1984. Обследование складских помещений с помощью ловушек. *Защита растиений*, 9: 38. **Strong, R.G. & Okumura, G.T.** 1966. *Trogoderma* species found in California, distribution, relative abundance and food habits. *Bulletin, Department of Agriculture, State of California,* 55: 23–30. **Varshalovich, A.A.** 1963. Капровый жук – опаснейший вредитель пищевых запасов. Сельхоиздат, Москва: 1–52. **Walker, K.** 2008. Khapra beetle (Trogoderma granarium). Pest and Diseases Image Library (PaDIL), ~~, a~~ Available online from ~~:~~http://www.padil.gov.au/viewPestDiagnosticImages.aspx?id=124~~,~~ (accessed 10 July 2008).		
[329]	159	Substantive	**9. Figures**	Add photomicrographs. EPPO Secretariat to source and provide these	EPPO,Russian Federation ,Ukraine ,Morocco ,Uzbekistan
[330]	159	Substantive	**9. Figures**	Figures are very helpful in correct and rapid identification in both larval and adult stages. As there are many characters difficult to use in the identification without their figures, the following with morphological terms should be added; -habitus of adults:see Peacock, E.R., 1993. P83 -habitus of larvae: see Peacock, E.R. 1993. P121 -patterns of elytra: see Beal, R.S. 1954. P39 -position of hastisetae: see Peacock, E.R. 1993. P122 -antennae of larvae: see Peacock, E.R. 1993. P138 Also, please put scales to know the actual size of the figures.	Philippines ,Thailand ,Korea, Republic of ,Lao People's Democratic Republic,Japan ,India
[331]	159	Substantive	**9. Figures**	Add photomicrographs from: [http://panels.eppo.org/getfile2.php/cd0c6092d6a6874f379fe4827ed1db8b/4921f95baf824205e1b13f22d60357a1]	European Union
[332]	160	Substantive	 **Figure 1**:***Trogoderma granarium***:(A) adult, (B) young larva, (C) mature	A general review of photographs and pictures would be necessary. The images that appear in the protocol are not of good quality making the observation of the details cited in the text	Costa Rica ,Uruguay ,Nicaragua ,El

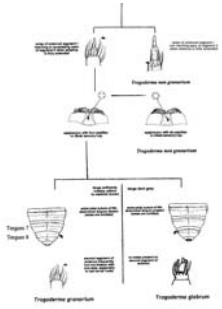
Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			larva ((A), (C), ICI Plant Protection Division; (B), Cornel Adler, BBA, Germany)	difficult.	Salvador
[333]	160	Substantive	 **Figure 1**:***Trogoderma granarium***:(A) adult, (B) young larva, (C) mature larva ((A), (C), ICI Plant Protection Division; (B), Cornel Adler, BBA, Germany)	Characteristics of male and female adult are different so we have to indicate if it is a male or a female.	Korea, Republic of ,Lao People's Democratic Republic,Japan ,India
[334]	160	Substantive	 **Figure 1**:***Trogoderma granarium***:(A) adult, (B) young larva, (C) mature larva ((A), (C), ICI Plant Protection Division; (B), Cornel Adler, BBA, Germany)	A general review of photographs and pictures would be necessary. The images that appear in the protocol are not of good quality, making it difficult the observation of the details cited in the text.	COSAVE,Paraguay ,Argentina ,Chile,Brazil
[335]	160	Substantive	 **Figure 1**:***Trogoderma granarium***:(A) adult, (B) young larva, (C) mature larva ((A), (C), ICI Plant Protection Division; (B), Cornel Adler, BBA, Germany)	A general review of photographs and pictures would be necessary. The images that appear in the protocol are not of good quality, making the observation of the details cited in the text difficult	OIRSA
[336]	160	Technical	 **Figure 1**:***Trogoderma granarium***:(A) adult, (B) young larva, (C) mature larva ((A), (C), ICI Plant Protection Division; (B), Cornel Adler, ~~BBA~~Julius Kuhn-Institut (JKI), Germany)	The name has been changed.	EPPO,Russian Federation ,Ukraine ,Morocco ,Uzbekistan
[337]	160	Technical	 **Figure 1**:***Trogoderma granarium***:(A) adult, (B) young larva, (C) mature larva ((A), (C), ICI Plant Protection Division; (B), Cornel Adler, BBA, Germany)	It's a pity that photographs arn't in colour as they assist in the identification of the pest – colour photos are available, not least on PaDIL. The precedent for use of colour photographs is in Annex 1 to ISPM 27 Thrips palmi.	Australia
[338]	160	Technical	 **Figure 1**:***Trogoderma granarium***:(A) adult, (B) young larva, (C) mature larva ((A), (C), ICI Plant Protection Division; (B), Cornel Adler, BBA,	Missing resolution and care the quality of the figures	Mexico

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			Germany)		
[339]	160	Technical	 ~~**Figure 1:*Trogoderma granarium*:(A) adult, (B) young larva, (C) mature larva ((A), (C), ICI Plant Protection Division; (B), Cornel Adler, BBA, Germany)~~ Figure 1:Trogoderma granarium:(A) adult, (B) young larva, (C) mature larva ((A), (C), ICI Plant Protection Division; (B), Cornel Adler, BBA, Germany)	1)suggest to use a scale for size specification. 2)As the character of wing spot is a little different, so it is better to indicate the male or female when provide the picture. 3) Two pictures: female and male adults are provided by Lab of Trogoderma in Shandong CIQ, China	China
[340]	160	Technical	 **Figure 1**:***Trogoderma granarium***:(A) adult, (B) young larva, (C) mature larva ((A), (C), ICI Plant Protection Division; (B), Cornel Adler, ~~BBA~~Julius Kuhn-Institut (JKI), Germany)	The name of the institute has been changed.	European Union
[341]	161	Editorial	Figure 2: ***Trogoderma*_spp**. elytral pattern (Beal, 1954)	Rewording to add clarity to the text because some words are all together in a same sentence	Mexico
[342]	162	Substantive	**Figure 3**: **Adult of *Trogoderma variabile*** (Berg, 1999b)	A general review of photographs and pictures would be necessary. The images that appear in the protocol are not of good quality making the observation of the details cited in the text difficult.	Costa Rica ,Uruguay ,Nicaragua ,El Salvador
[343]	162	Substantive	**Figure 3**: **Adult of *Trogoderma variabile*** (Berg, 1999b)	A general review of photographs and pictures would be necessary. The images that appear in the protocol are not of good quality, making it difficult the observation of the details cited in the text.	COSAVE,Paraguay ,Argentina ,Chile,Brazil
[344]	162	Substantive	**Figure 3**: **Adult of *Trogoderma variabile*** (Berg, 1999b)	A general review of photographs and pictures would be necessary. The images that appear in the protocol are not of good quality, making the observation of the details cited in the text difficult	OIRSA
[345]	165	Editorial	**Figure 5: Antennae of *Trogoderma granarium:*** (~~A~~) male antenna with normal number of segments; (~~B~~) female antenna with reduced number of segments; (~~C~~) female antenna with normal number of segments (Beal, 1956)	To be consistent with the figure.	Costa Rica ,Mexico ,Nicaragua ,El Salvador ,OIRSA

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
[346]	165	Editorial	**Figure 5: Antennae of *Trogoderma granarium:*** (~~A~~a) male antenna with normal number of segments; (~~B~~b) female antenna with reduced number of segments; (~~C~~c) female antenna with normal number of segments (Beal, 1956)	To be consistent with Figure 5	Uruguay
[347]	165	Editorial	**Figure 5: Antennae of *Trogoderma granarium:*** (~~A~~a) male antenna with normal number of segments; (~~B~~b) female antenna with reduced number of segments; (~~C~~c) female antenna with normal number of segments (Beal, 1956)	To be consistent with the figure.	COSAVE,Paraguay ,Argentina ,Chile,Brazil
[348]	173	Substantive	**Figure 10: Adult *Dermestes lardarius*;** copyright: Ministry of Agriculture Fisheries and Food, UK (Haines, 1991)	A general review of photographs and pictures would be necessary. The images that appear in the protocol are not of good quality making the observation of the details cited in the text difficult.	Costa Rica ,Uruguay ,Nicaragua ,El Salvador
[349]	173	Substantive	**Figure 10: Adult *Dermestes lardarius*;** copyright: Ministry of Agriculture Fisheries and Food, UK (Haines, 1991)	A general review of photographs and pictures would be necessary. The images that appear in the protocol are not of good quality, making it difficult the observation of the details cited in the text.	COSAVE,Paraguay ,Argentina ,Chile,Brazil
[350]	173	Substantive	**Figure 10: Adult *Dermestes lardarius*;** copyright: Ministry of Agriculture Fisheries and Food, UK (Haines, 1991)	A general review of photographs and pictures would be necessary. The images that appear in the protocol are not of good quality, making the observation of the details cited in the text difficult	OIRSA
[351]	174	Substantive	**Figure 11: Adult *Attagenus* sp.;** copyright: Natural History Museum, London, UK (Haines, 1991)	A general review of photographs and pictures would be necessary. The images that appear in the protocol are not of good quality making the observation of the details cited in the text difficult.	Costa Rica ,Uruguay ,Nicaragua ,El Salvador
[352]	174	Substantive	**Figure 11: Adult *Attagenus* sp.;** copyright: Natural History Museum, London, UK (Haines, 1991)	A general review of photographs and pictures would be necessary. The images that appear in the protocol are not of good quality, making it difficult the observation of the details cited in the text.	COSAVE,Paraguay ,Argentina ,Chile,Brazil
[353]	174	Substantive	**Figure 11: Adult *Attagenus* sp.;** copyright: Natural History Museum, London, UK (Haines, 1991)	A general review of photographs and pictures would be necessary. The images that appear in the protocol are not of good quality, making the observation of the details cited in the text difficult	OIRSA
[354]	175	Substantive	**Figure 12: Adult *Anthrenus verbasci*;** copyright: Natural History	A general review of photographs and pictures would be necessary. The images that appear in	Costa Rica ,Uruguay

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			Museum, London, UK (Haines, 1991)	the protocol are not of good quality making the observation of the details cited in the text difficult.	,Nicaragua ,El Salvador
[355]	175	Substantive	**Figure 12: Adult *Anthrenus verbasci*;** copyright: Natural History Museum, London, UK (Haines, 1991)	A general review of photographs and pictures would be necessary. The images that appear in the protocol are not of good quality, making it difficult the observation of the details cited in the text.	COSAVE,Paraguay ,Argentina ,Chile,Brazil
[356]	175	Substantive	**Figure 12: Adult *Anthrenus verbasci*;** copyright: Natural History Museum, London, UK (Haines, 1991)	A general review of photographs and pictures would be necessary. The images that appear in the protocol are not of good quality, making the observation of the details cited in the text difficult	OIRSA
[357]	178	Editorial	**Figure 15: Comparison of hastisetae morphology of various *Trogoderma* species:** (A), (B) *T. granarium*; (C), (D) *T. glabrum*; (E), (F) *T. variabile*; (G), (H) *T. inclusum*; copyright: Natural History Museum, London, UK (Peacock, 1993)	Figure sequence was originally C, D, A and B; and was changed to A, B, C and D to adjusted to the title of the figure.	COSAVE,Paraguay ,Argentina ,Chile,Brazil
[358]	178	Substantive	**Figure 15: Comparison of hastisetae morphology of various *Trogoderma* species:** (A), (B) *T. granarium*; (C), (D) *T. glabrum*; (E), (F) *T. variabile*; (G), (H) *T. inclusum*; copyright: Natural History Museum, London, UK (Peacock, 1993)	Figure 15 is missing in the OCS. However it should be reviewed because the order of the letters in the figure do not correspond with the explanatory text	OIRSA
[359]	178	Technical	**Figure 15: Comparison of hastisetae morphology of various *Trogoderma* species:** (A), (B) *T. granarium*; (C), (D) *T. glabrum*; (E), (F) *T. variabile*; (G), (H) *T. inclusum*; copyright: Natural History Museum, London, UK (Peacock, 1993)	Figure 15 is missing in the OCS. However, it should be reviewed because the order of the letters in the figure do not correspond with the explanatory text.	Costa Rica ,Uruguay ,Mexico ,Nicaragua ,El Salvador
[360]	179	Editorial	 **Figure 16: Pictorial key for distinguishing larvae of*Trogoderma granarium*from other *Trogoderma* spp.** (Berg (1999a); Kingsolver (1991))	Spaces missing	EPPO,Russian Federation ,European Union ,Ukraine ,Morocco ,Uzbekistan

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
[361]	179	Editorial	 **Figure 16: Pictorial key for distinguishing larvae of*Trogoderma granarium* from other*Trogoderma*spp.**(Berg (1999a); Kingsolver (1991))	Insert a space after 'granarium'.	United States of America
[362]	179	Editorial	 **Figure 16: Pictorial key for distinguishing larvae of *Trogoderma granarium* from other *Trogoderma* spp.**(Berg (1999a); Kingsolver (1991))	Rewording to add clarity to the text because some words are all together in a same sentence.	Mexico
[363]	179	Substantive	 **Figure 16: Pictorial key for distinguishing larvae**	A general review of photographs and pictures would be necessary. The images that appear in the protocol are not of good quality making the observation of the details cited in the text difficult.	Costa Rica ,Mexico ,Nicaragua ,El Salvador

Comment no.	Paragraph no.	Comment type	Comment	Explanation	Country
			of*Trogoderma granarium*from other*Trogoderma*spp.(Berg (1999a); Kingsolver (1991))		
[364]	179	Substantive	 **Figure 16: Pictorial key for distinguishing larvae of*Trogoderma granarium*from other*Trogoderma*spp.**(Berg (1999a); Kingsolver (1991))	A general review of photographs and pictures would be necessary. The images that appear in the protocol are not of good quality, making the observation of the details cited in the text difficult	OIRSA